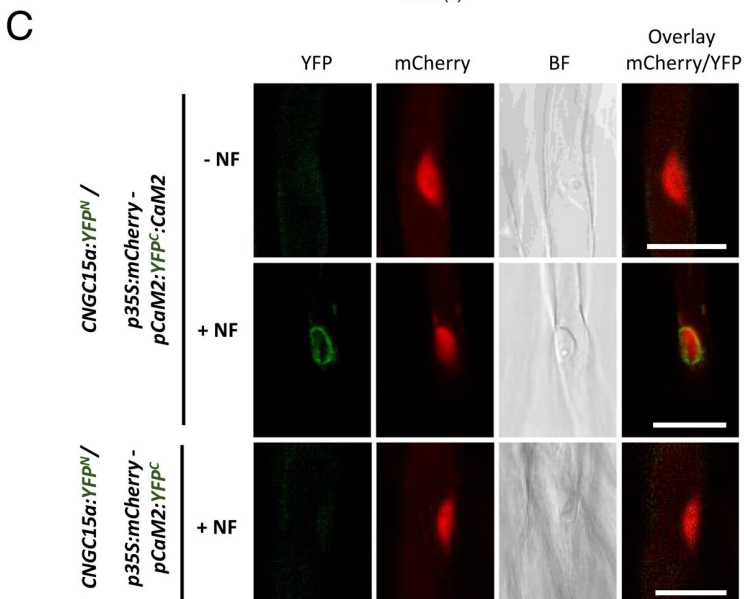
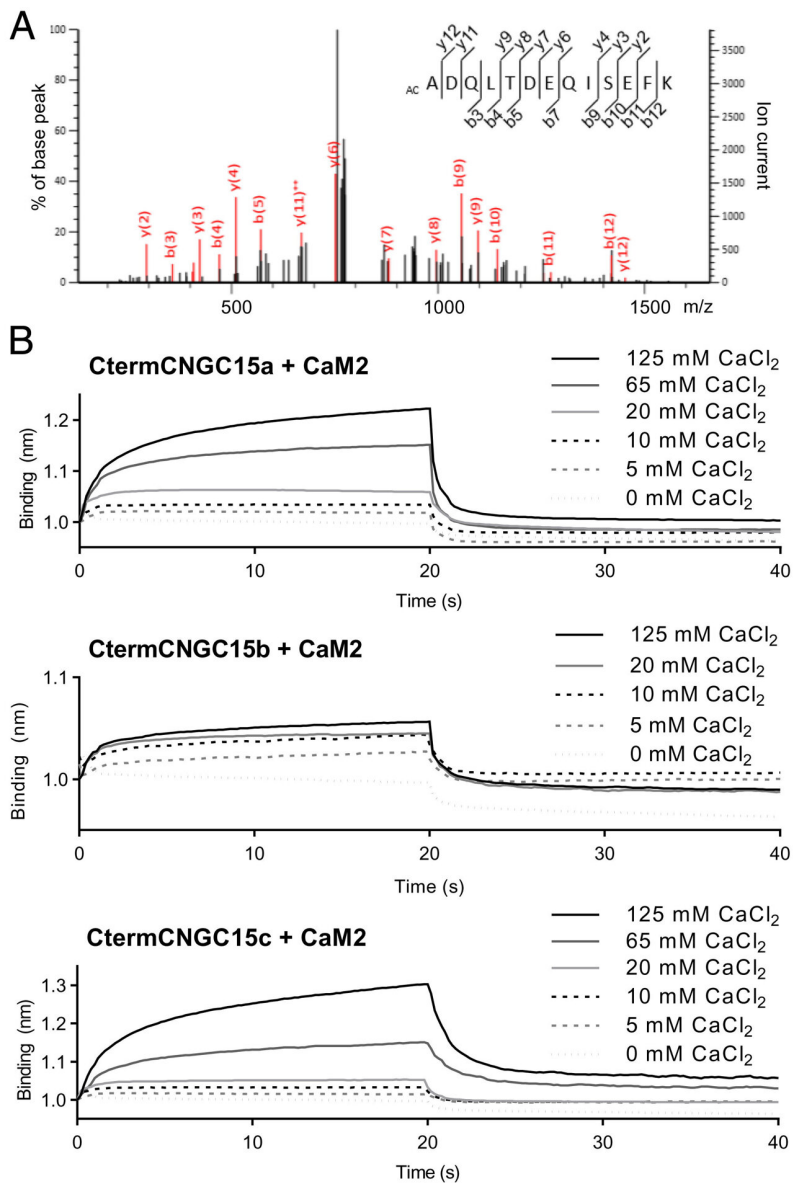


Modulation of calcium signaling to enhance root nodule symbiosis

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HoloCaM2 interacts with CNGC15a, -b, and -c. (A) Representative mass spectra of specific peptide of CaM2, ADQLTDEQISEFK, identified by IP of CNGC15a in *M. truncatula* roots in the presence of calcium, with a probability of 100%. (B) Biolayer interferometry analyses of CaM2 binding to CtermCNGC15a, CtermCNGC15b, or CtermCNGC15c. CaM2 binds to CtermCNGC15s specifically in the presence of calcium. The interactions are assessed with 50 μM of CtermCNGC15a, CtermCNGC15b, or CtermCNGC15c and 150 μM of CaM2 in the absence or presence of CaCl_2 at the concentration indicated. The raw curves represent the average of three replicates normalized to control run and performed using three independent protein purifications. Graphs show the association (0 to 20 s) and dissociation (20 to 40 s) steps of the interaction. (C) Bimolecular fluorescence complementation (BiFC) assessed in root hair cells of the *M. truncatula* transgenic line CNGC15a:Myc:YFP^N expressing under the promoter CaM2 CtermYFP (YFP^C) fused or not to CaM2. The interactions are visualized in the absence (–) or presence (+) of 10^{-8} M of Nod factor (NF) incubated for 1 h. Representative pictures of three biological replicates. (Scale bar, 20 μm .). Credit: *Proceedings of the National Academy of Sciences* (2022). DOI: 10.1073/pnas.2200099119

How plants acquire nutrients is a fundamental life process. Some plants have developed beneficial associations with bacteria and fungi to help them access essential elements such as phosphate and nitrogen.

Legumes such as beans and peas form symbiotic relationships with nitrogen fixing [bacteria](#) known as rhizobia. This capacity has positioned [legumes](#) as key crops to develop sustainable agricultural practices in both developed and developing countries. Understanding the process which allows these endosymbiotic relationships to happen may allow us to fine tune the physiology of legume crops.

By explaining the chemical signaling networks that underpin

endosymbiosis, researchers can help address agricultural pollution—one of the main challenges facing us.

Previous research by Dr. Myriam Charpentier's group at the John Innes Centre had unearthed a protein that controls oscillations of calcium produced in the nucleus membrane of plant root cells. These calcium signals are activated upon recognition by the plant of beneficial bacteria in the soil. This in turn triggers the formation of root nodules which become home to nitrogen fixing bacteria—which harvest atmospheric nitrogen and pass it onto the plant in return for nutrients.

But it was unclear how this oscillation-mechanism was being modulated. In a new study which appears in *PNAS (Proceedings of the National Academy of Sciences)*, the John Innes Centre researchers take an important new step in completing this picture by identifying the role of the signaling protein (CaM2) which regulates the [calcium channels](#).

Using an engineering strategy, they show that the newly described molecule shapes the pattern of calcium signals. It does this by closing down the calcium channel which accounts for the trough in the oscillations.

Plant roots expressing the engineered molecule showed accelerated calcium frequency, earlier signaling with bacteria and showed enhanced root nodule symbiosis.

"Our study shows that we can engineer specific signaling [molecules](#) to study their function directly in plants and that modulating calcium signaling could serve as a valuable strategy to enhance [nitrogen](#) fixing bacteria symbiosis," said Dr. Charpentier.

"Engineered CaM2 modulates nuclear [calcium](#) oscillation and enhances legume root nodule symbiosis" is published in *PNAS*.

More information: Pablo del Cerro et al, Engineered CaM2 modulates nuclear calcium oscillation and enhances legume root nodule symbiosis, *Proceedings of the National Academy of Sciences* (2022).

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Provided by John Innes Centre

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