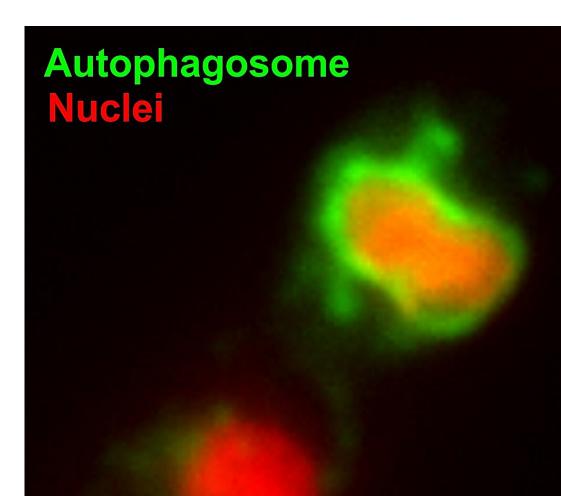


The world's most powerful anti-fungal chemistries cause fungal pathogens to selfdestruct

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2 micrometer

An autophagosome (green) in the process of "eating" a nucleus (red) in a azoletreated cell of Z. tritici. Image provided by co-author Dr. Martin Schuster. Credit: Dr. Martin Schuster



Scientists have discovered that the most widely-used class of antifungals in the world causes pathogens to self-destruct. The University of Exeterled research could help improve ways to protect food security and human lives.

Fungal diseases account for the loss of up to a quarter of the world's crops. They also pose a risk to humans and can be fatal for those with weakened immune systems.

Our strongest weapons against fungal plant diseases are azole fungicides. These chemical products account for up to a quarter of the world agricultural fungicide market, worth more than \$3.8 billion per year. Antifungal azoles are also widely used as a treatment against pathogenic fungi which can be fatal to humans, which adds to their importance in our attempt to control fungal disease.

Azoles target enzymes in the pathogen cell that produce cholesterol-like molecules, named ergosterol. Ergosterol is an important component of cellular bio-membranes. Azoles deplete ergosterol, which results in killing of the pathogen cell. However, despite the importance of azoles, scientists know little about the actual cause of pathogen death.

In a new study <u>published</u> in *Nature Communications*, University of Exeter scientists have uncovered the cellular mechanism by which azoles kill pathogenic fungi. The paper is titled "Azoles activate type I and type II programmed cell death pathways in crop <u>pathogenic fungi</u>." Coauthors are Dr. Martin Schuster and Dr. Sreedhar Kilaru at the University of Exeter.

The team of researchers, led by Professor Gero Steinberg, combined livecell imaging approaches and molecular genetics to understand why the inhibition of ergosterol synthesis results in cell death in the crop pathogenic fungus Zymoseptoria tritic (Z. tritici). This fungus causes



septoria leaf blotch in wheat, a serious disease in temperate climates, estimated to cause more than \$300 million per year in costs in the UK alone due to harvest loss and fungicide spraying.

The Exeter team observed living Z. tritici cells, treated them with agricultural azoles and analyzed the cellular response. They showed that the previously-accepted idea that azoles kill the pathogen cell by causing perforation of the outer cell membrane does not apply. Instead, they found that azole-induced reduction of ergosterol increases the activity of cellular mitochondria, the "powerhouse" of the cell, required to produce the cellular fuel that drives all <u>metabolic processes</u> in the pathogen cell.

While producing more "fuel" is not harmful in itself, the process leads to the formation of more toxic by-products. These by-products initiate a "suicide" program in the pathogen cell, named apoptosis. In addition, reduced ergosterol levels also trigger a second "self-destruct" pathway, which causes the cell to eat its own nuclei and other vital organelles—a process known as macroautophagy. The authors show that both <u>cell</u> <u>death</u> pathways underpin the lethal activity of azoles. They conclude that azoles drive the fungal pathogen into "suicide" by initiating self-destruction.

The authors found the same mechanism azoles killing pathogen cells in the rice-blast fungus Magnaporthe oryzae. The disease caused by this fungus kills up to 30% of rice, an essential food crop for more than 3.5 billion people across the world. The team also tested other clinically relevant anti-fungal drugs that target ergosterol biosynthesis, including terbinafine, tolfonate and fluconazole. All initiated the same responses in the pathogen cell, suggesting that cell suicide is a general consequence of ergosterol biosynthesis inhibitors.

Lead author Professor Gero Steinberg, who holds a Chair in Cell Biology and is Director of the Bioimaging Centre at the University of



Exeter, said, "Our findings rewrite common understanding of how azoles kill fungal pathogens. We show that azoles trigger cellular 'suicide' programs, which result in the pathogen self-destructing. This cellular reaction occurs after two days of treatment, suggesting that cells reach a 'point of no return' after some time of exposure to azoles. Unfortunately, this gives the pathogen time to develop resistance against azoles, which explains why azole resistance is advancing in fungal pathogens, meaning they are more likely to fail to kill the disease in crops and humans.

"Our work sheds light on the activity of our most widely used chemical control agents in crop and human pathogens across the world. We hope that our results prove to be useful to optimize control strategies that could save lives and secure food security for the future."

More information: Azoles activate type I and type II programmed cell death pathways in crop pathogenic fungi, *Nature Communications* (2024). <u>www.nature.com/articles/s41467-024-48157-9</u>

Provided by University of Exeter

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