

First atomic-scale 'movie' of microtubules under construction, a key process for cell division

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3D maps of the microtubule nucleation process, starting from the initial nucleation stages (left), obtained after sorting the data from cryo-electron microscopes using image processing and neural networks. Credit: Marina Serna/CNIO



Cells in the human body are constantly dividing. With each division, the genetic information contained in the chromosomes is duplicated, and each daughter cell receives a complete copy of the genetic material. It is a sophisticated process, a clockwork mechanism that involves refined and fast changes within the cell. To make this possible, the cell relies on microtubules, tiny structures that are indeed tube-shaped. Understanding how they start forming is a long-standing question.

Now, for the first time, researchers at the Center for Genomic Regulation (CRG), the Spanish National Cancer Research Center (CNIO) and the Spanish National Research Council (IBMB-CSIC) have succeeded in making the equivalent of a film showing how human cells initiate the construction of their <u>microtubules</u>.

The findings, <u>published</u> in the journal *Science*, solve a problem brought up years ago and thus lay the groundwork for future breakthroughs in the treatment of diseases ranging from cancer to <u>neurodevelopmental</u> <u>disorders</u>.

Long ropes that pull chromosomes apart

Óscar Llorca, director of the Structural Biology Program at the CNIO and co-lead author of the study, describes what happens inside the cell when cell division begins: "The chromosomes, once they have duplicated genetic information, move to the center of the cell and the cell, in a very remarkable way, quickly sprouts from its two ends large tubes that hook the chromosomes and pull each of the copies towards the two poles of the cell. Only then is it possible to encapsulate a copy of all our genetic material in each daughter cell."

The structures that are launched "like long ropes that reach the chromosomes to divide them," explains Llorca, are the microtubules. "That's why we say that microtubules play a key role in cell division. We



need to understand very well the mechanisms that trigger the formation of these microtubules, at the right place and at the right time."

They are also 'cellular highways'

Microtubules are tubes with a length of thousandths of a millimeter and a diameter of nanometers [millionths of a millimeter]. In addition to being key to cell division, they act as 'highways' for moving cellular components between different areas of the cell. They are also structural elements that shape the cell itself, among other tasks. A good understanding of their formation has implications for multiple areas of biomedicine.

"Microtubules are critical components of cells. Here we capture the process in action inside human cells. Given the fundamental role of microtubules in cell biology, this could eventually lead to new therapeutic approaches for a wide range of disorders," explains ICREA Research Professor Thomas Surrey, CRG researcher and co-lead author of the paper.

Molecular ring triggers microtubule formation

The <u>high-resolution images</u> now obtained answer a question that has been hanging in the air for years: how microtubule formation begins in the early stages of cell division.

We now know that it all starts when a <u>complex structure</u> made up of several proteins, and called 'g TuRC' (pronounced 'gammaturc'), closes, forming a ring.

The shape of g TuRC, its three-dimensional structure, was discovered a few years ago, and it surprised researchers. It was expected that g TuRC



would be a closed ring acting as a base mold on top of which the microtubule is built; but g TuRC appeared as an open ring. Its dimensions and shape were incompatible with those of a microtubule mold.

The new CRG and CNIO work unveils the mechanism by which g TuRC closes into a ring and effectively becomes a perfect mold, capable of launching microtubule formation. The closure of g TuRC occurs when the first molecular piece of a microtubule gets attached to it.

"That's the trick the cell uses to close g TuRC," explains Llorca. "As soon as this first brick enters, a region of g TuRC is able to hook it and, like a loop, acts as a latch that pulls the ring closed and launches the process."

Visualizing this process required purifying g TuRC from <u>human cells</u> and reproducing the microtubule initiation process in the test tube. The samples were observed with cryo-electron microscopes and artificial intelligence was used for data analysis.

One million frames in a movie at atomic scale

One of the challenges has been to deal with the high speed of the microtubule construction process. The CRG group succeeded in slowing it down in the laboratory, and also stopping the growth of microtubules in order to better analyze the initial stages of the process.

"We had to find conditions that allowed us to image over a million microtubules in the process of nucleation before they grow too long and obscure the action of γ -TuRC. We were able to achieve this using the molecular toolbox of our lab and then freeze the microtubule stubs in place," explains Cláudia Brito, a postdoctoral researcher at the CRG and first author of the study.



The microtubules under construction were observed at the IBMB-CSIC's Electron Cryomicroscopy Platform, located at the Joint Electron Microscopy Center (JEMCA), inside the ALBA Synchrotron.

"They were frozen in a thin layer of ice, preserving the natural shape of the molecules involved," explains Pablo Guerra, head of this Platform.

That's how the best experimental conditions for observing microtubules in formation were determined. The best frozen samples were then sent to BREM (Basque Resource for Electron Microscopy) for imaging, and the resulting images were transferred to Marina Serna and Oscar Llorca at the CNIO for analysis and determination of the three-dimensional structures at atomic resolution.





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Artificial intelligence for assembly

In practice, having more than a million microtubules in different stages of growth is equivalent to having many frames of a movie in high resolution. You 'just' have to arrange them in the right order to see the movie in progress. That task fell to the CNIO team, which used artificial intelligence techniques to complete it

"Determining the three-dimensional structure of growing microtubules from microscope images has been extremely complex. We needed multiple digital image-processing tools," explains Marina Serna, CNIO researcher.

For Llorca, "the great challenge has been to analyze at high resolution the images of a dynamic process, where we were observing several stages at the same time. This has been possible thanks to the use of neural networks, which have allowed us to organize all this complexity."

The result are three-dimensional structures at atomic resolution that represent the different stages of how the construction of a microtubule begins, and how the γ -TuRC ring becomes the mold that launches the formation of microtubules.



Implications for health

Llorca explains, "This finding is relevant because we have addressed a very basic mechanism of <u>cell division</u>, whose process in humans we did not know."

This basic knowledge is useful for learning how to correct errors in the functioning of microtubules, which are associated with cancer, neurodevelopmental disorders and other conditions ranging from respiratory problems to heart disease.

"Some of the drugs used today to treat cancer prevent the formation or dynamics of microtubules," says Llorca. "However, these drugs affect microtubules indiscriminately, both in cancer cells and in healthy cells, leading to side effects. Knowing in detail how microtubules are formed may contribute to the development of more targeted treatments that affect microtubule formation and allow progress in the treatment of cancer and other diseases."





 γ -TuRC starting the nucleation process (left) vs. γ -TuRC closing (right). Once closed, thhe foundation is set and tubulins can be added to make the bridge as long as necessary. Credit: Marina Serna/CNIO

Next step: Understanding regulation

Thomas Surrey explains the next steps in understanding microtubules, which involve deciphering how microtubule formation is regulated: "The process of nucleation decides where the microtubules are in a cell and how many you have in the first place. It is likely that the conformational changes we observe are controlled by yet-to-be-found regulators in cells. Several candidates have been described in other studies, but their mechanism of action is unclear."



Further work "clarifying how regulators bind to γ -TuRC and how they affect the conformational changes during nucleation, may transform our understanding of how microtubules work, and eventually offer alternative sites that one might want to target to prevent cancer cells from going through the cell cycle," Surrey concludes.

More information: Cláudia Brito et al, Transition of human γ-tubulin ring complex into a closed conformation during microtubule nucleation, *Science* (2024). DOI: 10.1126/science.adk6160. www.science.org/doi/10.1126/science.adk6160

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