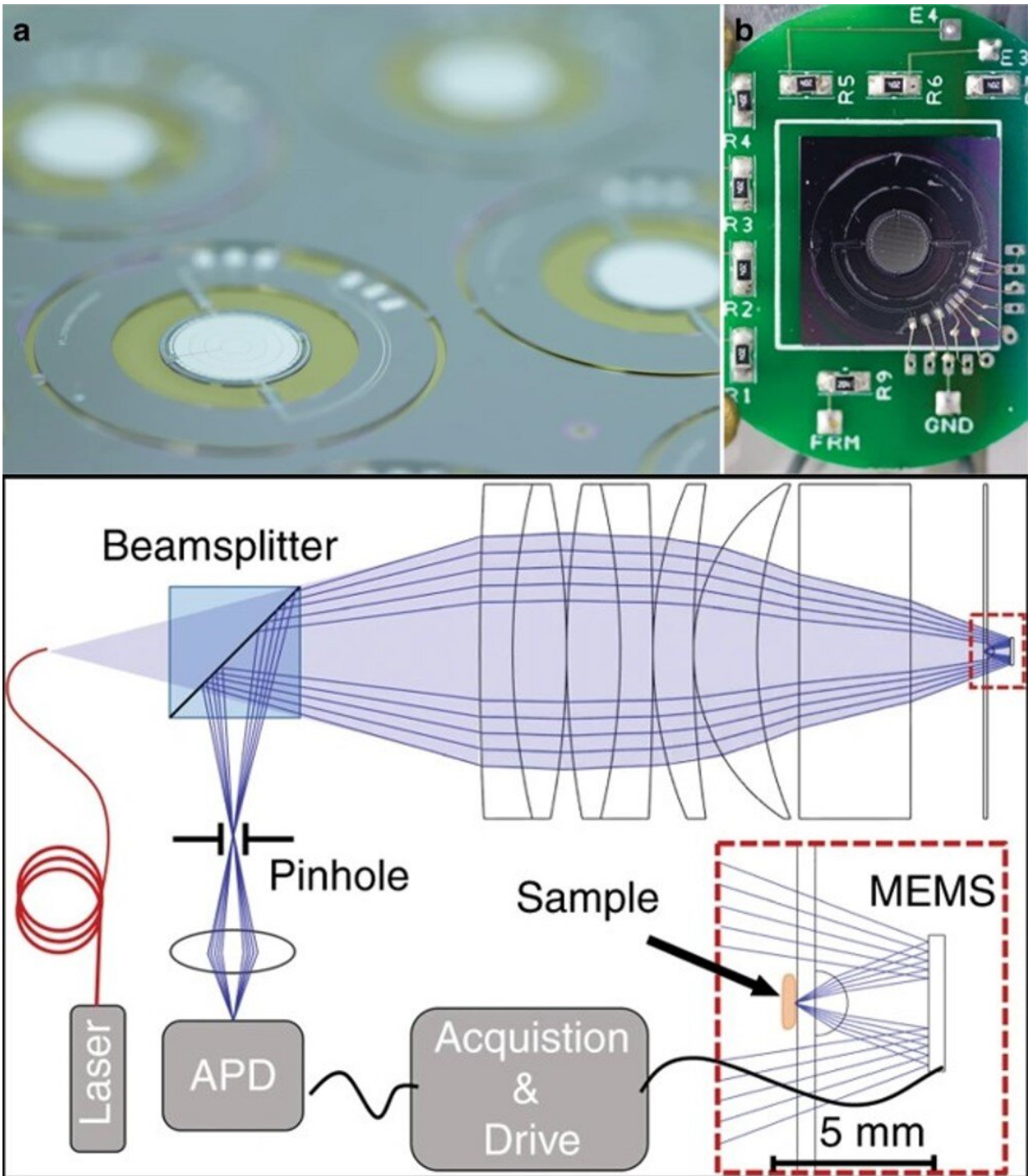


MEMS-in-the-lens architecture for laser scanning microscopy

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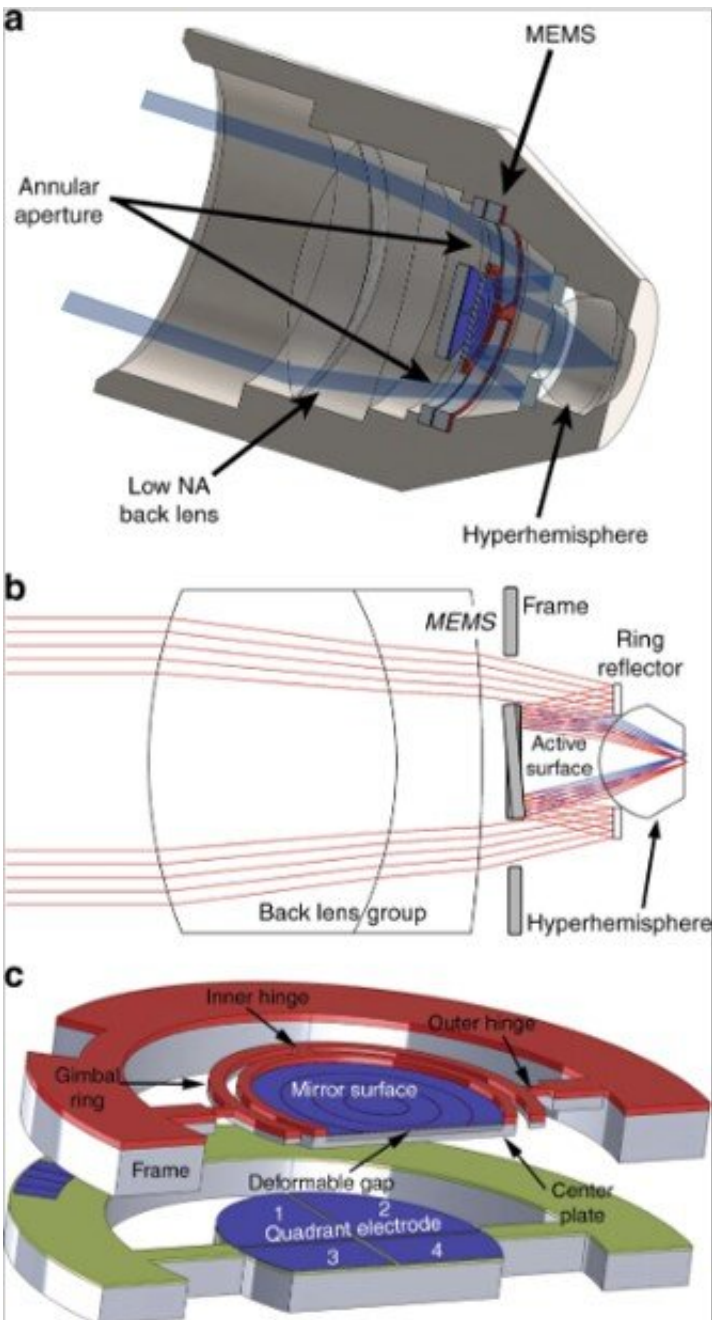
TOP: Inbuilt 3D MEMS scan mirror. (a) Devices on wafer after the release process. (b) The MEMS scanner after wire bonding to support PCB. BOTTOM: Schematic diagram of the confocal imaging setup. A magnified view of the MEMS scanner, hyperhemisphere, and sample stage. Credit: Light: Science & Applications, doi: 10.1038/s41377-019-0167-5

Laser-scanning microscopes can be miniaturized [to image microenvironments](#) in vivo via inclusion inside optical micromechanical system (MEMS) devices to replace the existing larger components. Multifunctional active optical devices are emerging components that support miniaturization for diffraction-limited performance with simpler optical system designs in optical devices. In a recent study, Tianbo Liu and a team of researchers in the departments of Electrical and Computer Engineering and Dermatology in the U.S. proposed a [catadioptric](#) (allowing both light reflection and refraction) microscope objective lens, featuring an integrated MEMS device to perform biaxial scanning, axial focus adjustment and control [spherical aberration](#).

The [materials scientists](#) included a reflective MEMS scanner into the MEMS-in-the-lens architecture to support high-numerical-aperture (NA) imaging that gathers light across a wider range of angles to generate images. Liu et al. implemented the MEMS-in-the-lens architecture by including the scan mirror into the objective lens, where the beam axis was normal to the mirror surface without the need for a [beam splitter](#) to separate the incident and reflected beam. They demonstrated the optical performance of the [catadioptric system](#) (an optical system that allows both light refraction and reflection with minimal aberration) by imaging hard and soft targets using a confocal microscope based on the new objective lens design. The improved imaging technique will allow advanced diagnosis of medical conditions. The results of the study are now published on *Light: Science & Applications*.

Unprepared and uncleared organs in live animals can be imaged in vivo using [scanning laser confocal and multiphoton microscopy](#) techniques. Technical advances have facilitated [bench top imaging of small animal models](#) such as mice, with suitable medical applications also emerging in dermatology clinics to [noninvasively examine optical skin biopsies](#).

However, conventional laser scanning microscopes are large and limit both medical and live animal imaging procedures. To access the [human body](#) and image ambulatory animals, scientists must therefore miniaturize these instruments.

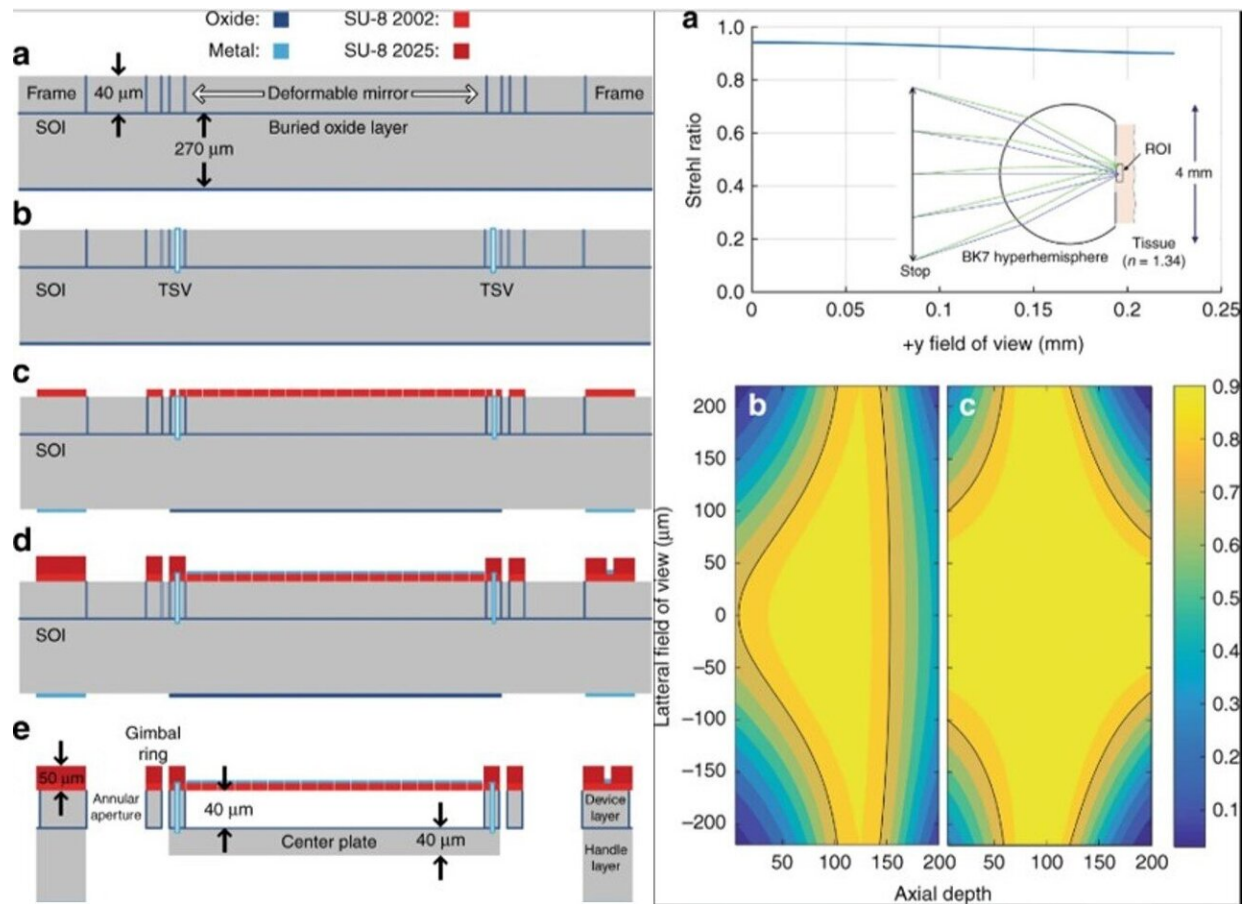


MEMS-in-the-lens architecture. (a) A cross-sectional view of the miniaturized

confocal microscope with a new objective lens that incorporates a MEMS 3D scanner. (b) An illustration of the light path through the annular aperture and the beam scan of the MEMS device. (c) A model of the MEMS 3D scanner. A gimbal platform is bonded to a set of quadrant electrodes. Credit: Light: Science & Applications, doi: 10.1038/s41377-019-0167-5

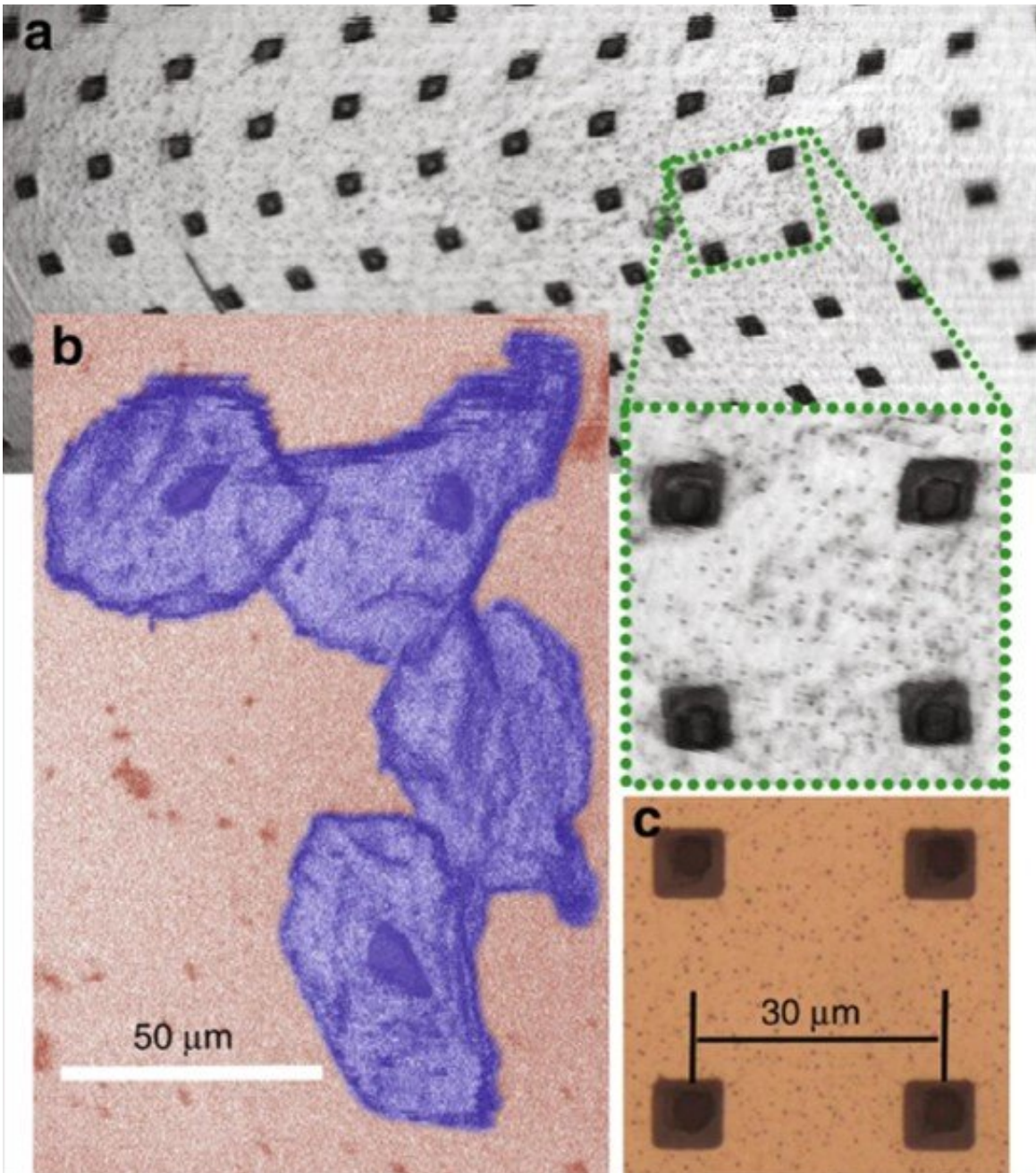
Miniaturized scanning mechanisms with smaller instruments such as micromechanical system devices can replace existing bulky mechanisms required to scan and focus the beam for hitherto improbable applications. For example, scientists were able to mount a MEMS-scanned miniaturized two-photon microscope weighing only 2.15 g [on the head of a freely moving mouse](#) for brain imaging. The devices have also facilitated laser scanning microscopy to be adapted in [endoscopic platforms](#) and during MEMS-based [optical biopsy experiments](#) to detect cancer in vivo. Alongside its smaller footprint, a MEMS scanner contributes to miniaturization by [combining multiple degrees of freedom](#) during its production alongside its optical architecture.

In the present work, Liu et al. explored a new optical architecture for a miniature, high-NA scanning laser microscope with a 3-D MEMS scanner within the objective lens. They illustrated the optical layout of the MEMS-in-the-lens to fabricate the device and operate it in vivo. The scientists engineered the MEMS 3-D scan mirror by successfully reproducing [a method previously introduced](#) by the same group. For in vivo microscopy, they operated the [hyperhemisphere](#) (that offers a broader field of view) in contact with tissue containing a [variable index of refraction](#) ranging from 1.3 to 1.4. Based on the parameters, the scientists simulated the imaging performance of the setup. They concluded hyperhemisphere of BK-7 glass to be effective as a front lens element for a tissue microscope with an active 3-D MEMS scanner deployed at the simulated aperture.



LEFT: Gimbal wafer fabrication schematic diagram. (a) Etching, oxidizing, and patterning the vertical etch stops. (b) Creating TSVs. (c) Spin-coating and patterning the deformable membrane. (d) Depositing and patterning the top-side metal and spin-coating and patterning the SU-8 hinges. (e) Fully released gimbal platform. RIGHT: Simulated imaging performance. (a) A Zemax simulation of a 2-mm-radius BK-7 hyperhemisphere that is in contact with tissue. The aperture stop is 2.5 mm to the left of the lens, with $NA = 0.7$, with a plot of the Strehl ratio vs. the lateral field, simulated for a depth of $125 \mu\text{m}$. (b), (c) A contour plot of the Strehl ratio over a 2D axial cross-section of the 3D field of view. The black line represents the contour for $S = 0.8$. b Without depth-dependent adjustment of the spherical aberration. (c) With depth-dependent adjustment of the spherical aberration. Credit: Light: Science & Applications, doi: 10.1038/s41377-019-0167-5

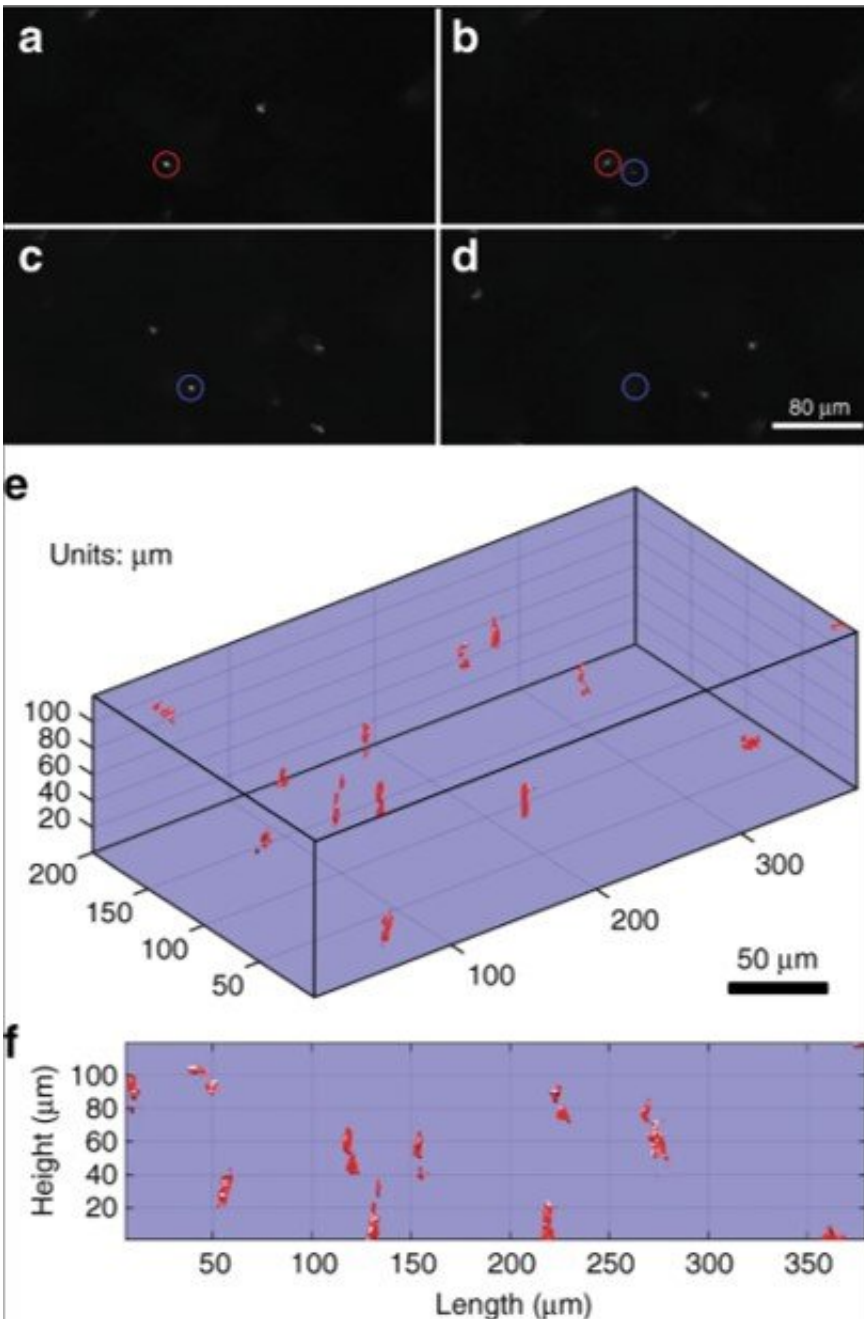
To demonstrate confocal imaging, the scientists used a bench-top mockup of the objective lens with an integrated 3-D MEMS mirror. Liu et al. attached the mirror onto the sample stage using a thin-layer of water-based ultrasound gel. As an example, they introduced samples of human cheek cells ($\sim 80 \mu\text{m}$) on to the sample stage, and captured their images using the microscope thereafter. During imaging, the scientists used a 633 nm helium neon laser for illumination. They then attached the sample of interest on the glass wafer opposite the hyperhemisphere lens. Liu et al. included a 50/50 beam splitter between the optical fiber and compound lens element to separate the reflected light, and a $10 \mu\text{m}$ pinhole to spatially filter the reflected light.



Experimental imaging results. a A confocal image of the surface of a prototype three-dimensional scanner. A subsection of the image is digitally enlarged to show details. b A confocal image of human cheek cells (with false coloring). The nucleus and cell membranes are clearly visible. c A digitally cropped brightfield epi-illumination microscope image of the surface of a similar prototype mirror that was recorded using a $\times 50$ objective lens ($NA = 0.8$). Credit: Light: Science & Applications, doi: 10.1038/s41377-019-0167-5

The MEMS [confocal microscope](#) also allowed imaging beneath the surface of the sample and Liu et al. demonstrated this via imaging a sample of interest. For the sample, they suspended 6 μm polystyrene microbeads in an ultrasound transmission gel then followed up the imaging process with volumetric reconstruction of the images to better illustrate confocal sectioning at different focal planes. Although the images were well resolved, the scientists observed that the 3-D profiles of the beads were neither uniform nor symmetric requiring further optimization of the technique.

The 3-D MEMS mirror developed provided complete scanning and focus control for the instrument, alongside electronic control of the spherical aberration. The new work showed improved resolution compared with previously described 3-D MEMS mirrors, to allow its inclusion in a compact MEMS-in-the-lens system.



3D imaging demonstration. (a–d) Confocal sectioning of 6- μm -diameter polystyrene beads suspended in ultrasound gel. Two beads have been circled using different colors to show their focus change from frame to frame. (e) A volumetric reconstruction from the images recorded at each focal plane. (f) A first-angle projection through the volumetric rendering to better illustrate the confocal sectioning at different focal planes. Credit: Light: Science & Applications, doi: 10.1038/s41377-019-0167-5

In this way, Tianbo Liu and co-workers proposed and developed a catadioptric MEMS-in-the-lens [microscope](#) objective lens and integrated a MEMS 3-D scanner to perform biaxial scanning with controlled spherical aberration during imaging applications. Liu et al. simulated the development of the proposed instrument architecture to indicate considerable promise for future, miniaturized and high-NA laser scanning microscopes for in vivo imaging applications.

More information: Tianbo Liu et al. MEMS-in-the-lens architecture for a miniature high-NA laser scanning microscope, *Light: Science & Applications* (2019). [DOI: 10.1038/s41377-019-0167-5](https://doi.org/10.1038/s41377-019-0167-5)

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