

# **A protein-RNA structure hints at how viruses commandeer human proteins**

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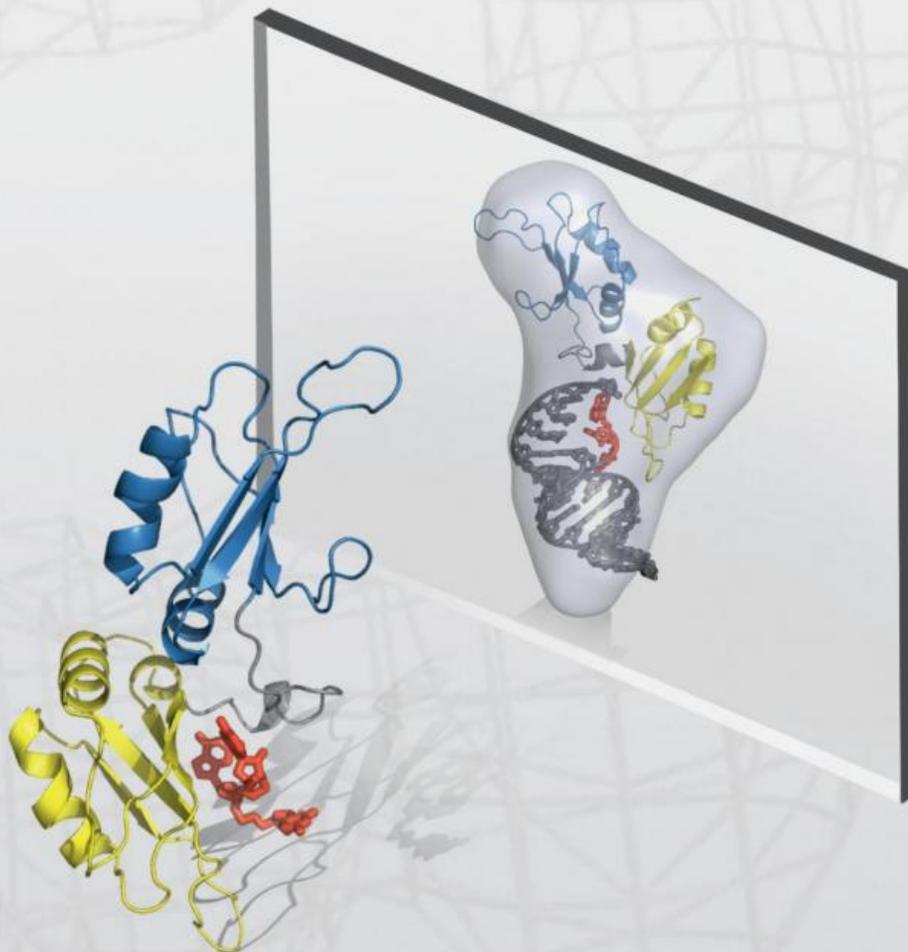
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Researchers at Case Western Reserve university and the University of Michigan combined three technologies to produce the image, showing how hnRNP A1 binds to the hairpin loop of RNA. The discovery may offer clues to how some viruses, including HIV, control expression of their genetic material. Credit: *Journal of Molecular Biology*

Researchers at Case Western Reserve University and the University of Michigan have produced the first image of an important human protein as it binds with ribonucleic acid (RNA), a discovery that could offer clues to how some viruses, including HIV, control expression of their genetic material.

That information could lead to new strategies to block viruses from replicating, thereby limiting or halting infection.

RNA is one of three macromolecules—along with DNA and proteins—essential to all forms of life. By understanding how hnRNP A1 binds to RNA, the scientists may find ways to jam up components of the replication machinery when the protein is coopted by disease.

The team of scientists reveals the mechanism used by the protein, hnRNP A1 to link to the section of RNA, called the 'hairpin loop.' Their research is published in a cover article in the *Journal of Molecular Biology*.

They found that hnRNP A1, a protein essential to cell function and virus replication, has a significantly different structure than its only previously known form: binding to DNA.

"We solved the three-dimensional structure of the protein bound to an RNA hairpin derived from the HIV virus," said Blanton Tolbert, a

chemistry professor at Case Western Reserve. "But because the hairpin loop is found in other viruses and throughout healthy cells, our findings may help explain how the protein connects to the other hairpin targets."

Tolbert began this research six years ago, frustrated that the only information available was the structure of the protein bound to a synthetic DNA, which isn't its natural target.

Proteins that bind hairpins sense both the structure and the sequence information presented in the loop. The structure of the DNA complex did not demonstrate the molecular recognition that must take place to bind RNA hairpins.

## **The process**

To discover the structure bound to RNA, the researchers combined three techniques: X-ray crystallography, [nuclear magnetic resonance spectroscopy](#) and small angle x-ray scattering. Each technique yielded a piece of the puzzle.

To bind to RNA, hnRNP A1 has two domains, RRM1 and RRM2, which are akin to hands. Scientists already knew both hands are needed to connect to RNA.

But the researchers found that, instead of each domain grabbing a section of the loop, only RRM1 makes contact with the RNA. RRM2 acts as support, helping organize RRM1 into the [structure](#) needed to conform to a certain section of the loop.

To confirm that the structures are key to binding, the researchers inserted mutations by changing amino acids on the surface of the domains.

Surprisingly, mutations on the far side of RRM1—the surface not in contact with the RNA but with the RRM2—caused decoupling at that site and substantially weakened the affinity for RNA.

Without the normal connection between the two domains, RRM1 fails to adopt the geometric shape that conforms to the RNA hairpin loop.

The researchers are further investigating how the protein transmits the effects of RRM2 to RRM1 and bind. They are also exploring the development of antagonistic agents that would disrupt the interaction of the [protein](#) with viruses.

**More information:** Christopher E. Morgan et al. The First Crystal Structure of the UP1 Domain of hnRNP A1 Bound to RNA Reveals a New Look for an Old RNA Binding Protein, *Journal of Molecular Biology* (2015). [DOI: 10.1016/j.jmb.2015.05.009](https://doi.org/10.1016/j.jmb.2015.05.009)

Provided by Case Western Reserve University

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