Chemists develop faster, more efficient protein labeling
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North Carolina State University researchers have created specially engineered mammalian cells to provide a new "chemical handle" which will enable researchers to label proteins of interest more efficiently, without disrupting the normal function of the proteins themselves or the cells in which they are found.

Protein labeling is used by researchers in a variety of fields to help them understand how these important molecules affect the normal functioning of cells. Currently, proteins are labeled for study simply by fusing them to other fluorescent proteins, which allows researchers to use microscopy to track their movements through a cell. This approach has several drawbacks, however, not least being that the fluorescent proteins are often large enough to affect the function of the protein of interest.

Dr. Alex Deiters, associate professor of chemistry, along with colleague Dr. Jason Chin of the Laboratory of Molecular Biology at the Medical Research Council in Cambridge, U.K., have developed a way to attach a fluorophore - a fluorescent molecule about 20 times smaller than the fluorescent proteins currently in use - to a protein that is expressed in a mammalian cell.

Deiters and Chin developed a special 21st amino acid that they added to cells that were specially engineered to incorporate this amino acid into the protein they wanted to study (there are normally only 20 amino acids). This 21st amino acid has a "chemical handle" that only reacts with a specifically designed fluorophore, but not any cellular components. According to Deiters, "The reaction between the modified protein and the fluorophore is extremely fast, high yielding, and generates a stable link between both reaction partners. This novel methodology enables future cell biological studies that were previously not possible."

The research appears in the Feb. 5 issue of Nature Chemistry.

"We found that our approach gave us a higher yield of labeled proteins and that the binding reaction was 50 times faster than with current methods," Deiters says. "Additionally, it took less reagent to complete the reaction, so overall we have a faster, more efficient method for protein labeling, and less chance of interfering with the normal function of the proteins and cells being studied."

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