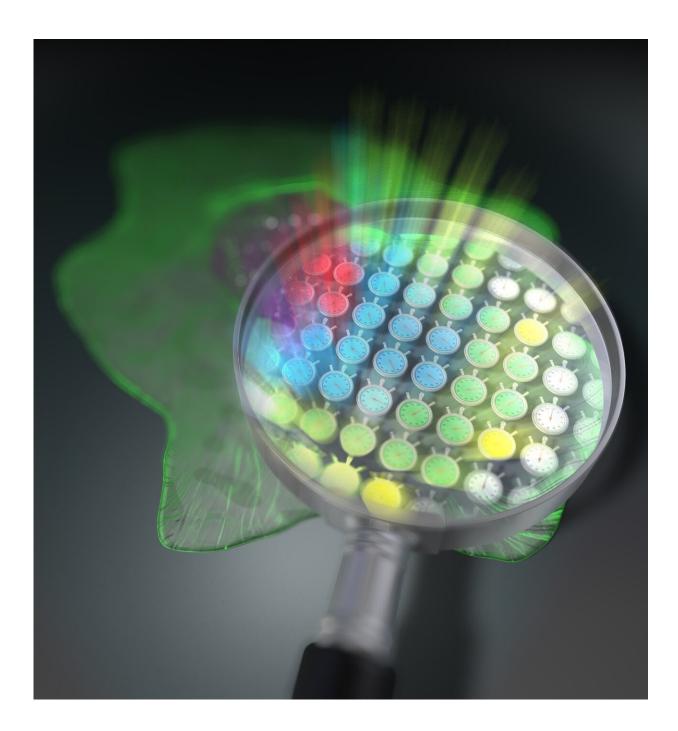


## **Comb of a lifetime: A new method for fluorescence microscopy**

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2-D arrangement of 44,400 light stopwatches enables scan-less fluorescence lifetime imaging. Credit: Tokushima University

Fluorescence microscopy is widely used in biochemistry and life sciences because it allows scientists to directly observe cells and certain compounds in and around them. Fluorescent molecules absorb light within a specific wavelength range and then re-emit it at the longer wavelength range. However, the major limitation of conventional fluorescence microscopy techniques is that the results are very difficult to evaluate quantitatively; fluorescence intensity is significantly affected by both experimental conditions and the concentration of the fluorescent substance. Now, a new study by scientists from Japan is set to revolutionize the field of fluorescence lifetime microscopy.

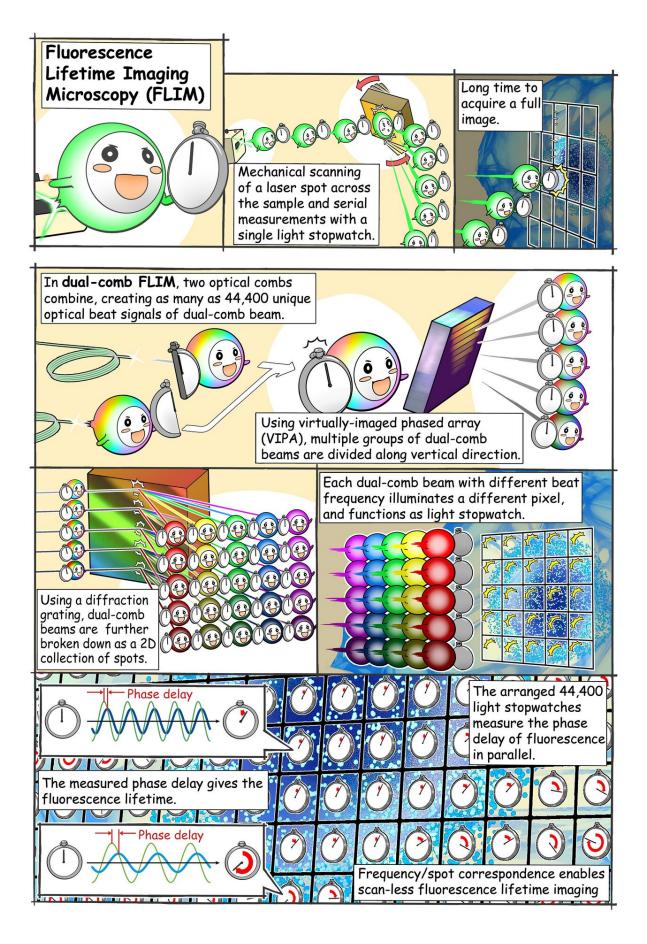
A way around the conventional problem is to focus on <u>fluorescence</u> lifetime instead of intensity. When a fluorescent substance is irradiated with a short burst of light, the resulting fluorescence does not disappear immediately but actually "decays" over time in a way that is specific to that substance. The fluorescence lifetime microscopy technique leverages this phenomenon, which is independent of experimental conditions, to quantify fluorescent molecules and changes in their environment. However, fluorescence decay is extremely fast, and ordinary cameras cannot capture it. While a single-point photodetector can be used instead, it has to be scanned throughout the <u>sample</u>'s area to be able to reconstruct a complete 2-D picture from each measured point. This process involves movement of mechanical pieces, which greatly limits the speed of image capture.

In this recent study, published in Science Advances, the team of scientists



developed a novel approach to acquire fluorescence lifetime images without the need for mechanical scanning. Professor Takeshi Yasui, from Institute of Post-LED Photonics (pLED), Tokushima University, Japan, who led the study, says, "Our method can be interpreted as simultaneously mapping 44,400 light-based 'stopwatches' over a 2-D space to measure fluorescence lifetimes—all in a single shot and without scanning."







This new fluorescence microscopy technique will measure both fluorescence intensity and lifetime and it will not require mechanical scanning of a focal point; instead, it will produce images from all points in the sample simultaneously, enabling a more quantitative study of dynamic biological and chemical processes. Credit: Suana Science YMY

One of the main pillars of their method is the use of an optical frequency comb as the excitation light for the sample. An <u>optical frequency comb</u> is essentially a <u>light signal</u> composed of the sum of many discrete optical frequencies with a constant spacing in between them. The word "comb" in this context refers to how the signal looks when plotted against optical frequency: a dense cluster of equidistant spikes rising from the optical frequency axis and resembling a hair comb. Using special optical equipment, a pair of excitation frequency comb signals is decomposed into individual optical beat signals (dual-comb optical beats) with different intensity-modulation frequencies, each carrying a single modulation frequency and irradiated on the target sample. The key here is that each light beam hits the sample on a spatially distinct location, creating a one-to-one correspondence between each point on the 2-D surface of the sample (pixel) and each modulation frequency of the dualcomb optical beats.

Because of its fluorescence properties, the sample re-emits part of the captured radiation while preserving the frequency-position correspondence. The fluorescence emitted from the sample is then simply focused using a lens onto a high-speed single-point photodetector. Finally, the measured signal is mathematically transformed into the frequency domain, and the fluorescence lifetime at each "pixel" is easily calculated from the relative phase delay that exists between the



excitation signal at that modulation frequency versus the one measured.

Thanks to its superior speed and <u>high spatial resolution</u>, the microscopy method developed in this study will make it easier to exploit the advantages of fluorescence lifetime measurements. "Because our technique does not require scanning, a simultaneous measurement over the entire sample is guaranteed in each shot," says Prof. Yasui, "This will be helpful in <u>life sciences</u> where dynamic observations of living cells are needed." In addition to providing deeper insight into <u>biological processes</u> , this new approach could be used for simultaneous imaging of multiple samples for antigen testing, which is already being used for the diagnosis of COVID-19.

Perhaps most importantly, this study showcases how optical frequency combs, which were only being used as "frequency rulers," can find a place in microscopy techniques to push the envelope in life sciences. It holds promise for the development of novel therapeutic options to treat intractable diseases and enhance life expectancy, thereby benefitting the whole of humanity.

**More information:** T. Mizuno et al. Full-field fluorescence lifetime dual-comb microscopy using spectral mapping and frequency multiplexing of dual-comb optical beats, *Science Advances* (2021). DOI: 10.1126/sciadv.abd2102,

advances.sciencemag.org/content/7/1/eabd2102

Provided by Tokushima University

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