

DNA strong bonding—a long-term commitment or many brief relationships?

January 27 2022



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In an article in the *Science*, researchers at Uppsala University show how a DNA-binding protein can search the entire genome for its target sequence without getting held up on the way. The result contradicts our current understanding of gene regulation—the genetic code affects how often the proteins bind, but not for how long.

Over an organism's lifetime, its genome changes very little. What does change, constantly, are which proteins the cell produces in response to damage, changes in the environment, or stages in the reproductive cycle. The [protein](#) production is regulated by DNA-binding proteins that have evolved the ability to turn different genes on or off. Because the environment can change quickly, rapid adaptation is key. The DNA-binding proteins must find the correct DNA code among millions of base pairs, and do so fast.

When DNA-binding proteins search the genetic code for their target sequence, they slide along the DNA helix to speed up the process. When they finally find the right spot, they stay there; the interaction with the "correct" sequence prevents them from sliding along. This mechanism has been widely accepted to describe the search process. It is an appealing hypothesis, yes, but it presents an annoying problem—the DNA code is full of "almost correct" sequences. If the time a protein resides on a particular DNA motif was determined by the sequence, the searching proteins would constantly linger on sequences that resembled their target.

"If the textbook explanation was correct, the DNA-binding proteins would get stuck all the time off target. Gene regulation would be very ineffective, but we know from previous studies that this is not the case.

Our favorite protein, LacI, finds its target sequence among 4.6 million base pairs in a matter of minutes", says Emil Marklund, one of the researchers behind the discovery.

In an attempt to resolve this paradox, the researchers allowed the DNA-binding protein LacI to slide back and forth on thousands of different DNA sequences mounted on a microchip. A [fluorescent molecule](#) was attached to the LacI protein and made it possible to measure how fast LacI adhered to the different sequences and how quickly it was released. The result was striking. Contradicting previous assumptions, the DNA sequence had little effect on how long LacI remained bound to the DNA. However, it was much more likely that the sliding LacI was held up briefly when the sequence was similar to the target sequence. In other words, DNA-binding proteins often leave also the sequence they are intended to regulate, but at the target site, they all but always make a very short journey before finding their way back again. On the macroscopic time scale, this looks like a stable interaction.

"Our result, that DNA-binding proteins bind often rather than protractedly, explains how LacI can slide on the DNA sequence in search of its [target](#) without getting held up unnecessarily. LacI regulates the uptake of lactose in bacteria, but is of course just an example. The hundreds of different transcription factors that regulate our own genes likely act according to a similar principle," says Johan Elf, Professor at the Department of Cell and Molecular Biology at Uppsala University and the national research infrastructure SciLifeLab.

More information: Emil Marklund et al, Sequence specificity in DNA binding is mainly governed by association, *Science* (2022). [DOI: 10.1126/science.abg7427](https://doi.org/10.1126/science.abg7427).
www.science.org/doi/10.1126/science.abg7427

Provided by Uppsala University

Citation: DNA strong bonding—a long-term commitment or many brief relationships? (2022, January 27) retrieved 20 June 2024 from <https://phys.org/news/2022-01-dna-strong-bondinga-long-term-commitment.html>

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