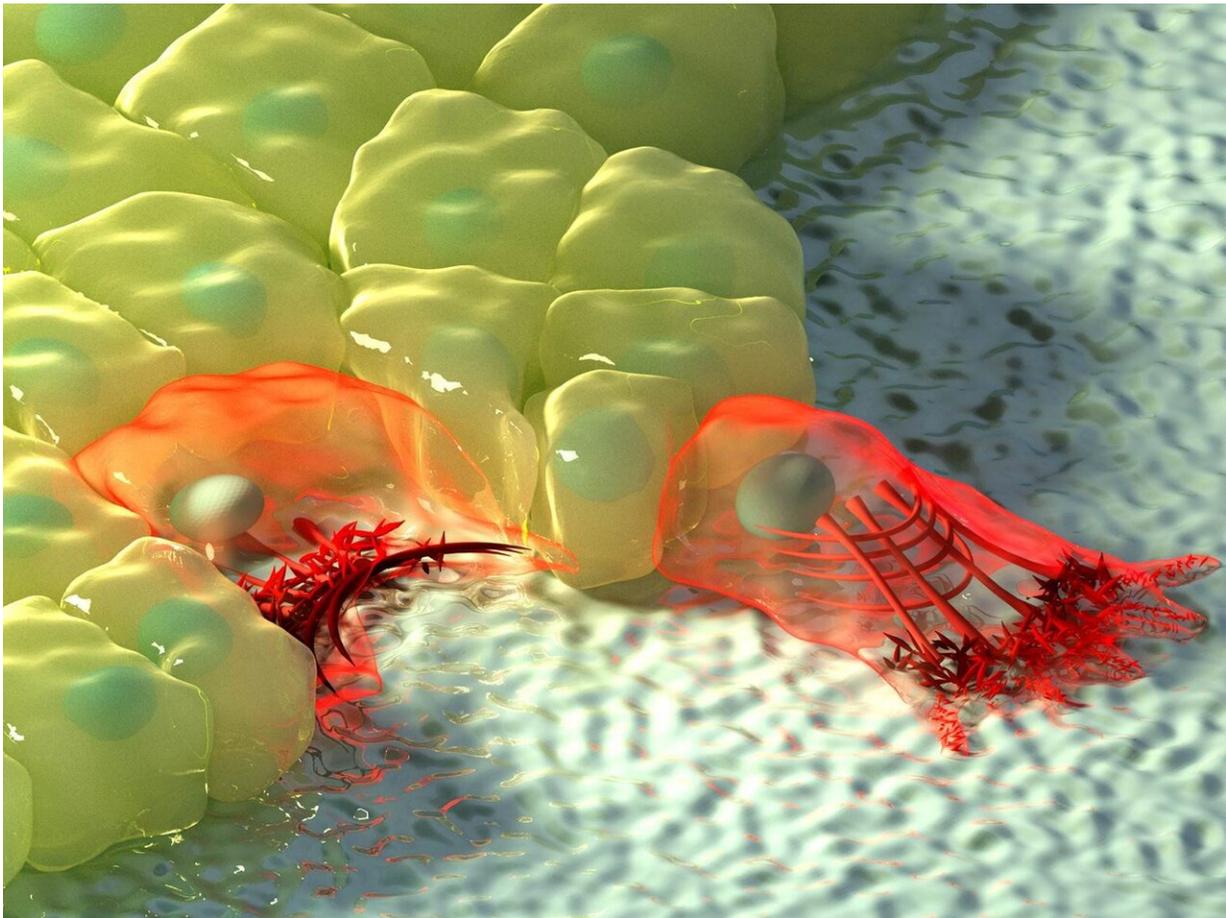


The mechanism of cellular migration mode switching

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The figure above shows an artistic illustration of cells at regions of negative (left) and positive curvature (right). At negative curvature, the actin flow moves towards the actin cable at the edge of the cell, depicted by the increase in colour intensity, and the contractile cable lifts the edge above the substrate. At positive curvature, lamellipodia extend from the cell to crawl along the substrate, and these protrusions are characterised by retrograde actin flow towards the inside of

the cell. Credit: Mechanobiology Institute, National University of Singapore

When faced with difficult terrain, off-road vehicles can switch from two- to four-wheel drive to keep moving forward. Similarly, cell migration can be driven either by protrusion-directed crawling, or by contractile pulling forces, but how the cell switches between these two methods remains a mystery. A collaborative study led by MBI doctoral student Tianchi Chen and Professor Benoit Ladoux of the Mechanobiology Institute (MBI) at the National University of Singapore, has uncovered that the direction in which actin filaments flow within the cell allows it to sense the physical curvature of its surroundings, and this directional flow is the key switch that determines which method of migration is selected. The work was published in the April 2019 issue of *Nature Physics*.

Curvature induced reorganisation of actin flow

Cell [migration](#) is an essential biological process that drives tissue and organ formation during embryo development, and also helps protect the body through immune response and wound healing mechanisms. The shape changes necessary for cell migration depends on dynamic organisation and force generation from the cell's internal actomyosin cytoskeleton, which is made up of structural [actin filaments](#) and contractile myosin motor proteins.

Reorganisation of these components enables two mechanisms of cell migration: the growth and extension of actin-based lamellipoidal protrusions that allow the cell to 'crawl' outwards, or the formation of large contractile actomyosin cables that can pull the cell forward while generating traction forces on the substrate. However, the process by which the cell chooses to form protrusions or contractile cables remains

unclear.

Depending on the topography of the physical environment, [cells](#) may encounter regions of positive or negative [curvature](#). Positive curvature refers to a convex shape that curves outward, much like a capital letter 'D', whereas an example of the opposite negative curvature can be seen in the letter 'C', which forms a concave shape that curves inward. By observing epithelial cell migration on flower-shaped substrate patterns that are used for wound closure assays, an international, multidisciplinary research team led by Prof Ladoux in collaboration with MBI Principal Investigators Associate Professor Boon Chuan Low and Associate Professor Yusuke Toyama, along with theoretical physicists and research peers in France, Chile, Israel and Spain found that at regions of positive curvature, cells developed lamellipodia for cell crawling, and at regions of negative curvature, strong actin cables assembled that followed the concave edge.

Further observation of actin dynamics in real-time using live cell microscopy revealed a difference in the direction of actin [flow](#) at the two curvatures. In positive curvature induced lamellipodia, the flow of actin was retrograde, i.e. away from the [cell membrane](#) in the opposite direction to the protrusion. Conversely, at negative curvatures the actin flow was anterograde, i.e. towards the cell membrane where the actin cable was located.

This anterograde flow drives the assembly of distinct, polarised actin structures at these regions of negative curvature. The actin cable at the concave edge is located at the front, followed by transverse actin fibres that are oriented in the same direction as the actin cable, and bringing up the rear of the network are radial actin fibres oriented at 90 degrees to the actin cable. Interestingly, the formation of these polarised actin structures in response to negative curvature was observed in single cells and cell sheets migrating on a 2-D surface, and in cells embedded in 3-D

fibrous networks. While these results confirmed the existence of two mechanisms of [cell migration](#), there was still an open question about how cells were able to sense the curvature of the environment.

Fortunately, the live cell imaging provided the researchers with a potential clue—the switch in the direction of actin flow depending on curvature could potentially be the 'sensor' for switching the mode of migration. This was confirmed by plating epithelial cells on a micro-patterned surface coated with an adhesive substrate, which was dotted with non-adherent circular patches. When the cell sheet encountered a non-adherent circle, lamellipodial protrusions crawl around the circle, pulling the cells forward on the adhesive substrate. This automatically created a concave shape across the non-adherent circle, which caused a rapid switch in the direction of actomyosin flow from retrograde to anterograde, resulting in assembly of an actin cable and a switch in migration mechanism. The contractile cable can lift up the edge of the cell over the non-adherent circle, enabling migration of the cell sheet to continue.

While cells do contain curvature sensors in their membrane, such as BAR proteins, these only allow the detection of curvature at the molecular level. Essentially, the size of these sensor proteins means that they can only sense curvature in their immediate vicinity at a scale of nanometres, not at the level of the whole cell. However, this study reveals how cells can sense larger cell-size curvature changes at a scale up to tens of micrometres, through dynamic adaptation of the actin flow.

This ability allows the cell to select an appropriate mechanism to continue migration, even when faced with diverse changes in the physical environment, much like how an off road vehicle can switch between two- and four-wheel drive depending on the terrain. With the new knowledge of this [actin](#) flow switch, scientists may be better able to understand how cells can continue migration during wound healing, or in

situations where migration can be damaging such as the metastatic progression of cancer.

More information: Tianchi Chen et al. Large-scale curvature sensing by directional actin flow drives cellular migration mode switching, *Nature Physics* (2019). [DOI: 10.1038/s41567-018-0383-6](https://doi.org/10.1038/s41567-018-0383-6)

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