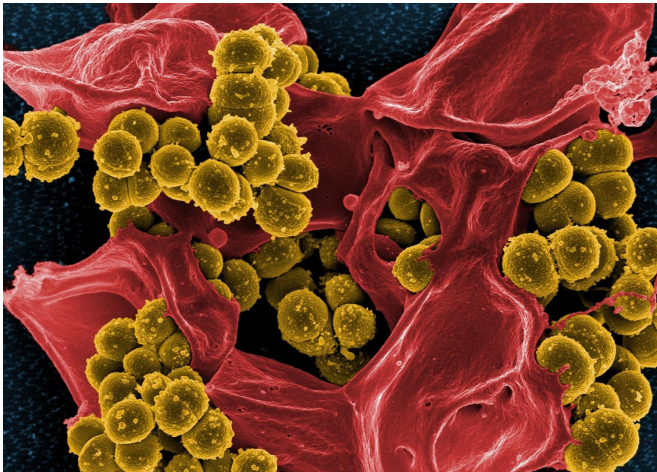


# DNA islands effective as 'anti-bacterial drones'

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Genomic "islands" that evolved from viruses can be converted into "drones" that disable *Staphylococcus aureus*, bacteria that are often resistant to antibiotics and pose a threat to safe hospital care, a new study finds.

Conducted by researchers from NYU School of Medicine and published online in the journal *Nature Biotechnology* on September 24, the study found that a certain type of bacterial DNA can be engineered to replace disease-causing genes with ones that kill or cripple bacteria.

The type of DNA featured in the study, a "pathogenicity island," evolved from viruses that stayed permanently in the bacteria they infected to become a part of their genetic system. The result is a hybrid entity that contains useful genes passed on by the bacteria when they reproduce, but that is also in some cases cut out of the bacterial superstructure, and packaged like a virus in a protein shell (capsid) that can inject its DNA into other bacterial cells.

This mix of evolutionary leaps has fashioned genomic [islands](#) as perfect drone-like vehicles to deliver genetic payloads throughout bacterial populations, say the study authors. When injected into mice with an otherwise lethal staph [infection](#), the research team's engineered *S. aureus* pathogenicity islands (SaPIs) killed the bacteria and rescued the treated animals.

"Given efficacy seen so far, and the safety record of related treatment attempts, we are getting set to test our drones against a Staph infection that interferes with milk production in cattle, and if successful there, in humans with Staph infections," says senior study author Richard P. Novick, MD, the Recanati Family Professor of Science at the Skirball Institute of Biomolecular Medicine within NYU School of Medicine, and a member of the National Academy of Sciences.

"It is an extraordinarily rewarding experience to spend a long career studying an infection, and then to arrive at a potentially new way to treat it," says Novick, who earned his MD at NYU School of Medicine and has been studying Staph ever since.

One-third of the human population are carriers of *S. aureus*, generally without symptoms. But those with weakened immune systems may develop life-threatening infections in the bloodstream, lungs (pneumonia), or organs (sepsis). *S. aureus* has long posed a threat in hospitals, where there are numerous surfaces for bacteria to colonize, and has more recently become a growing problem in the community (e.g. gyms, playgrounds, schools, military barracks, and doctor's offices).

In addition, antibiotics used against Staph are becoming less effective because an increasing number of strains have become resistant to these drugs. With many Staph infections now untreatable after decades of antibiotic over-use, the field urgently seeks new ways to counter infections.

## Borrowing Viral Tricks

Provided by NYU Langone Health

It was during research in the 1980s on a gene called TSST1—which causes a dangerous complication of Staph infections called toxic shock syndrome—that Novick and colleagues first discovered that a pathogenicity island carried TSST1 as part of its cargo. The team's subsequent drone design effort grew from the understanding that Staph bacteria depend on genomic islands to share useful genes. In this way, an entire population benefits when any one cell stumbles on a change that helps it to survive, and not just its offspring.

"The natural role of islands in such gene transfers set them up as a new treatment approach, if they could be made to contain genes that hampered bacteria instead of encouraging infection," says Hope Ross, Ph.D., a longtime member of the lab. In 2013, it was Ross that first saw this potential, and suggested that the lab focus on it.

As a first test, the team added to their island a CRISPR/Cas9 sequence, a genetic system that targets and cuts the DNA chain within a targeted gene, a lethal event in bacteria. Another drone studied by the team contains a gene for the enzyme lysostaphin, which directly kills bacteria by breaking down their cell walls. The team also studied a CRISPR approach with the potential to disable several, disease-causing bacterial [genes](#) without killing the bacteria, which promises to prevent infections from becoming drone resistant.

Novick's study also led to another important conclusion. "The drone system would not, like antibiotics, disrupt patients' microbiomes, the mix of [bacteria](#) in the gut, some species of which are essential to digestion and to general health," he says.

**More information:** Conversion of staphylococcal pathogenicity islands to CRISPR-carrying antibacterial agents that cure infections in mice, *Nature Biotechnology* (2018). [DOI: 10.1038/nbt.4203](#) , [www.nature.com/articles/nbt.4203](http://www.nature.com/articles/nbt.4203)

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