

In vivo production of CAR-T cells using virusmimetic fusogenic nanovesicles

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The virus-mimetic fusogenic nanovesicle (FuNV) can efficiently fuse with T cell in vivo, thereby delivering the loaded anti-CD19 (α CD19) chimeric antigen receptor (CAR) onto T cells to produce α CD19 CAR-T cells. These in vivo engineered α CD19 CAR-T cells show effective inhibition of B-cell lymphoma. Credit: Science China Press



Chimeric antigen receptors (CARs) are synthesized membrane proteins that enable lymphocytes to recognize and respond to the specific antigens of target cells. Despite the impressive efficacy of CAR-T cell therapy in treating B-cell lymphoma or leukemia, the expensive and complex manufacturing process has hindered its widespread clinical application.

Previous research has explored the use of nanoparticles for delivering <u>nucleic acids</u> to program circulating T cells in vivo, streamlining CAR-T cell generation and obviating the need for isolating T cells from patients. Meanwhile, inserting the CAR protein directly into the T cell membrane could present a straightforward method, circumventing complications such as cytokine release syndrome (CRS) and the tumorigenic risk associated with random viral gene insertion into the genome.

Led by Prof. Jun Wang and Prof. Cong-Fei Xu from the School of Biomedical Sciences and Engineering at the South China University of Technology, researchers have developed a promising strategy that involves the direct fusion of CAR molecules, pre-expressed on fusogenic nanovesicles (FuNVs), to T cells, thereby constructing CAR-T cells in vivo.

They engineered the T-cell fusogen by adding an anti-CD3 single-chain variable fragment to the reovirus or the measles virus fusogen. They demonstrated that FuNVs derived from T-cell fusogen-expressing cells carried a substantial quantity of T-cell fusogen, which efficiently induced the fusion between NVs and T cells both in vitro and in vivo.

Subsequently, considering the clinical success of anti-CD19 (α CD19) CAR-T cells, the <u>engineered cells</u> expressing T-cell fusogen and α CD19 CAR protein were constructed to produce α CD19 CAR-carrying FuNVs (FuNV_{CAR}). CAR-T cell production was successfully achieved by delivering CAR protein onto T-cells via FuNV_{CAR} in vitro and in vivo.



Meanwhile, intravenous injection of $FuNV_{CAR}$ effectively inhibited B-cell lymphoma growth.

To further explore the potential toxicity of $FuNV_{CAR}$, blood counts and serum biochemical analyses were conducted at 2 days and 14 days, demonstrating comparability to the control group. Throughout the treatment with $FuNV_{CAR}$, no significant alterations in <u>body weight</u> were observed in mice.

Furthermore, in contrast to traditional CAR-T cell treatment, treatment with $FuNV_{CAR}$ did not induce an elevated release of inflammatory cytokines. This observed difference can be attributed to transient CAR-T cells produced by $FuNV_{CAR}$, which undergoes limited and temporary activation, mitigating the sustained release of inflammatory cytokines.

In summary, this study introduces a novel approach for in vivo CAR-T cell production through FuNV-mediated CAR protein delivery. It is essential to note, however, that this strategy may not be suitable for patients with compromised T cell function.

The findings are **<u>published</u>** in the journal *Science Bulletin*.

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