

New research on how fungal cells respond to stress

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Researchers at the University of Maryland, Baltimore County (UMBC) have published new findings in *Molecular and Cellular Proteomics* on critical cellular processes triggered when cells respond to environmental stress. Mark Marten, professor of chemical, biochemical, and environmental engineering, led the research team, which identified three

coordinated pathways involved in the response to cell wall stress in filamentous fungi. Cynthia Chelius, who recently earned her Ph.D. in chemical engineering at UMBC, is the first author on the paper.

A previous NSF grant supported the work that Marten conducted with Ranjan Srivastava, University of Connecticut, and Steven Harris, University of Manitoba.

Numerous species of filamentous fungi are pathogens that can make people sick, especially people who are immunocompromised. Different species of fungi play an important role in the development of pharmaceuticals and enzymes, and agriculture, where fungi can help improve the quality of soil and make nutrients more readily available for crops. By understanding how cells work and respond to stress, researchers can reverse-engineer processes that could have a broad range of applications.

To understand how the fungal cell walls respond to environmental stressors, Marten and his team studied what he describes as the cell's "software"—rules that control how the cell behaves. When fungi experience stress, Marten's team found an increase in the number of septa (or cross-hyphal bulkheads) created. "When you stress cells, they sense it and try to protect themselves," Marten explains. He adds that fungi try to repair damage to their cell walls so that they can resume normal growth and function.

The study used a multi-omic methodology, which researchers say can be applied to studying how signaling networks in cells work in general. The methodology allowed researchers to get a more detailed understanding of how cells respond to stressors. They found that when cell walls experience stress, there is a coordinated response through various pathways. By combining short time-scale phosphoproteomic sampling and longer scale transcriptomic sampling, the researchers were able to

see a broader view of how cells respond to stress.

Marten, Srivatava, and Harris's teams will continue to collaborate on related research, which will be supported by a new three-year grant totaling \$1.23 million grant from the National Science Foundation. This work will further explore how [filamentous fungi](#) repair their cell walls when exposed to stressors.

The team will examine how the parts of the fungal cell are assembled and how fungal gene regulatory networks function. They hope to understand how proteins in cells interact with each other, and how [cells](#) can turn on and off certain parts of their DNA to respond to [stress](#).

"We were excited to see the results from this paper, as they both revealed a novel connection between different aspects of gene regulation in [fungi](#) and served as the basis for a new hypothesis regarding gene regulation in our most recent NSF Collaborative Research Award," says Marten.

More information: Cynthia Chelius et al, Dynamic Transcriptomic and Phosphoproteomic Analysis During Cell Wall Stress in *Aspergillus nidulans*, *Molecular & Cellular Proteomics* (2020). [DOI: 10.1074/mcp.RA119.001769](#)

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