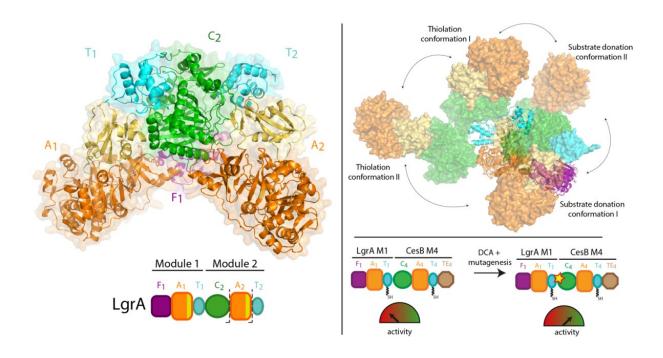


New understanding of antibiotic synthesis

November 11 2019, by Jason Clement



Credit: McGill University

Researchers at McGill University's Faculty of Medicine have made important strides in understanding the functioning of enzymes that play an integral role in the production of antibiotics and other therapeutics. Their findings are published in *Science*.

"Many of the medicines that we rely on today are natural products, made by the Earth's flora," explains Dr. Martin Schmeing, Associate Professor in the Department of Biochemistry at McGill and the study's senior



author. "This includes compounds made in microbes by massive enzymes called nonribosomal peptide synthetases, or NRPSs. NRPSs synthesize all sorts of antibiotics, which can kill dangerous fungi and bacteria, as well as compounds to help us fight off viral infections and cancers. For example, these compounds include viomycin, an antibiotic used for the treatment of multidrug-resistant tuberculosis; cyclosporin, which has been widely used as an immunosuppressant in organ transplants; and the familiar antibiotic penicillin."

In order to synthesize these drugs, NRPSs operate similar to a factory assembly line, consisting of a series of workstations. Each station, called a "module," has multi-step workflows and moving parts that allow it to add one building block component to the growing drug.

Understanding the assembly line's inner workings

Previous work by Dr. Schmeing and others has led to a solid understanding of how one module works. Now, using a technique called X-ray crystallography at the Canadian Light Source in Saskatchewan, and the Advanced Photon Source in Illinois, the team was able to take ultra-high resolution 3-D pictures of the NRPSs.

For the first time, they were able to make high quality observations about how an individual module relates to the bigger assembly line, by visualizing a two-module portion of the NRPSs that makes the antibiotic linear gramicidin (found in Polysporin treatments). The study found a surprising lack of synchronisation between modules at all points other than when they must coordinate to pass the intermediate from one workstation to the next. Additionally, they found that the modules don't line up in a straight line or other organized fashion, but instead can line up in many different relative positions. "This level of massive flexibility was not expected," notes Dr. Schmeing, who is also Director of McGill's Centre for Structural Biology. "The enzymes are performing



gymnastics."

Because the proteins are trapped in a crystal, care was taken to confirm that the results were representative of what happens in real life. Dr. Schmeing worked with his colleague, Dr. Alba Guarné, Professor in the Department of Biochemistry at McGill, to use complementary solution data, collected at the Advanced Light Source in Berkeley to validate the observations. "The structural biology community is very strong at McGill. We work together to help each other in collaborations, to obtain the biophysical equipment required for cutting-edge experiments, and to train our students" says Dr. Schmeing, noting that the experimentalists on the paper, Janice Reimer, Max Eivaskhani and Ingrid Harb, are all talented McGill graduate students. "The environment and colleagues at the McGill Centre for Structural Biology are important for the continued success of our labs."

Future implications for therapeutic design

The results could have implications for the production of new antibiotics and therapeutics in the long term. Since they were first discovered, scientists have been excited about the possibility of bioengineering NRPSs by mixing and matching workstations to produce designer compounds. "Our study shows that it should be possible to mix and match these modules, but that the bioengineered NRPSs must be modified at the points involved in passing the compound from one module to the next to get them to work well," explains Dr. Schmeing. "This is something we teamed up with Martin Weigt from the Sorbonne to do as a proof of principle in the paper, but which will need to be optimized for production of designer therapeutics."

"Structures of a dimodular nonribosomal peptide synthetase reveal conformational flexibility," by Reimer, J. M., Eivaskhani, M, Harb, I., Guarne, A, Weigt, M., Schmeing, T. M. was published in the journal



Science.

More information: Janice M. Reimer et al. Structures of a dimodular nonribosomal peptide synthetase reveal conformational flexibility, *Science* (2019). DOI: 10.1126/science.aaw4388

Provided by McGill University

Citation: New understanding of antibiotic synthesis (2019, November 11) retrieved 3 April 2024 from https://phys.org/news/2019-11-antibiotic-synthesis.html

This document is subject to copyright. Apart from any fair dealing for the purpose of private study or research, no part may be reproduced without the written permission. The content is provided for information purposes only.