

The world in a drop of water: DNA tool transforms nature tracking

September 7 2021, by Kelly MacNamara



Some animals, like the Amazon pink river dolphin are tricky to track using conventional methods.

In their search for pink river dolphins, researchers in the Peruvian Amazon scooped up river water sloshing with genetic material that they

hoped could trace the elusive creatures.

They found what they were looking for. And then some.

The environmental DNA collected yielded information on 675 species, including dozens of land-based mammals like deer, jaguar, giant anteaters, monkeys and 25 species of bat.

"It's kind of mind blowing," said Kat Bruce, founder of the eDNA firm NatureMetrics, which carried out the study for the wildlife charity WWF.

The technology is increasingly used to track rare species.

Bruce hopes eDNA will help revolutionise the way the world measures and monitors nature.

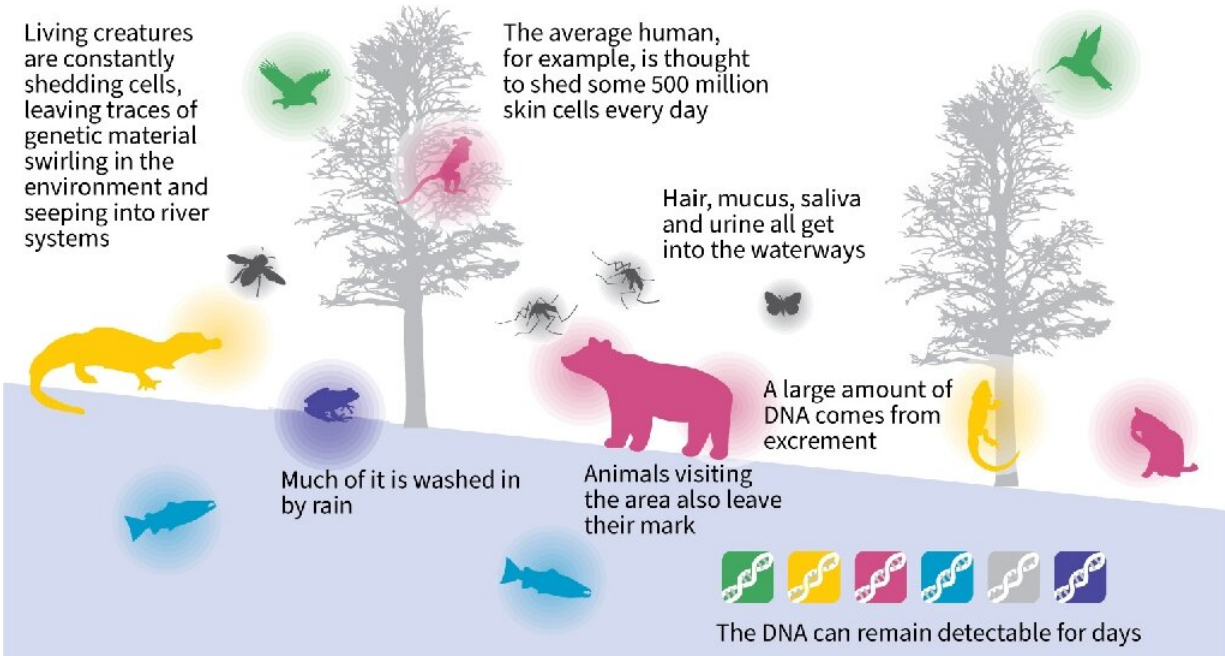
It is now at the heart of a \$15 million dollar project with the International Union for Conservation of Nature (IUCN) to collect and analyse 30,000 freshwater samples over three years from major river systems—including the Amazon, Ganges and Mekong Delta.

With species in precipitous decline and growing calls for international targets on biodiversity protection, organisers say this "eBioAtlas" can help inform policy and focus scarce conservation resources.

"What the eBioAtlas will do in the middle of this mass extinction, is hopefully start to fill those gaps in in a way that is scalable," said Mike Morris, who heads the project for NatureMetrics, at an event showcasing the project at the IUCN conference in Marseille this week.

Traces of life

Tracking waterborne environmental DNA (eDNA) to profile the life surrounding us



Traces of life.

The IUCN plans to feed the information into conservation tools like its Red List of Threatened Species.

It offers a "simple and I would say precise way to tell us where species are," said Paola Geremicca, who leads IUCN's corporate engagement.

'Tickle a fish'

The method relies on that fact that living creatures constantly shed cells, leaving traces of [genetic material](#) swirling in the environment that wind up in river systems: skin, mucus, saliva.

But most of all "it's from a lot of poo," Bruce told AFP, both from fish and animals in the rivers and those just passing by.

Scientists have been using eDNA for at least a decade, with early work focused on microbes.

Bruce started using the technique during her PhD studies, blending assortments of insects into a "soup" and figuring out what creatures were inside using genetic sequencing.

That led her to set up NatureMetrics, which focuses on finding DNA "fingerprints" in freshwater.

Researchers collect a litre or two of water then pass it through a small filter, which traps the sample.



There is a shortage of reference DNA for identifying fish species.

To sequence the DNA they need to decide what to look for—mammals for example—because otherwise the result would be swamped with ubiquitous bacteria and microbes.

After two days their machine spits out about 30 million DNA sequences—imagine a text file 30 million lines long and crammed with A's, T's, C's and G's, the molecular building blocks of life.

While they are confident in the accuracy of the sampling itself, Bruce said the problems come with anomalies and gaps in the identification reference database.

In the Amazon for example, only 20 percent of fish could be identified to species level because the genetic information did not match any sequences in the library.

In these types of situations, they contact local zoos and museums looking for a reference specimen.

Or they could find the species in the wild.

"We've got these simple kits, where people can just tickle the fish with the swab and send that back to us to generate the reference sequence," she said.

'Game changer'

When results are transmitted back to local conservationists, Morris said people are often "astonished" at the array of hidden wildlife.



All living things shed DNA traces that often gets into waterways.

Other researchers have called this "dark diversity".

In research published in the journal *Science Advances* in 2018, scientists took 20 coastal water samples in New Caledonia for eDNA sequencing and found more shark species than they had previously identified in two decades of visual and camera surveys.

"Even if people in Noumea never see sharks when they go diving... they are always there," said David Mouillot from the University of Montpellier and co-author of the research, at the IUCN congress.

He works with another eDNA initiative, Vigilife, an international public-private alliance developing long-term biodiversity monitoring and sharing technological expertise with local conservationists.

Morris acknowledged that eDNA sampling should be seen as one biodiversity tool among many.

A blindspot is plants, which are harder to identify at [species](#) level.

The eBioAtlas developers, who plan to create an open source database, hope to spread their net beyond freshwater—with plans for marine and soil eDNA surveys.

After that they might snatch samples from thin air.

Separate preliminary studies this year in Britain and Denmark sucked air through filters in zoo enclosures and not only identified resident animal DNA, but also local wildlife.

"For things like caves, bat roosts and burrows, that's potentially a game changer," said Bruce.

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