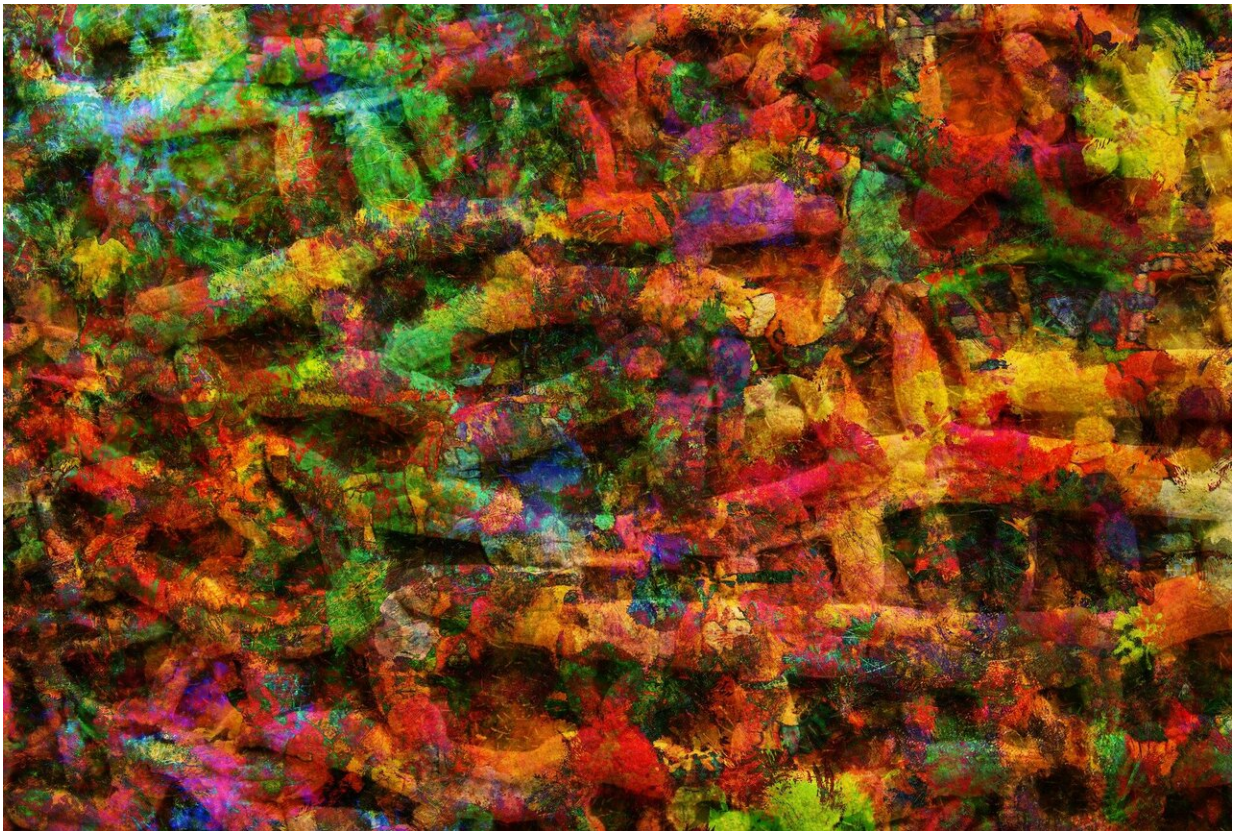


Herbivore gut fungi found to produce unique building blocks of antibiotics

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For the past several years, chemical engineer Michelle O'Malley has focused her research on the anaerobic fungi found in the guts of herbivores, which make it possible for those animals to fuel themselves

with sugars and starches extracted from fibrous plants. O'Malley's work, reflected in multiple research awards and journal articles, has centered on how these powerful fungi might be used to extract value-added products from the nonedible parts of plants—roots, stems and leaves—that are generally considered waste products.

Now, her lab has discovered that those same [fungi](#) likely produce novel "[natural products](#)," which could function as [antibiotics](#) or other compounds of use for biotechnology. The research is described in a paper titled "Anaerobic gut fungi are an untapped reservoir of natural products," published in the *Proceedings of the National Academy of Sciences (PNAS)*.

All living things are equipped with natural defenses to ensure their survival. Microbes often depend on synthesized natural products that act as defenses against environmental threats and allow them to compete with other microbes. Many such natural products have served as the source of antibiotics used to fight disease in humans.

Anaerobic fungi, by definition, do not use oxygen to fuel their production of natural products. Thus, said O'Malley, "It costs them a lot more metabolically to make something than it does an aerobe." Further, anything an anaerobic microbe produces has to be carefully "designed" and extremely efficient, since energy is scarce in the oxygen-free microbes.

O'Malley's lab in 2017 provided the U.S. Department of Energy with several anaerobic gut fungi, which were then sequenced as part of a larger collaboration.

"We started seeing something we didn't expect to find: The [building blocks](#)—biosynthetic gene clusters, or BGCs—that antibiotics are typically built from," O'Malley said of that work. "BGCs sit near each

other on the genome and participate in stepwise chemical reactions. In this case, they are loading a molecule onto an enzyme—chemically decorating it, if you will—and then passing it to another enzymatic module, and in this way, making it into an increasingly complex molecule."

That molecular assembly-line process is important, she says, because it emulates how antibiotics are often made. "BGCs could also be of useful for making value-added chemicals, because they have lot of complex chemistry built in," she added. "So, they might be able to be used as drop-in biofuels, as coatings, and as monomers to make novel materials."

But the most important implications, she said, are the antibiotic possibilities. In the paper, lead author Candice Swift, who earned her Ph.D. in O'Malley's lab and is now a postdoctoral researcher at the University of South Carolina's School of Public Health, demonstrates not only that the BGCs are present where they were not expected to be—anaerobic fungi have never been thought to have antibiotic properties—they are, even more surprisingly, active and actually making something. "They are transcribed," O'Malley noted.

Swift's analysis suggests that the compounds they found have not been identified previously. "We discovered them, and, based on what is known about these biosynthetic gene clusters, they could be making novel antibiotics," O'Malley said. "We haven't shown that definitively here, but it's one possibility that would make sense, because the fungi exist as kind of minority players in their community, so they probably have some edge or ability that allows them to stick around in the face of overwhelming competition. That's what we think they might be doing. We haven't proven it, but it's an attractive way to look at it."

So, if the anaerobic fungi are producing natural antibiotics, how difficult would it be to translate them into use in humans? "If you identify a novel

antibiotic, the key to making that at scale is knowing how it's built," O'Malley explained. "That's a big question. We'll try to address that in a follow-up paper, but if you can figure out the genetic recipe for how they're made, it's straightforward to apply it in another system."

O'Malley said that the scientific community "has become pretty good at making antibiotics, recombinantly or through genetic engineering," and that all, or nearly all antibiotics, are derived from natural products—either made in nature or inspired by the compounds that nature builds.

The next stage in this research will involve using genetic tools to boost production and isolate the compounds to figure out exactly what compounds are being made. For that work, O'Malley is partnering with other researchers in the BioPolymers, Automated Cellular Infrastructure, Flow, and Integrated Chemistry Materials Innovation Platform (BioPACIFIC MIP), which the National Science Foundation funded last year as a collaboration between UCSB and UCLA.

More information: Candice L. Swift et al, Anaerobic gut fungi are an untapped reservoir of natural products, *Proceedings of the National Academy of Sciences* (2021). [DOI: 10.1073/pnas.2019855118](https://doi.org/10.1073/pnas.2019855118)

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