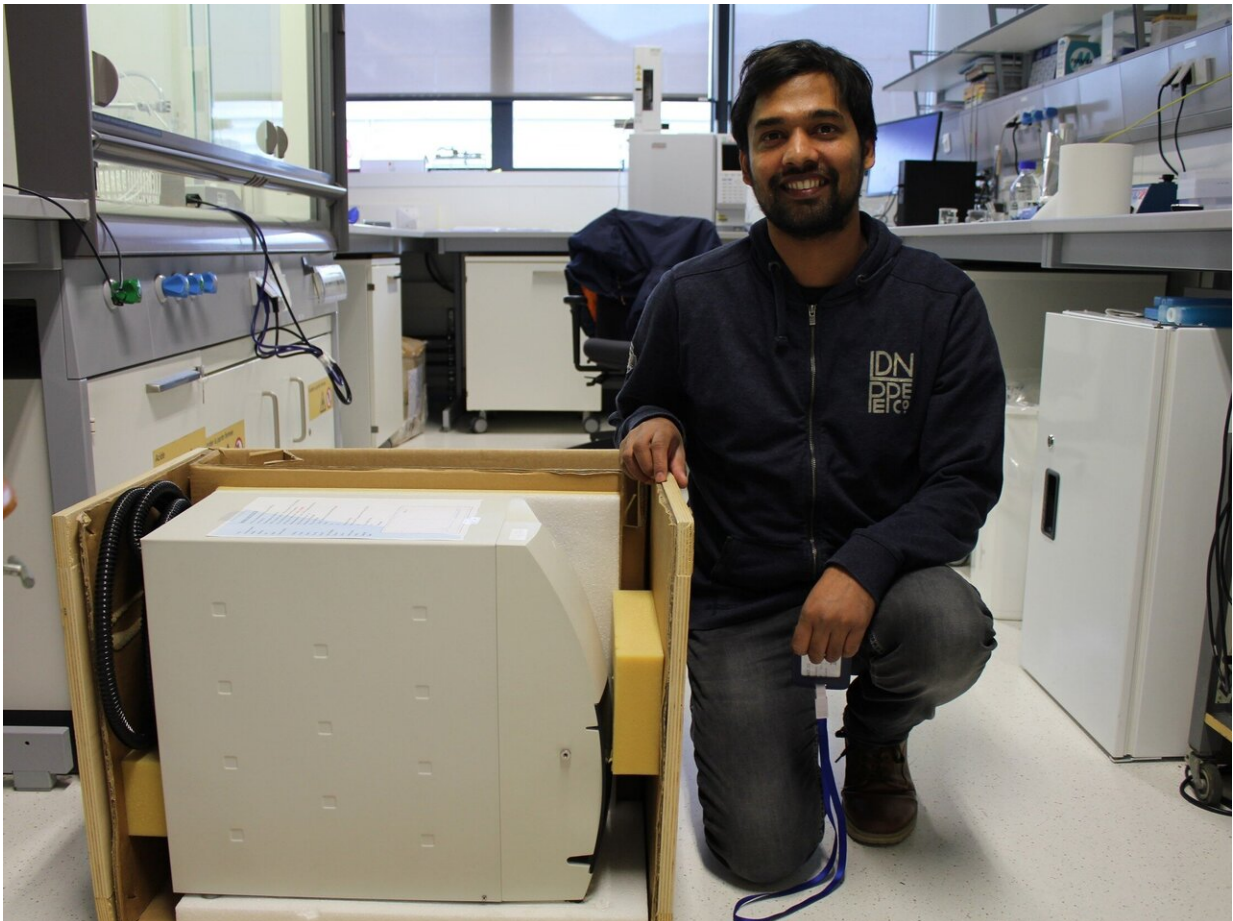


# Preparing protocols for making deuterated biomolecules

June 26 2019

---



Credit: SINE2020

Neutron techniques are good for studying light atoms like

hydrogen—great for biological molecules that contain large numbers of them. Neutrons are particularly sensitive to isotopic substitution of hydrogen ( $1\text{H}$ ) with deuterium ( $2\text{H}$ ) and this allows contrast techniques to be used to study molecules in detail. For this, users need to prepare deuterated versions of the biological molecules that they want to study. However, these per-deuterated molecules are rarely available from commercial suppliers because the market for them is of limited value. Here SINE2020 aims to fill the gap.

Task 5.2 of the Chemical Deuteration work package is to setup procedures for extraction, purification and analysis of small-deuterated biomolecules. These molecules will then be available to users of neutron techniques and via the DEUNET platform partners. The main biomolecules involved are phospholipids, sterols (e.g. cholesterol) and sphingolipids—all types of molecules that can be found in [biological membranes](#) and therefore have important applications in pharmaceuticals. Lipids also have roles in the cosmetics, food and nanotechnology sectors so the ability to study them using neutrons would be invaluable to many researchers and industrial companies.

Krishna Batchu, with Giovanna Fragneto, at ILL is carrying out the research for SINE2020. Work has concentrated on various phospholipids that mainly include phosphatidylethanolamine (PE), phosphatidylcholine (PC) and phosphatidylserine (PS). They are prepared using cell cultures of the yeast strain *Pichia pastoris* as it works well for lipid extraction and grows successfully in deuterated media.

Phospholipids are amphipathic molecules that consist of a water-loving head group and two water-hating "legs". The "legs" are chains of [carbon atoms](#) attached to hydrogen (or deuterium atoms). Their compositions in the cellular membranes are enormously complex with membranes typically comprising several thousand chemically distinct molecular species of lipids. The chains within can vary in length, usually from

between 6 and 24 carbon atoms, and contain between 0 and 12 double bonds between the carbon atoms—the so-called saturation level. The more [double bonds](#), the more unsaturated the chain.

Phospholipid homeostasis in a biological system is maintained via three key cellular processes: 1) Biosynthesis 2) Remodeling and 3) Degradation. In the [yeast cells](#) (in the culture) exists a dedicated molecular machinery for [fatty acid metabolism](#) causing both elongation and unsaturation of the chain, catalyzed by elongases and desaturases respectively, during the production process. Once biosynthesized, their incorporation into various phospholipids is catalyzed by a number of acyltransferases thus leading to the existence of a larger repertoire of individual molecular species. The challenge is that with so many variables the number of types of phospholipids produced is vast.

Part of the project has studied how growth conditions affect the type of lipids produced. No significant differences have been found that suggest harvesting time affects the lipid profile but there is some indication that carbon source has an effect and that more unsaturation occurs if growth occurs at [lower temperatures](#).

Once a cell culture has grown, the lipids need to be extracted, separated and purified. This has been achieved, for both hydrogenated and deuterated PC, PS and PE [molecules](#), using a two-step process: a solid phase extraction followed by purification via a normal phase column coupled to a HPLC system. Hydrogenated versions have been prepared in order to compare them with the deuterated species and for initially testing processes to avoid wasting expensive deuterium.

Now the challenge is to purify individual molecular species, identify them and analyse them using a Mass Spectrometric approach. The ultimate aim is to carefully document all the procedures and protocols for producing these deuterated biomolecules for future users.

Provided by CORDIS

Citation: Preparing protocols for making deuterated biomolecules (2019, June 26) retrieved 24 April 2024 from <https://phys.org/news/2019-06-protocols-deuterated-biomolecules.html>

This document is subject to copyright. Apart from any fair dealing for the purpose of private study or research, no part may be reproduced without the written permission. The content is provided for information purposes only.