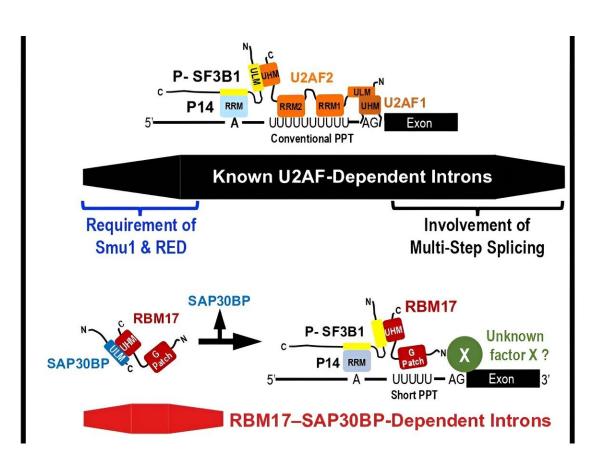


## Discovery of a subset of human short introns that are spliced out by a novel mechanism

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Human introns have extensively varying lengths. Previously, only U2AFdependent splicing was known. Researchers from Fujita Health University now show that splicing in a subset of human short introns is mediated by the intermediary RBM17–SAP30BP complex, instead of the U2AF heterodimer. Credit: Kazuhiro Fukumura and Akila Mayeda from Fujita Health University, Japan. *Cell Reports* (2023). DOI: 10.1016/j.celrep.2023.113534



The well-known essential pre-mRNA splicing factor U2AF heterodimer (U2AF2–U2AF1) has been identified as mediating early splicing reactions in all introns of different lengths. However, Dr. Kazuhiro Fukumura in the Akila Mayeda lab at Fujita Health University has discovered that a subset of short introns with truncated polypyrimidine tracts are spliced by the RBM17–SAP30BP complex instead of U2AF heterodimer. Dr. Fukumura's team has proposed a unique mechanism in which SAP30BP guides RBM17 to active early spliceosomes.

In humans, the length of pre-mRNA varies extensively (from 30 to 1,160,411 nucleotides by recent studies). The fundamental mechanism of splicing has been studied with model pre-mRNAs including 158- and 231-nt introns, for example, that are spliced very efficiently in vitro and in vivo.

Such an ideal pre-mRNA contains good splicing signal sequences, i.e., the 5' splice site, the branch-site (BS) sequence, and the polypyrimidine tract (PPT) followed by the 3' splice site that are recognized by U1 snRNP, U2 snRNP and U2AF2–U2AF1, respectively. Prof. Mayeda says, "Given the diverse lengths of human introns, it is likely that more than one mechanism exists. This is our motivation to initiate our study of splicing focused on human short introns."

Dr. Fukumura explains, "Our previous research on the splicing process on short intron revealed that the authentic splicing factor U2AF2 cannot bind to the truncated PPT and then RBM17 is replaced with U2AF to promote splicing. You know, this is reasonable because short introns are often too tight for the sufficient length of PPT. We published this finding in 2021.

"However, RBM17 cannot bind to the truncated PPT in vitro, so we did not know how the truncated PPT and the following 3' splice site are recognized by RRM17. Therefore, we hypothesized that another protein



factor is involved in RBM17-dependent splicing."

The Mayeda group eventually identified this protein cofactor behind the RBM17-dependent splicing, which is the SAP30BP. Their study was <u>published</u> in the journal *Cell Reports* on December 7, 2023.

Dr. Fukumura states, "It was critical to investigate previous references. From three papers, I was convinced that SAP30BP is the strongest candidate for the cofactor of RBM17." They showed that the existence of SAP30BP in human early splicing complex, the fruit fly SAP30BP, and RBM17 were detected in fly spliceosome formed on a short <u>intron</u>, and the binding between SAP30BP and RBM17 was indeed detected by yeast two-hybrid analysis.

"Nowadays, siRNA-mediated depletion of SAPBP in human cell line is the easy straightforward way to check the repression of RBM17-dependent splicing. And it was bingo!" says Dr. Fukumura.

The transcripts in SAP30BP-depleted human cells were analyzed by a next-generation sequencer (RNA-Seq analysis), and many RBM17- and SAP30BP-dependent introns were found. These introns were distributed in the shorter range and the truncated PPT was indeed a critical determinant of the RBM17/SAP30BP-dependency. Thus, RBM17 and SAP30BP are the general splicing factors.

Prof. Mayeda remarks, "It was a lucky coincidence that Prof. Michael Sattler, who is an expert in structural analyses, was keenly interested in our study, and we could start a productive collaboration."

Protein–protein interactions through UHM (U2AF-homology motif)–ULM(UHM-ligand motif) binding play essential roles in general splicing reactions. The Sattler lab found a hidden ULM sequence in SAP30BP, and demonstrated this was critical to interact with UHM in



RBM17 by NMR (<u>nuclear magnetic resonance</u>) and ITC (isothermal titration calorimetry) analyses.

However, the role of RBM17–SAP30BP interaction remained enigmatic. Since RBM17 has only one UHM, the RBM17–SAP30BP binding has to be released before the RBM17 interaction with SF3B1, one component of U2 snRNP, that is essential to promote splicing. So, what is the role of the RBM17–SAP30BP interaction?

Prof. Mayeda says, "Fukumura designed a smart binding assay using antiphospho-SF3B1 antibodies to address this curious question, and we could provide an elegant working model."

The researchers propose that the intermediary RBM17–SAP30BP complex prevents non-functional RBM17 binding to free unphosphorylated SF3B, which promotes functional RBM17 binding to active phosphorylated SF3B1 on pre-mRNA.

**More information:** Kazuhiro Fukumura et al, SAP30BP interacts with RBM17/SPF45 to promote splicing in a subset of human short introns, *Cell Reports* (2023). DOI: 10.1016/j.celrep.2023.113534

Provided by Fujita Health University

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