

Improved CRISPR gene drive solves problems of old tech

March 4 2020



Credit: CC0 Public Domain

Gene drives use genetic engineering to create a desired mutation in a few individuals that then spreads via mating throughout a population in fewer than 10 generations.

In theory, such a mechanism could be used to prevent malarial mosquitoes from transmitting disease, or possibly to wipe out an invasive species by disabling its ability to reproduce.

Though scientists have had success proving the concept in the lab, they have found that [wild populations](#) invariably adapt and develop resistance to the scheme. And when gene drives work, they are all or nothing—without nuance—they spread to all individuals, which can be a drawback.

Now, a Cornell study, "A Toxin-Antidote CRISPR Gene Drive System for Regional Population Modification," published Feb. 27 in the journal *Nature Communications*, describes a new type of gene drive with the potential to delay resistance. The method could also be applied to a regional population, limiting its spread other populations where it could have undesired effects.

"Those are two things that this new drive that we developed here addresses to some extent," said Philipp Messer, an assistant professor of computational biology, and the paper's senior author. Jackson Champer, a postdoctoral researcher in Messer's lab, is the first author.

In a classic gene drive, called a homing drive, an offspring inherits one set of genes, or genome, from the mother and another from the father. If an offspring inherits a gene with a drive from one parent and not the

other, the drive copies itself into the genome from the parent without the drive.

"Now that individual has that drive in both of its genomes and it will pass it on to every offspring," Messer said.

The drives are engineered with CRISPR-Cas9 gene-editing technology, so when the drive copies itself into a new genome, the CRISPR machinery makes a cut into the chromosome without the drive, and pastes in the new code. But sometimes, cells will repair the incision and, in doing so, randomly delete DNA letters. When this happens, the CRISPR gene drive can no longer find a [genetic sequence](#) it recognizes in order to make the incision, which creates a resistance and stops the gene drive from spreading.

Natural genetic variation—another source of changes in DNA sequences—can also create resistance, since CRISPR gene drives must recognize short genetic sequences in order to make incisions.

"We were among the first labs to show that this is a tremendous problem," Messer said.

The paper describes a new gene drive, called TARE (Toxin-Antidote Recessive Embryo), which works by targeting a gene that is essential for an organism to function. At the same time, the organism can survive with only one intact copy of this essential gene. Instead of cutting and pasting DNA as homing drives do, the TARE drive simply cuts the other parent's gene, disabling it.

Meanwhile, the engineered TARE drive gene has a DNA sequence that has been recoded; the gene works but it won't be recognized or cut in future generations. If an offspring inherits two disabled genes, those individuals won't survive, thereby removing those copies from the

population. Meanwhile, as viable individuals mate, more and more surviving offspring will carry TARE drive [genes](#).

Just a few individuals with homing drives can spread a trait through an entire population. TARE drives, on the other hand, do not cut and paste a drive into a target gene; instead they destroy one of the target gene copies in the offspring. Because of this, the drive requires a higher frequency of engineered individuals in the population to spread. For this reason, TARE drives are less likely to transfer from one distinct population to another.

In lab experiments, when [fruit flies](#) with TARE [gene drives](#) were released in cages of wild-type fruit flies, all the flies in the cage had the TARE drive in just six generations.

The researchers pointed out that resistance can indeed evolve with a TARE drive in the wild, especially in very large populations, but they believe it will take longer and evolve at a much lower rate, Messer said.

More information: Jackson Champer et al. A toxin-antidote CRISPR gene drive system for regional population modification, *Nature Communications* (2020). [DOI: 10.1038/s41467-020-14960-3](https://doi.org/10.1038/s41467-020-14960-3)

Provided by Cornell University

Citation: Improved CRISPR gene drive solves problems of old tech (2020, March 4) retrieved 16 April 2024 from <https://phys.org/news/2020-03-crispr-gene-problems-tech.html>

This document is subject to copyright. Apart from any fair dealing for the purpose of private study or research, no part may be reproduced without the written permission. The content is provided for information purposes only.