

Nanocapsule carrying a CRISPR-Cas9 editing tool used for noninvasive brain delivery and tumor cell targeting

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Fabrication, physical properties, and cellular function of Cas9/sgRNA nanocapsules. (A) In situ free-radical polymerization was used to synthesize disulfide-cross-linked nanocapsules containing Cas9/sgRNA and functionalized with angiopep-2 targeting ligand. (B) Size distribution of ANC_{SS}(Cas9/sgRNA) nanocapsules determined by dynamic light scattering. (C) TEM images of ANC_{ss}(Cas9/sgRNA) with or without GSH treatment. (D) Gel electrophoresis analysis of the ANC_{SS}(Cas9/sgPLK1) or free Cas9/sgPLK1 with or without RNase treatment (1 mg/ml, 30 min). (E) Flow cytometry of U87MG cells following 4-hour incubation with ANC_{SS}(Cas9/sgRNA) or controls. (F) Confocal laser scanning microscopy (CLSM) images of U87MG cells following 4-hour incubation with ANC_{SS}(Cas9/sgRNA) or controls. Cas9 was labeled with Alexa Fluor 647 (AF647; red); the cytoskeleton was stained with Alexa Fluor 488 (green), and the nuclei was stained with Hoechst 33342 (blue). For (E) and (F), the AF647-Cas9 concentration was 20 nM. Scale bars, 20 µm. (G) Luciferase gene editing efficiency in U87MG-Luc cells incubated with ANC_{SS}(Cas9/sgRNA) or controls for 72 hours. Data are presented as means \pm SD (n = 5; *P $_{SS}$ (Cas9/sgPLK1) or controls for 48 hours. (I) Schematic of gene editing in the nucleus. (J) Expression levels of PLK1 in U87MG cells after 72-hour incubation with ANC_{SS}(Cas9/sgPLK1) or controls. (K) Apoptosis assay of U87MG cells after 72-hour incubation with ANC_{ss}(Cas9/sgRNA) and other controls. For (G) to (K), the Cas9 concentration was 20 nM. bp, base pairs; PBS, phosphate-buffered saline. Credit: Science Advances (2022). DOI: 10.1126/sciadv.abm8011

An international team of researchers has developed a nanocapsule that is capable of crossing the blood brain barrier (BBB) to carry the CRISPR-Cas9 editing tool to treat a brain tumor. In their paper published in the journal *Science Advances*, the group describes how they created their capsule and how well it worked when tested in mice with a glioblastoma.

Glioblastomas are notoriously difficult to treat. The tumors appear in the brain and their growth damages tissue. Treatment options include surgical removal, direct injections of therapies meant to kill the cancer



cells or inserting CRISPR viruses into the bloodstream. Each of these options has a downside, from damaged brain tissue to ineffectiveness due to difficulties with therapies crossing the BBB. In this new effort, the researchers have tried a new approach, using a nanocapsule to carry the CRISPR-Cas9 editing tool to the brain tumor where it targets a gene that is responsible for the development of new cells—a capsule that is capable of safely crossing the BBB.

The shell of the nanocapsule was created using a disulfide, cross-linked polymer that was then dotted with an angiopep-2 peptide. The peptide was added to create a neutral surface charge so that it would not be attacked by ribonuclease. The shell was just large enough to hold a Cas9 complex but still small enough (approximately 30 nanometers in length) to allow it to pass through the BBB.

The researchers tested their nanocapsule delivery system in <u>glioblastoma</u> mouse models. Each was given a single tail injection—some received the newly developed nanocapsule delivery system while others received a control. The researchers found that the mice given the new therapy had a mean survival time of 68 days compared to 24 days for the control group. They also found a less than 0.5% undesired genetic mutation rate in other brain tissue.

The researchers suggest their work represents a new step toward noninvasive and nonviral approaches to treating glioblastomas, though they acknowledge that much more work needs to be done before it can be determined if the same approach will be both safe and effective in humans.

More information: Yan Zou et al, Blood-brain barrier–penetrating single CRISPR-Cas9 nanocapsules for effective and safe glioblastoma gene therapy, *Science Advances* (2022). <u>DOI: 10.1126/sciadv.abm8011</u>



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