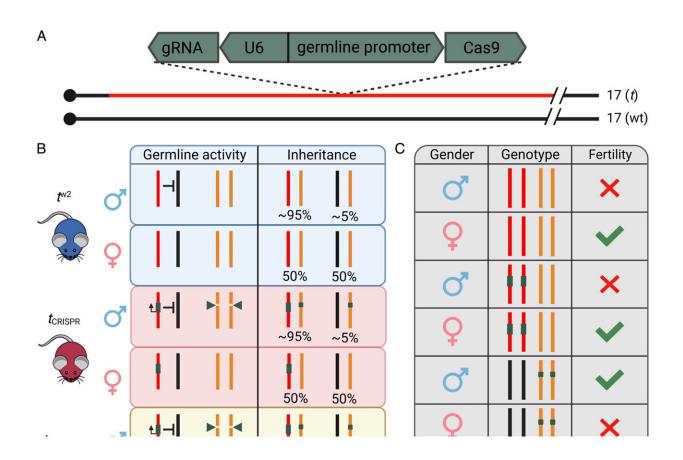


Gene drive technology to suppress invasive mice

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Overview of t haplotype modification strategies for population suppression. (A) The integration of a transgene within the t haplotype expressing Cas9 under the control of a male-specific promoter or a germline-specific promoter, coupled with a ubiquitously expressed gRNA targeting a haplosufficient female fertility gene. (B) Inheritance of t^{w^2} is biased in males but not females. In the t_{CRISPR} system, Cas9 is only active in males and, with a ubiquitously expressed gRNA, disrupts a haplosufficient female fertility gene in the germline. t_{CRISPR} males transmit the t_{CRISPR} transgene and a disrupted fertility gene to ~95% and 100%



of offspring, respectively. The t_{CRISPR(2)} strategy is identical except that the Cas9 is active in the male and female germline, thereby generating a more rapid increase in female infertility alleles. Prl* mice contain a sequence difference at the gRNA target site but maintain a functional protein, making these mice resistant against any further cleavage at that site. (C) Fertility of male and female mice carrying the various versions of chromosome 17 and Prl within the target chromosome. Credit: *Proceedings of the National Academy of Sciences* (2022). DOI: 10.1073/pnas.2213308119

Researchers at the University of Adelaide have released their first findings on the potential effectiveness of revolutionary gene drive technology to control invasive mice.

The team has developed a world-first proof of concept for the <u>technology</u>—called t-CRISPR—using <u>laboratory mice</u>.

Using sophisticated computer modeling performed by co-first author Dr. Aysegul Birand, the researchers also found about 250 gene-modified mice could eradicate an island population of 200,000 mice in around 20 years.

The results of the study have been published today in *Proceedings of the National Academy of Sciences*.

"This is the first time that a new genetic tool has been identified to suppress invasive mouse populations by inducing female infertility," said lead researcher Professor Paul Thomas from the University of Adelaide, and the South Australian Health and Medical Research Institute (SAHMRI).

"The t-CRISPR approach uses cutting-edge DNA editing technology to make alterations to a female fertility gene. Once the population is



saturated with the <u>genetic modification</u>, all the females that are generated will be infertile.

"We are also developing new versions of t-CRISPR technology that are designed to target specific pest populations to prevent unwanted spread of the gene drive."

Post-graduate student Luke Gierus, a co-first author of the research paper, said t-CRISPR was the first genetic biocontrol tool for invasive mammals.

"Up until now, this technology has been aimed at insects to try and limit the spread of malaria, which causes up to 500,000 deaths worldwide per year," Mr. Gierus said.

"The use of t-CRISPR technology provides a humane approach to controlling invasive mice without the release of toxins into the environment. We are also working on strategies to prevent failed eradication due to the emergence of gene drive resistance in the target population."

Professor Thomas said the research team had worked closely with Australia's National Science Agency CSIRO, the Center for Invasive Species Solutions, the Genetic Biocontrol for Invasive Rodents (GBIRd) consortium and the US Department of Agriculture to consider next steps towards safely implementing the new technology.

"Our broader project includes consideration of societal views and attitudes, and is integral to our ongoing research relating to this gene drive," Professor Thomas said.

CSIRO Group Leader for Environmental Mitigation and Resilience Dr. Owain Edwards added, "This particular prototype has been designed to



be highly specific for mice, but it is also evidence that gene drives can be developed against other invasive pest animals.

"As part of this research, we conduct the safety assessments for this technology to the highest standards. Because this is the first prototype for a vertebrate gene drive, interested stakeholders will include many from the <u>international community</u>."

The research was supported by the South Australian Government and NSW Government.

South Australian Deputy Premier, the Hon. Dr. Susan Close MP said, "These promising findings demonstrate how gene drive technology may be a game changer in managing the impacts of <u>mice</u> on our environment, community, and <u>agricultural sector</u>.

"This cutting-edge research also highlights the global leadership of the South Australian research sector, in finding solutions to social, environmental and economic challenges."

More information: Luke Gierus et al, Leveraging a natural murine meiotic drive to suppress invasive populations, *Proceedings of the National Academy of Sciences* (2022). DOI: 10.1073/pnas.2213308119

Provided by University of Adelaide

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