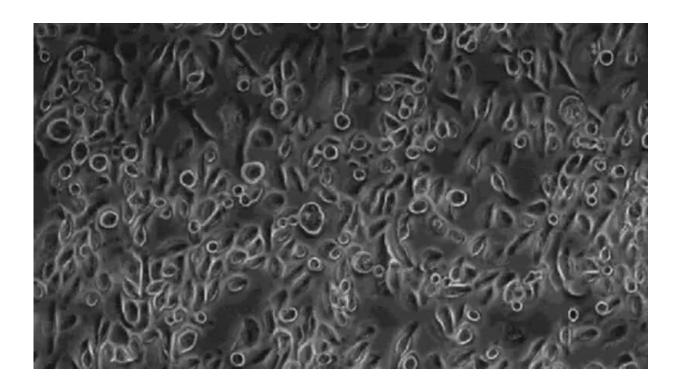


Researchers take step toward using cellular motion to help wound healing

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With a technique that overcomes cells' innate social behaviors, researchers have taken an important step in directing skin cells to migrate en masse to close wounds—"literally making skin crawl," said principal investigator Daniel Cohen.

In a new study, the researchers overcame the inertia typical of mature



skin tissue by breaking the molecular connections between cells, applying an <u>electrical field</u> to direct their migration and then rebuilding the connections. This novel approach improves the controllability of tissues and may one day help optimize wound healing through electrical stimulation.

The German physician Emil du Bois-Reymond first described an electric current flowing from a cut on his finger in 1848. Later research showed that cells in the body can sense and follow an electric field, a process called electrotaxis. Electric fields generated in the body promote healing by directing cells to move toward the wound and are also vital for growth and development.

"There are a lot of reasons why people think electrical stimulation might help wound healing," said Gawoon Shim, the study's lead author and a graduate student in Cohen's lab. But despite promising clinical evidence from decades of use in patients, scientists have yet to work out how cells detect and respond to electric fields or how electrical stimulation can best be applied therapeutically. "It's kind of a black box," said Shim.

"We discovered this basic phenomenon 175 years ago and we don't yet have commercial electric band aids," said Daniel J. Cohen, an assistant professor of mechanical and aerospace engineering and senior author of the <u>study</u>, published online in the July 20 issue of the *Proceedings of the National Academy of Sciences*. "No one really knows the rules for how to design these things."

In their previous work, Cohen's group used electric fields to program thousands of <u>individual cells</u> to move <u>in circles and around corners</u>. Their new study used a model of more mature skin—a single layer of mouse skin cells all latched together—which is harder to control. Instead of moving with the speed and precision of a marching band in response to an <u>electric current</u>, the mature skin cells inched along like a crowd of



people holding hands with their neighbors.

The mature skin also posed another problem: Once the leading edge of cells advanced, it would peel away from the petri dish and die. "If you apply a command that differs from what the cells naturally 'want' to do, you get a tug-of-war," Cohen said. "The result was the tissues ripped themselves apart."

Cohen and Shim suspected that the "handshakes" between cells prevented the tissue from fluidly following the electrical commands. These handshakes are proteins called cadherins that anchor neighboring cells together. They make tissues cohesive so they can move together but can also create traffic jams when cells don't have space to move.

Cadherins need <u>calcium ions</u> to complete their connections, so Shim grew the cells with different amounts of calcium and measured their response to <u>electrical stimulation</u>. She saw that the less calcium the cells had, the more fluid they became and the quicker they moved. "It goes really fast—I was very surprised," said Shim.

Calcium has many effects on living tissues, however, so Shim had to confirm that the handshakes were to blame for the slow movement. She grew cells with an antibody that attaches to cadherins. With blocked handshakes, these cells moved more quickly.

After working out the ground rules of cell adhesiveness, the researchers developed a solution to their sticky cell problem. Shim grew a layer of skin cells in a high calcium solution so they made their normal connections. Then she treated the cells with a chemical that grabs up calcium ions to break up the cellular handshakes. When Shim lowered the calcium level and applied the electric field, the cells moved on command. Finally, she restored the high calcium level to reinstate the handshakes, resulting in a healthy and cohesive layer of <u>skin cells</u>.



To demonstrate that this approach has the potential to accelerate healing, Shim performed the above experiment using an electrobioreactor developed in the Cohen lab that mimics the closing of a wound. Unlike other models of electrotaxis where the electric field moves cells in one direction, their new system exposes cells to an <u>electric field</u> focused on the center of the injury. Shim showed that the stimulated tissues successfully came together while the unstimulated ones remained largely separate. Cohen's group described their electrobioreactor in a <u>new paper</u> in *Biosensors and Bioelectronics*.

"This concise and exciting study from Cohen's lab results in an intuitive yet previously unknown lesson: collectively migrating cells follow directional cues more readily if their mutual cohesion is weaker," said Alex Mogilner, a Professor of Mathematics and Biology at New York University who studies the biophysics of collective behaviors and was not involved in the research. "This paper is not just basic science but also has far-reaching biomedical implications... [Cohen] asks not only how the cells do it, but also how can we manipulate the cells and make them do better."

Next, Shim and Cohen plan to move from their two-dimensional model into a 3D one. Human skin, for example, is composed of different tissues, much like a layer cake. Depending on how these techniques work in a 3D skin model, the results could indicate whether the same approach will work on actual wounds. Many tissues use cadherin handshakes to stick <u>cells</u> together, but there are other ways tissues stay connected.

This work could be applied not only to help wounds heal faster, but also to fine-tune the healing process to prevent tightness and reduce scarring.

"We started out with this nice, warm and fuzzy approach to engineering group behaviors by saying, let's see what the <u>tissue</u> wants to do and



harmonize with that. It turns out, that's too simple," said Cohen. "Sometimes, they don't want to listen to you properly. Sometimes you just have to change the rules."

More information: Gawoon Shim et al, Overriding native cell coordination enhances external programming of collective cell migration, *Proceedings of the National Academy of Sciences* (2021). DOI: 10.1073/pnas.2101352118

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