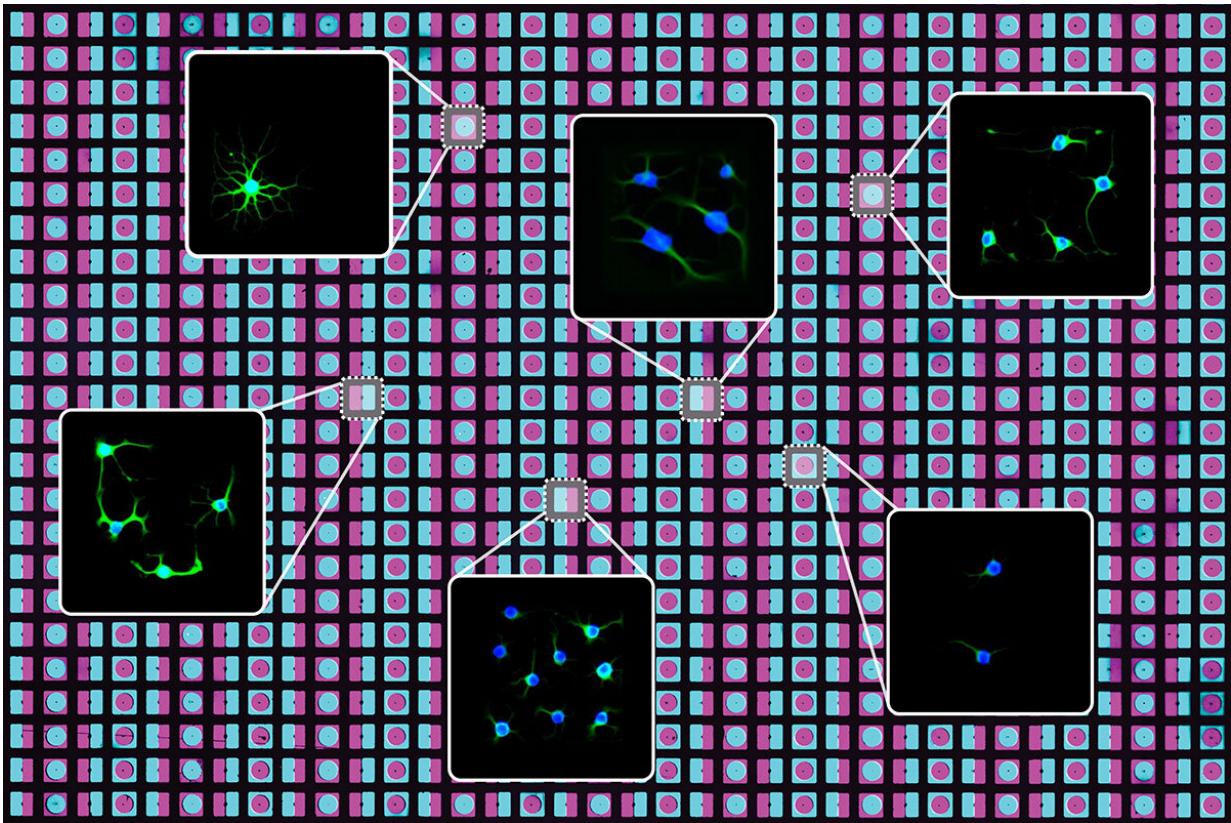


New technique 'prints' cells to create diverse biological environments

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University of California, Berkeley, researchers have created a new technique that utilizes photolithography and programmable DNA to rapidly "print" two-dimensional arrays of cells and proteins that mimic a wide variety of cellular environments in the body. Credit: (UC Berkeley graphic by Olivia Scheideler)

Like humans, cells can be easily influenced by peer pressure.

Take a [neural stem cell](#) in the brain: Whether this cell remains a stem cell or differentiates into a fully formed brain cell is ultimately determined by a complex set of molecular messages the cell receives from countless neighbors. Understanding these messages is key for scientists hoping to harness these stem [cells](#) to treat neurological conditions like Alzheimer's or Parkinson's.

With the help of photolithography and a creative use of programmable DNA, University of California, Berkeley, researchers have created a new technique that can rapidly "print" two-dimensional arrays of cells and proteins that mimic a wide variety of cellular environments in the body—be it the [brain tissue](#) surrounding a neural stem cell, the lining of the intestine or liver or the cellular configuration inside a tumor.

This technique could help scientists develop a better understanding of the complex cell-to-cell messaging that dictates a cell's final fate, from neural stem cell differentiating into a brain cell to a tumor cell with the potential to metastasize to an embryonic stem cell becoming an organ cell.

"What's really powerful about this platform is you can create in vitro tissues that capture the spatial organization of cells in the body, from the intestinal lining of your digestive tract to the arrangements of different cell types in the liver," said Olivia Scheideler, who completed the research as a graduate student at Berkeley. "I think you could apply this technique to recreate any tissue where you want to explore how cellular interactions contribute to tissue function."

In a paper appearing today in the journal *Science Advances*, Scheideler and her collaborators show that the new technique can be used to rapidly print intricate patterns of up to 10 different kinds cells or proteins onto a [flat surface](#).

"Essentially, what this technique allows us to do is pattern different kinds of conditions in one shot and in a high-throughput manner," said Lydia Sohn, Chancellor's Professor of Mechanical Engineering at UC Berkeley and senior author of the paper. "It provides a whole range of options for what you could study, because it's so flexible. You can pattern many different kinds of cells or proteins."

Caught on a DNA tether

In the new technique, each cell or [protein](#) is tethered to a substrate with a short string of DNA. While similar methods have been developed that attach tethered cells or proteins one by one, the new technique takes advantage of a patterning process called photolithography to attach, or print, each type of cell protein in one quick batch, greatly speeding up the process.

"It's like color laser printing, where you print one color and then print another," Sohn said.

Like photography, photolithography works by exposing a coated surface or substrate to a pattern of light, which initiates a chemical reaction that dissolves the coating in the illuminated areas, leaving a templated substrate. In the new technique, the substrate is then bathed in strands of single-sided DNA, whose ends have been chemically altered to firmly latch on where the coating has been dissolved.

Each single-sided DNA strand is programmed have a specific sequence of the nucleotides adenine (A), thymine (T), guanine (G) and cytosine (C). Single-sided DNA strands with the complementary nucleotide sequence are embedded or attached to cells or proteins of interest.

Finally, the surface is washed with a mixture of cells or proteins attached to the complementary strands of single-sided DNA, which bond with the

single-sided DNA already attached to the surface to form double-helix "tethers."

"All the cells and proteins attach exactly where they should be because of the DNA programming," Sohn said.

By repeating the process, up to 10 different kinds of cells or proteins can be tethered to the surface in an arbitrary pattern.

Conflicting messages

To demonstrate one of the many applications of the technique, Scheideler and co-author David Schaffer, Hubbard Howe Jr. Distinguished Professor of Biochemical Engineering at UC Berkeley, used the platform to study the chemical signaling that cues neural stem cells to differentiate into mature cells.

"Stem cells have programs embedded inside their DNA that tell them (to) either stay a stem cell or to differentiate into a mature cell," Schaffer said. "And they receive a lot of information about what to do and which programs to activate from the environment, from other cells around them. If we could learn how to make stem cells do our bidding, how to turn them into a particular cell type, then we could harness the stem cells to mass produce specialized cell types that were lost due to disease or injury."

Neural stem cells in the brain regularly receive conflicting messages from their neighbors about how they should behave, Scheideler said. One messenger, the FGF-2 protein, tells them to make more stem cells. The other, the ephrin-B2 protein, tells them to differentiate into a mature neuron.

Scheideler used the new technique to pattern neural stem cells onto

thousands of different arrays of the two proteins, FGF-2 and ephrin-B2, to see how the spatial organization of the two signals helps determine the cells' ultimate fate.

She found that many [stem cells](#) differentiated into mature neurons, even when they were primarily in contact with FGF-2, or "stay a stem cell," messengers. When she looked closer, however, she found that those cells that differentiated were more likely to have small, finger-like extensions, or "neurites," that touched the ephrin-B2 or "differentiate" messengers.

"The great thing about this patterning technology is you can easily replicate these little patterns hundreds or thousands of times across a slide," Schaffer said. "It is like running a thousand little independent experiments, each of which is a trial run to see how a stem cell listens to the cells around it. And then you can get very, very deep statistics about the various ways that it can be regulated."

More information: "Recapitulating complex biological signaling environments using a multiplexed, DNA-patterning approach" *Science Advances* (2020). DOI: [10.1126/sciadv.aay5696](https://doi.org/10.1126/sciadv.aay5696) , advances.sciencemag.org/content/6/12/eaay5696

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