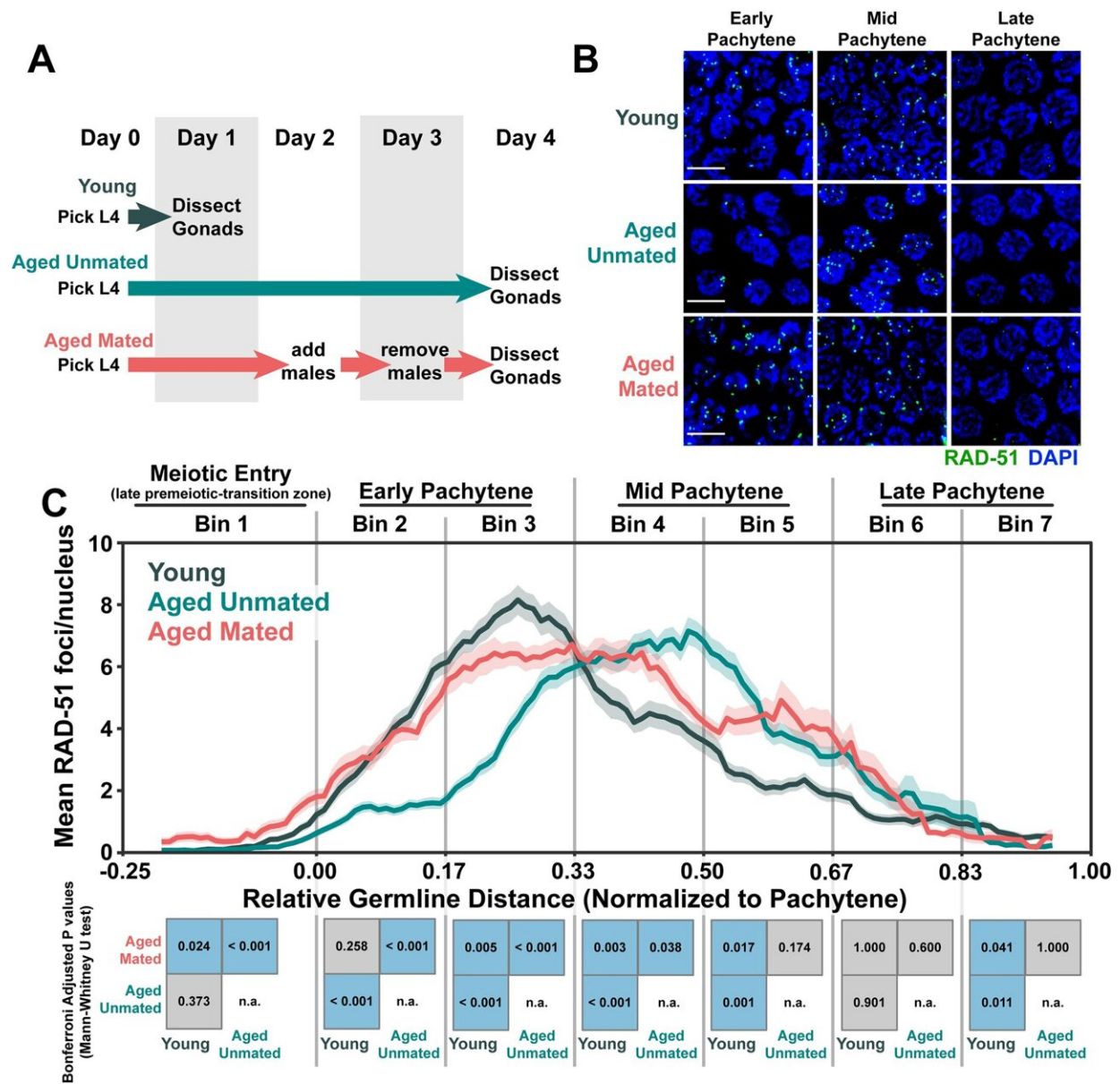


# Study finds DNA repair declines with age, limiting fertility

December 5 2022, by Laurel Hamers



DNA double strand break levels are altered during *C. elegans* germline aging. A) Schemes used to isolate young (1 day post-L4) and aged (4 days post L4) worms for experiments. Days count ~18–24 hour periods after hermaphrodites were isolated as L4 larvae and are separated by alternating grey shaded boxes. B) Representative images of RAD-51 foci in meiotic nuclei of young and aged germlines. Scale bars represent 5 $\mu$ m. C) RAD-51 foci per nucleus in oocytes. Line plots represent the mean RAD-51 foci per nucleus along the length of the germline in a sliding window encompassing 0.1 units of normalized germline distance with a step size of 0.01 germline distance units. Mean RAD-51 foci were calculated from nuclei analyzed in  $n = 9$  total germlines derived from  $\geq 3$  experimental replicates within each age group. Shaded areas around each line represent  $\pm$  SEM. Total nuclei analyzed (Bins 1/2/3/4/5/6/7) Young: 185/117/146/107/117/97/83; Aged Mated: 234/205/192/173/154/177/96; Aged Unmated: 268/147/161/175/191/152/110. Germline distances were normalized to the start (0) and end (1) of pachytene based on DAPI morphology (see Methods). For analysis, the germline was divided into 7 bins encompassing the transition zone (Bin 1), early pachytene (Bins 2–3), mid pachytene (Bins 4–5), and late pachytene (Bins 6–7). The germline positions at which each bin start and end are marked on the X axis as vertical grey lines. Heat maps below each bin display the p values of pairwise comparisons of RAD-51 foci per nucleus counts within that bin. P values were calculated using Mann-Whitney U tests with Bonferroni correction for multiple comparisons. Blue tiles indicate significant differences (adjusted p value = less than 0.05) and grey tiles indicate nonsignificant effects (adjusted p value = greater than 0.05). n.a. = not applicable. Numerical data associated with this figure is presented in S1 Data. Credit: *PLOS Genetics* (2022). DOI: 10.1371/journal.pgen.1010282

Even worms have a ticking fertility clock. Older worms are less efficient at repairing broken DNA strands while making egg cells—part of a process that's essential for fertility. A new study from University of Oregon (UO) biologists suggests one possible reason that reproduction slows with age.

Researchers from the lab of Diana Libuda report the findings in a paper published Nov. 7 in *PLOS Genetics*.

Each sperm or egg cell has only half the number of chromosomes found in a regular cell. During meiosis, the cell division process that forms sperm and eggs, the parent cells must evenly divide their DNA. The costs of error can be high, since incorrectly divided chromosomes are a major cause of birth defects.

To get it right, cells use a surprising strategy: They deliberately break their strands of DNA, and then repair them.

"It's one of the most amazing processes in biology," said Erik Toraason, a former graduate student in Libuda's lab who led the work. The repair process "physically locks the chromosomes together and provides an organization point" that ensures that the [chromosomes](#) are divided evenly.

Toraason wanted to understand how aging affects those processes. Eggs lose their viability relatively early in an organism's lifespan, but "the reason these cells are so susceptible to aging is not well understood," said Toraason, who is now a postdoctoral researcher at Princeton University. "But one of the factors that's considered to be important is genome repair, and the idea that that might decline, leaving [oocytes](#)—developing eggs—vulnerable to defects."

Worms proved to be an ideal testing ground for those questions. *C. elegans* worms are usually hermaphroditic, meaning each worm produces both eggs and sperm. But thanks to a genetic mutation that nixes sperm production, researchers can create egg-producing worms that are functionally female.

Toraason and research assistant Vikki Adler used that trick to separate

the effects of aging from the effects of running out of sperm, which happens normally as hermaphroditic worms age and also leads to a decline in fertility.

Running out of sperm seemed to affect the rate at which the hermaphroditic worms created breaks in their egg cell DNA, the researchers found. But the ability to repair DNA breaks declined with age even in the female worms, independent of sperm levels.

"We found that that forming breaks is altered when you run out of sperm, but break repair is only affected by the aging process, not sperm," Libuda said.

The team doesn't yet know why this process changes with age, but at least for worms, it could be related to a shift in resources. In worms that have used up all their sperm, unfertilized oocytes sometimes become a food source for offspring. Since the oocytes aren't being used for reproduction, the worms no longer need to spend resources breaking and repairing their DNA.

**More information:** Erik Toraason et al, Aging and sperm signals alter DNA break formation and repair in the *C. elegans* germline, *PLOS Genetics* (2022). [DOI: 10.1371/journal.pgen.1010282](https://doi.org/10.1371/journal.pgen.1010282)

Provided by University of Oregon

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