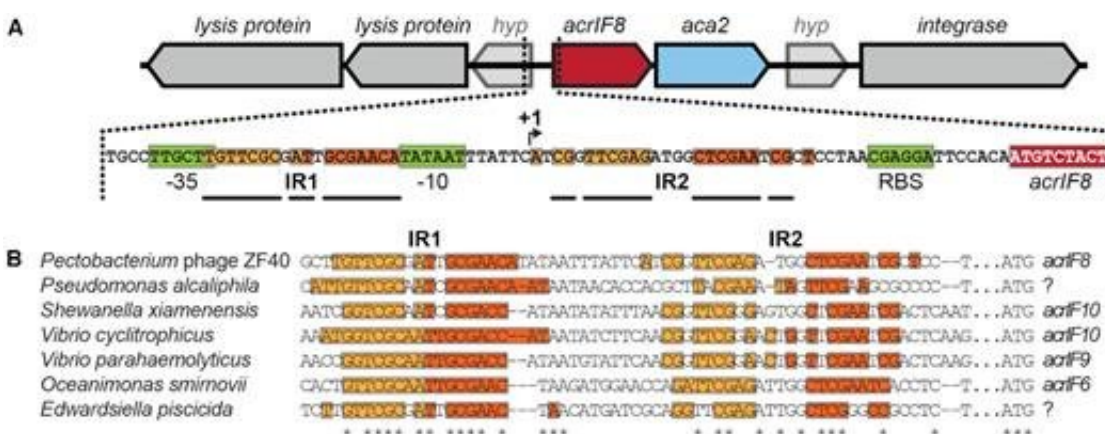


Bacteria-attacking phages could provide clues to antibiotic resistance

August 29 2019



Inverted repeat pairs are conserved in *acr*–*aca2* operon promoters. (A) Genomic context of the *acrIF8*–*aca2* locus of phage ZF40 with inverted repeat pairs (shades of orange; bold bars illustrate symmetry and distance between each half-site of the respective repeat). Predicted regulatory sequences (–35 and –10 sites, and ribosome binding site (RBS)) in green, predicted transcription start site (+1) indicated by an arrow. (B) Alignment (24) of *acr*–*aca2* operon promoters with inverted repeats displayed as in (A). Invariant residues are indicated by an asterisk and the *acr* genes encoded downstream are given where known. Question marks indicate genes that have no matches among known *acr* genes.

Is there a solution to bacteria becoming resistant to antibiotics? One answer may be found by studying the world's largest and most brutal army, new University of Otago microbiology research shows.

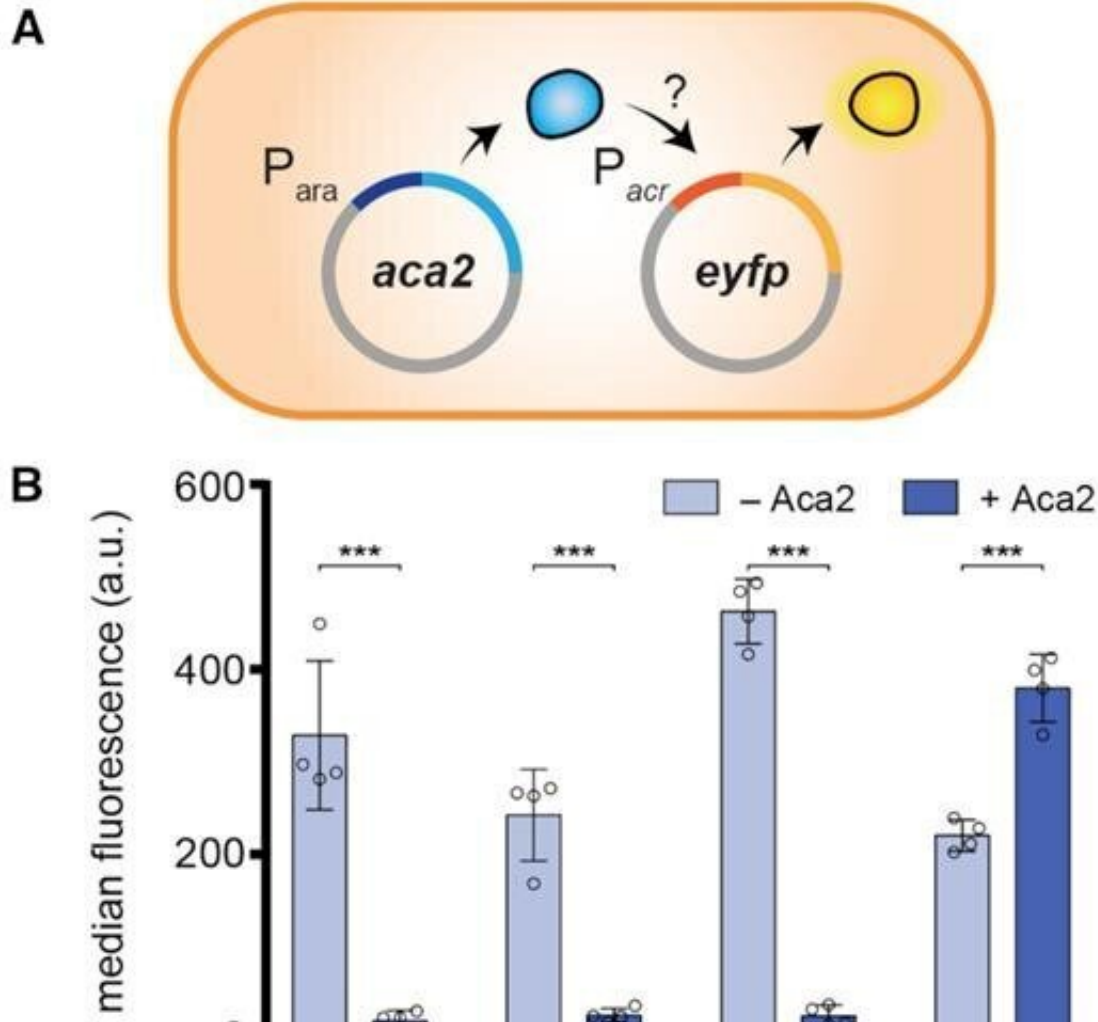
In fact that army—bacteria-attacking viruses known as phages—outnumber bacteria nearly ten-fold, making them the most abundant biological entity on earth. They are intricately adapted to invading, weakening and controlling their targets, then keeping them alive long enough to feed off them and use them to breed.

To defend themselves from the phage invasion, bacteria have developed "CRISPR" defense systems—immune systems within the bacteria. But the phages have their own weapon, called "anti-CRISPR," which blocks these bacterial defenses.

The Otago study's lead author, Ph.D. student Nils Birkholz, of the University's Department of Microbiology and Immunology, says the new research shows phages use a special protein to control the production of the anti-CRISPR when they invade a bacteria.

Initially they rapidly ramp up anti-CRISPR production, conquering the bacteria's defense system. Then, with the bacteria beaten, the phages use the protein to switch off anti-CRISPR production, ensuring the [bacteria](#) survives—conquered, but alive.

That makes sense, Mr Birkholz says, as phages, like all viruses, are not just conquerors but hijackers that reproduce inside living hosts.



Aca2 represses the *acrIF8-aca2* operon. (A) Schematic of the plasmid setup for the assay to measure autoregulation of the *acrIF8-aca2* promoter by Aca2 in a Pca ZF40⁻ host (Pca RC5297). (B) Activity of *acrIF8-aca2* promoter variants in Pca ZF40⁻ in the presence and absence of Aca2, determined as the median eYFP fluorescence. The IR sites were mutated as indicated; sc: scrambled or Δ: deleted. (C) Schematic of the *acrIF8-aca2* promoter assay in the ZF40⁺ strain (Pca lysogen ZM1). (D) Activity of *acrIF8-aca2* promoter variants in the Pca ZF40⁺ strain, determined as the median eYFP fluorescence. The Pca ZF40⁻ control strain lacks *aca2* and in the Pca ZF40⁺ strain *aca2* is expressed natively from the ZF40 prophage. (E) Activity of *acrIF8-aca2* promoter variants in the Pca ZF40⁻ strain in the presence of different concentrations of arabinose to induce *aca2* expression. In (B) and (D), data are presented as the mean \pm standard deviation of four biological replicates and statistical significance was tested by two-tailed unpaired t-tests (* P 0.05).

"We know from previous research that too much anti-CRISPR production can be bad for the cell and an important question was how anti-CRISPR abundance is controlled. This has now been answered by our research.

"Our results suggest that right after infection, the [phage](#) produces a large enough anti-CRISPR quantity to inhibit bacterial defense, but then turns down production to avoid any negative side effects.

"So this protein ensures that, once the virus has beaten its host it keeps it alive and devotes its resources to its own reproduction."

That is, until the phages finish reproducing, at which point they explode out of the cells, killing them, before moving on to infect other cells.

The results emphasize the "delicate balancing act" phages need to perform to subdue their hosts, Mr Birkholz says. And as gruesome as it sounds, the research could lead to very "real world" outcomes.

"Phages have a strong impact on our lives in both negative and positive ways. Particularly in this era of increasing antibiotic resistance, phages are being considered as a means to treat bacterial infections. So these new details provide information that might help us choose, or design, more effective antimicrobial phages."

The [research paper](#), called "The autoregulator Aca2 mediates anti-CRISPR repression" and published in Nucleic Acids Research this month, was written by Nils Birkholz, Robert D. Fagerlund, Leah M. Smith, Simon A. Jackson and Peter C. Fineran.

More information: Nils Birkholz et al. The autoregulator Aca2 mediates anti-CRISPR repression, *Nucleic Acids Research* (2019). [DOI: 10.1093/nar/gkz721](https://doi.org/10.1093/nar/gkz721)

Provided by University of Otago

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