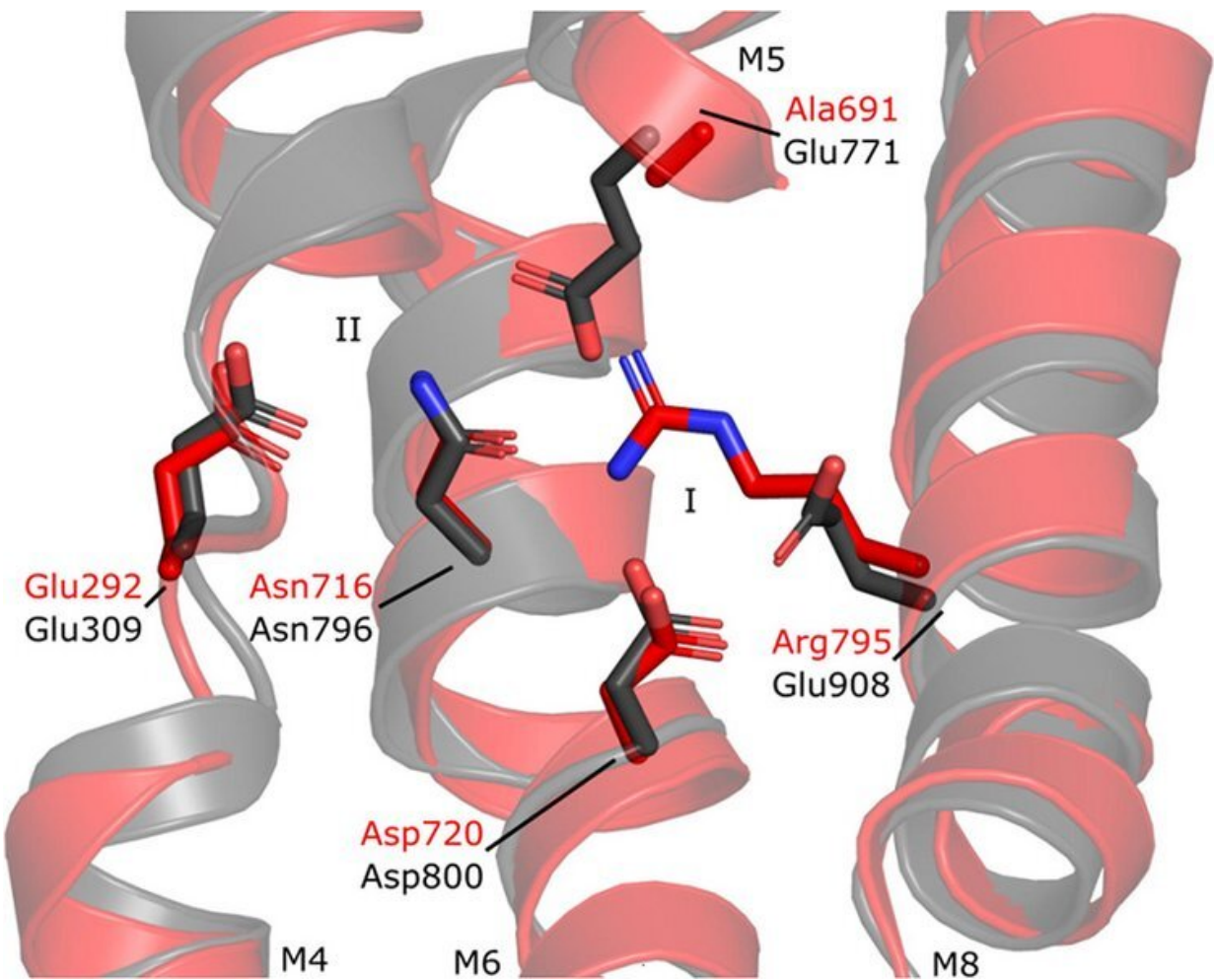


Ion and lipid transporters specialize for their niche

June 10 2021, by Lisbeth Heilesen



Binding site I of SERCA is not conserved in LMCA1. Alignment between LMCA1 G4 E2-BeF₃⁻ (red) and SERCA E2-AlF₄⁻ (pdb: 3b9r) (dark grey). Relevant Ca²⁺-coordinating residues are shown as sticks. The structures are aligned by the residues shown as sticks. SERCA Ca²⁺ binding site I and II are indicated. Credit: Sara Basse Hansen.

Cell viability requires that a variety of functions at the cell membrane are maintained properly. P-type ATPases translocate substrates across the membrane, and they have evolved into different types taking care of specific substrates within a diverse range. Now, key structural aspects have been described on how two different types of P-type ATPases—a Ca^{2+} transporting Ca^{2+} -ATPase and a lipid transporting P4-ATPase—have adapted to different substrates and physical environments.

Many bacteria export intracellular calcium using active transporters homologous to the well-described mammalian Ca^{2+} -ATPases such as plasma-membrane Ca^{2+} -ATPase and sarco-endoplasmic reticulum Ca^{2+} -ATPase (PMCA and SERCA, respectively). Crystal structures of Ca^{2+} -ATPase 1 from *Listeria monocytogenes* (LMCA1) suggest that LMCA1 is pre-organized for dephosphorylation upon Ca^{2+} release, which can explain the rapid dephosphorylation observed earlier in single-molecule studies.

Also, variation in the architecture of the calcium binding sites explains why LMCA1 transports a single Ca^{2+} ion similar to PMCA, in contrast to two transported Ca^{2+} ions in SERCA. The LMCA1 structures provide insight into the evolutionary divergence and conserved features of this important class of ion transporters that also inform us on central mechanisms of mammalian Ca^{2+} -ATPases and how they can be regulated or affected by pathological conditions.

For the P4-ATPase study, researchers took a different perspective. The [transport](#) cycle of a P-type ATPase consist of two half-reactions. Phosphorylation where a phosphate is transferred from ATP to the transporter, and dephosphorylation, where the phosphate is again released. In contrast to ion transporters such as LMCA1, the

P4-ATPases transport lipids and are known as lipid flippases. Importantly, the lipid transport is coupled to the dephosphorylation reaction of the cycle, where for ion transporting P-type ATPases it is mainly coupled to the phosphorylation reaction.

Through new structures determined by cryo-electron microscopy (cryo-EM) of a yeast lipid flippase, Drs2p/Cdc50p, it was investigated how the [lipid](#) flippases have diverged from ion transporters and have adapted the enzymatic mechanism for the 'flipped' purpose. Cryo-EM was a critical technique for this study, and multiple structures of the transport cycle could be determined by locking Drs2p/Cdc50p using different inhibitors and electron microscopy data collected at the [electron microscopy](#) infrastructure facility at Aarhus University (EMBION).

The two studies have been spearheaded by Ph.D. student Sara Basse Hansen and Postdoc Milena Timcenko—under the supervision of Professor Poul Nissen (and Sara also of Associate Professor Magnus Kjærgaard)—and are being published in *Journal of Molecular Biology*.

More information: Sara Basse Hansen et al, The Crystal Structure of the Ca²⁺-ATPase 1 from *Listeria monocytogenes* reveals a Pump Primed for Dephosphorylation, *Journal of Molecular Biology* (2021). [DOI: 10.1016/j.jmb.2021.167015](https://doi.org/10.1016/j.jmb.2021.167015)

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