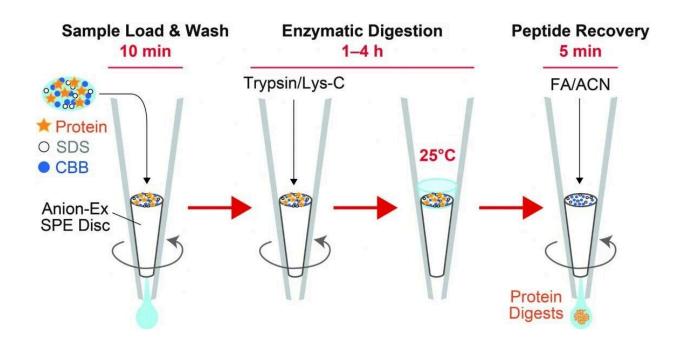


Fast and easy protein analysis from trace amounts of cell samples

March 2 2022



Sample preparation that used to take more than 20 hours with conventional methods can now be completed in less than 5 hours with AnExSP. Credit: *Chemical Communications*, DOI: 10.1039/D1CC05529A

Mass spectrometry is an excellent protein analysis method widely used in medicine and agriculture, but it requires complicated and time-consuming sample preparation. In this study, we developed a new method for significantly reducing preparation time using an anion-exchange solid-phase extraction spin column that can be made from

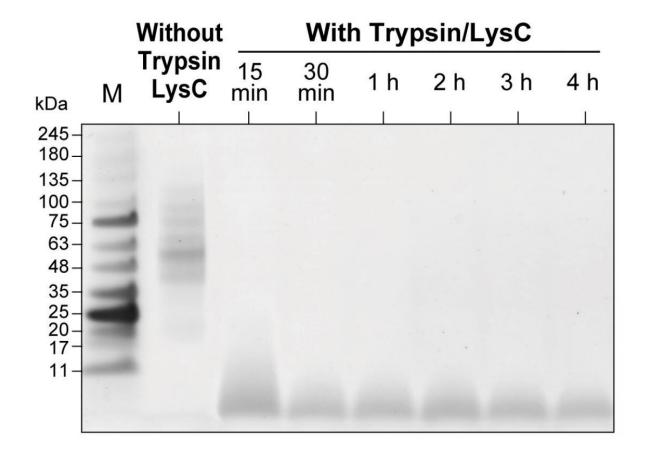


readily available materials. This new and reliable method does not require highly specialized lab technique and enables reproducible mass spectrometry from trace cell and tissue samples.

In recent years, proteomics research using mass spectrometry to comprehensively investigate the protein components of biological samples has become very popular. In order to detect proteins by mass spectrometry, it is necessary to extract the protein components contained in the cells and digest them with proteases to a peptide size that is easy to analyze by mass spectrometry. Protein digestion in this manner typically requires a long reaction time of 20 hours or more. Sodium dodecyl sulfate (SDS), a surfactant often used in protein extraction processes, interferes with mass spectrometry and protein digestion and must be thoroughly removed prior to analysis. However, that process comes with a cost, namely sample loss. To overcome these challenges, we developed an innovative sample preparation method, AnExSP (anion-exchange disk-assisted sequential sample preparation), which uses a microliter-sized spin column fitted with anion-exchange solid-phase extraction disks in a pipette tip called a StageTip as a tool for protein digestion.

We succeeded in completing the enzymatic digestion process in a minimum of 60 minutes by enriching the protein components on the disk surface. Furthermore, we established optimal conditions for eluting only the digested peptides from the disks while keeping the SDS contained in the sample on the disks, and achieved sample pretreatment with simple operation and minimal loss. By combining AnExSP with data-independent acquisition (DIA) analysis by Orbitrap mass spectrometry, we succeeded in detecting approximately 7,000 different protein components in one microgram of human cultured cell protein extract. By establishing sample pretreatment conditions using the reduced-size StageTip in the future, AnExSP pretreatment can be used for single-cell proteomics research, which is of growing interest in recent years.





AnExSP allows for digestion of human cell extracts in just 60 minutes. Credit: *Chemical Communications*, DOI: 10.1039/D1CC05529A

AnExSP can also be used for pretreatment of proteins separated by SDS-polyacrylamide gel electrophoresis (SDS-PAGE), which is commonly used in protein experiments. Compared to conventional pretreatment methods, AnExSP demonstrated superior performance, especially in the detection of long-chain digested peptides. In this study, we combined AnExSP and SDS-PAGE with a chemical cross-linking method for protein complexes, and succeeded in simple purification of target protein complexes and efficient detection of long-chain cross-linked peptides that are difficult to detect with conventional methods. Because



of its far-reaching versatility, AnExSP is expected to be applied to highly sensitive structural analysis methods for trace <u>protein</u> complexes in biological samples.

More information: Ayako Takemori et al, Bottom-up/cross-linking mass spectrometry via simplified sample processing on anion-exchange solid-phase extraction spin column, *Chemical Communications* (2021). DOI: 10.1039/D1CC05529A

Provided by Ehime University

Citation: Fast and easy protein analysis from trace amounts of cell samples (2022, March 2) retrieved 23 June 2024 from https://phys.org/news/2022-03-fast-easy-protein-analysis-amounts.html

This document is subject to copyright. Apart from any fair dealing for the purpose of private study or research, no part may be reproduced without the written permission. The content is provided for information purposes only.