

Commonly used chemical fixation causes aggregation artifact

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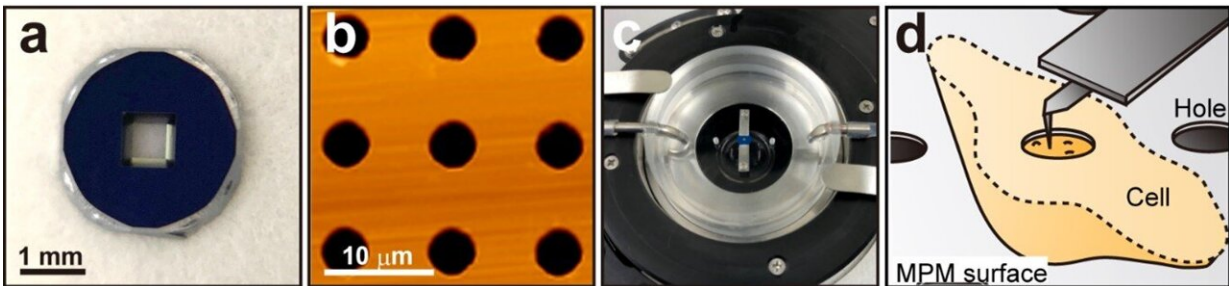


Figure 1. A new technique for observing living cell membranes for AFM using porous silicon nitride membrane (MPM). (a) The appearance of MPM. (b) The holes of MPM observed by AFM. (c) The MPM when set in the chamber for observation. (d) Schematic of AFM cell surface observation using MPM. Credit: Kanazawa University

Researchers at Kanazawa University report in *Communications Biology* that using common chemicals for fixing living cell samples for microscopy studies causes membrane proteins to aggregate.

For histological investigations of biological tissues, i.e. anatomical studies under the microscope, samples are usually fixated to prevent them from decaying. Fixation is typically done by immersing or perfusing the sample in a chemical—aldehydes and alcohols are common fixatives. It has been speculated that membrane proteins moving to some extent on a [cell membrane](#) can form aggregates during

[fixation](#). Yet, detailed [cell surface](#) studies with the nanometer-scale resolution are necessary for obtaining definitive insights into this potential issue. Now, Takehiko Ichikawa and colleagues from Kanazawa University have performed atomic force microscopy (AFM) studies of living mammalian cell surfaces. By comparing non-fixated and fixated samples, they found that fixation indeed leads to structural changes.

The researchers developed a method of using microporous silicon nitride membrane (MPM), used in [transmission electron microscopy](#) (Figure 1), for AFM imaging. Importantly, MPM can make the cell surface flat and prevent fluctuations by supporting the area outside the observation area. In AFM images of the surfaces of the cultured colon cancer cells on MPM, biomolecular structures on the cell membranes showed up as protrusions with a typical size of a few nanometers (Figure 2 living cell surface).

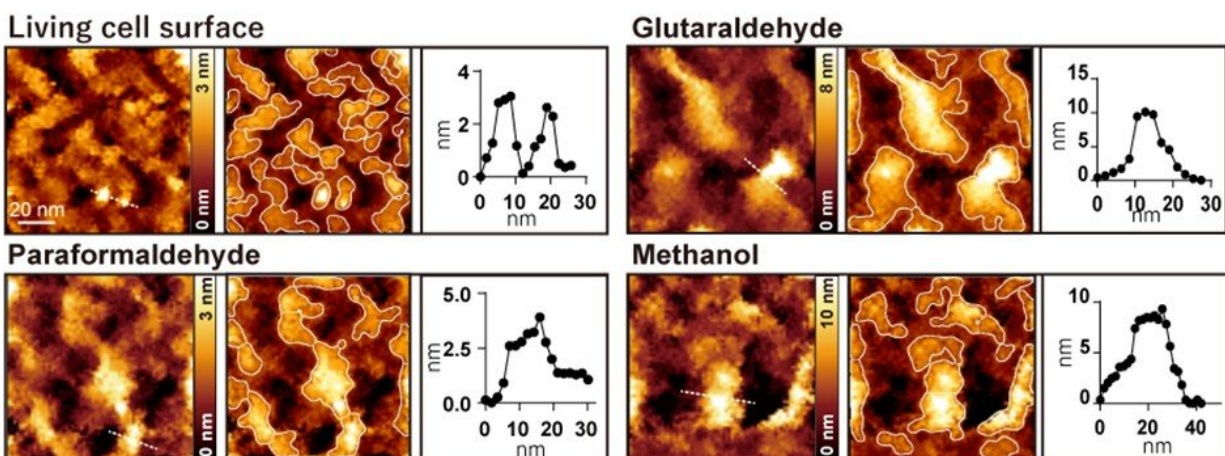


Figure 2. Observation of AFM cell surface before and after treatment with glutaraldehyde, paraformaldehyde, and methanol, respectively. The left image is an AFM image, the center image is an AFM image with the protrusions framed, and the right image is a height profile along the dotted line in the left image.

Credit: Kanazawa University

When the cells were treated with commonly used fixatives such as paraformaldehyde, glutaraldehyde, and methanol, a few nanometer structures disappeared, and only large protrusions with diameters ranging from 20 to 100 nanometers were observed (Figure 2). The researchers performed several fluorescence experiments and concluded that large protrusions observed after fixation were formed by the aggregation of [membrane proteins](#).

The study demonstrates that the observed aggregates are artifacts resulting from the fixation process. This should call for caution among the community of researchers working with chemical fixatives. Quoting Ichikawa and colleagues: "Researchers who observe nanoscale clusters also should be careful in interpreting their experimental results when using fixed cells. We recommend that researchers use living cells as much as possible to avoid the effect of fixation when investigating nanoscale clusters."

The general principle behind [atomic force microscopy](#) (AFM) is to make a very small tip scan the surface of a sample. During this horizontal (xy) scan, the tip, which is attached to a small cantilever, follows the sample's vertical (z) profile, inducing a force on the cantilever that can be measured. The magnitude of the force at the xy position can be related to the z value; the xyz data generated during a scan then result in a height map providing structural information about the investigated sample. AFM is not affected by the [diffraction limit](#) due to the use of light or electron beams and can observe the intact surface topography with high resolution.

More information: Takehiko Ichikawa et al, Chemical fixation creates nanoscale clusters on the cell surface by aggregating membrane proteins, *Communications Biology* (2022). [DOI: 10.1038/s42003-022-03437-2](#)

Provided by Kanazawa University

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