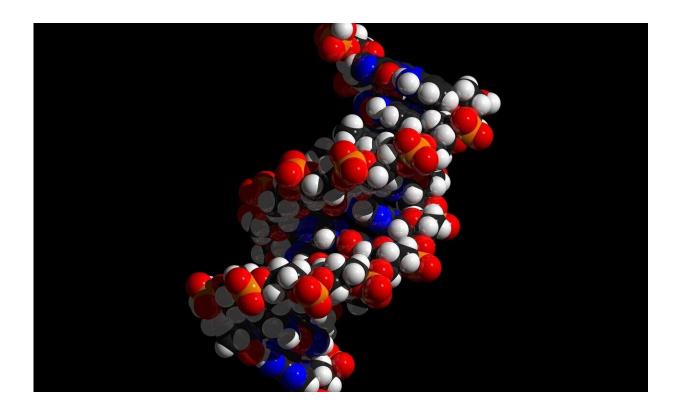


Researchers create bioluminescent tag to detect DNA break repair

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A new bioluminescent reporter that tracks DNA double stranded break (DSB) repair in cells has been developed by researchers from Massachusetts General Hospital (MGH) and the Academia Sinica in Taiwan. The international team's novel bioluminescent repair reporter (BLRR)-based system can be used to monitor DNA repair pathways



directly in animals as well as cell lines. No such system previously existed for in vivo studies. These pathways play a crucial role in multiple conditions, including cancer.

"One of the main reasons cancer cells are resistant to treatment is that they can inherently repair the DNA damage caused by radiation and chemotherapy," explains Christian Elias Badr, Ph.D., investigator in the Department of Neurology at MGH and co-senior author of the paper. The study's other co-senior author is Charles Pin-Kuang Lai, Ph.D., at the Academia Sinica in Taiwan.

Their study appeared this month as an online advance paper in *Nucleic Acids Research*.

DSB damage repair is key to maintaining genomic integrity and cell viability. It also plays a role in cancer treatment, which often includes chemoradiotherapy (radiation and chemotherapy), which disrupts DSB. A cell may recognize the damage and use its intrinsic DNA damage response (DDR) to reduce DSB-caused cell death. As a result, the cancer cell's own DNA repair mechanisms can promote drug resistance and recurrence in some malignancies. Researchers would like to know more about them.

The BLRR approach builds on earlier work members of the team did on enzymes called luciferases. These produce bioluminescence, making them useful for tracking molecules in cells. BLRR uses secreted Gaussia and Vargula luciferases to detect homology-directed repair (HDR) and non-homologous end joining (NHEJ)—the two major pathways to DSB repair. Using BLRR. Researchers can track HDR and NHEJ-related activities over time in cells. It also detects DSB repairs in xenografted tumors in vivo.

"You can study DNA damage in cells with next generation sequencing



(NGS), but that's more costly and time consuming," Badr says. "And our system's accuracy is comparable to NGS."

The researchers used their new tag to carry out multiple studies. In one, they found a significant difference in the efficiency of CRISPR/Cas9-mediated editing with guide RNAs 1-10bp apart. They also used BLRR analysis to detect altered dynamics for DSB repair induced by small molecule modulators. Lastly, they used the system to discover HDR-suppressing functions of anticancer cardiac glycosides in human glioblastomas and glioma cancer stem-like cells by inhibiting DNA repair protein RAD51 homolog 1.

In their paper, the authors describe the BLRR system as: "A highly sensitive platform to simultaneously and longitudinally track HDR and NHEJ dynamics that is sufficiently versatile for elucidating the physiology and therapeutic development of DSB <u>repair</u>." The authors plan on using this reporter system in high throughput drug screening to identify novel therapeutics that sensitize cancer <u>cells</u> to radiation and chemotherapy.

More information: Jasper Che-Yung Chien et al, A multiplexed bioluminescent reporter for sensitive and non-invasive tracking of DNA double strand break repair dynamics in vitro and in vivo, *Nucleic Acids Research* (2020). DOI: 10.1093/nar/gkaa669

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