

Exploring the interaction of polystyrene nanoplastics and blood plasma proteins

August 22 2019



Proposed structural model of PS-,/HSA complexes in pH 7.4 (top) and pH 5.0 (bottom). Credit: Australian Nuclear Science and Technology Organisation



(ANSTO)

Collaborative research at ANSTO led by Mr Shinji Kihara and A/Prof. Duncan McGillivray of The MacDiarmid Institute, New Zealand with ANSTO's Dr. Jitendra Mata, scientists from the University of Auckland and A/Prof Ingo Köper from Flinders University, SA, is contributing to a better understanding of how nanoplastics interact with blood plasma proteins and other biological molecules within the body.

The motivation for this study, which was recently published in *Bioconjugate Chemistry*, arose from concerns regarding the increasing quantities of plastic waste in the environment.

In nature, these plastics undergo physical and chemical breakdown processes to form <u>minute particles</u> on the micro- and nanometer scale.

Studies on the toxicology of engineered <u>nanoparticles</u> have suggested that these particles gain easy access and mobility within the body, often side-stepping important biological barriers and defence mechanisms against foreign bodies. Unlike the engineered nanoparticles used in biomedical applications, however, the potential effects and interactions of these nanoplastics are not well-understood.

The toxicity of nanoparticles is directly linked to its physical and chemical properties. When nanoparticles enter the body, they are surrounded by layers of proteins, which is known as a "corona." Loosely-bound proteins form a "soft corona," while tightly bound proteins form a "hard corona."

With a model system of polystyrene nanoparticles and human serum albumin (HSA) protein, the investigators used a range of techniques to



determine the size, composition and geometry of the polystyrene nanoparticle-protein corona complex.

HSA was selected due to its natural abundance, while positively and negatively charged nanoparticles, of two different sizes, were used to assess how <u>particle size</u> influenced the formation of the corona in solution under different pH conditions. The investigators used <u>small</u> <u>angle neutron scattering</u> (SANS) with contrast matching on the Bilby instrument (with Dr. Andrew Whitten) to determine the diameter of the nanoparticle and characterize its distinct structural features.

"Our facilities are quite unique in that we can explore the interaction between protein and nanoparticle at length scales from one nanometer to 10 microns, which is very difficult to do with other techniques," said Dr. Jitendra Mata, instrument scientist and co-author on the paper.

"Contrast matching allows you to see two components together, such as nanoparticles and protein corona, or we can mask out one of the components of interest. We were able to determine if there was a strong or weak interaction with the protein or if there was any shape change in the <u>protein</u>," he added.

The study found that both particle size and pH played a role in determining the nature of the corona. Larger particles favored the formation of a soft corona, with the hard corona being completely absent in some cases. HSA actively participated in the formation of these complexes, dotting the surface of negatively-charged nanoparticles.

Additionally, they discovered that the interaction between the soft corona and the nanoparticle surface was governed by a subtle balance of electrostatic forces.

The ongoing research seeks the usage of other neutron instruments



including Kookaburra USANS and Platypus neutron reflectometry to understand how these complex corona/nanoplastic would interact with other biological entities such as cellular membranes.

The researchers anticipate that these findings will have implications on further research on nanoparticle toxicity, by providing a clearer picture of the interactions of nanoparticles with biomolecules.

More information: Shinji Kihara et al. Soft and Hard Interactions between Polystyrene Nanoplastics and Human Serum Albumin Protein Corona, *Bioconjugate Chemistry* (2019). <u>DOI:</u> <u>10.1021/acs.bioconjchem.9b00015</u>

Provided by Australian Nuclear Science and Technology Organisation (ANSTO)

Citation: Exploring the interaction of polystyrene nanoplastics and blood plasma proteins (2019, August 22) retrieved 27 June 2024 from <u>https://phys.org/news/2019-08-exploring-interaction-polystyrene-nanoplastics-blood.html</u>

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