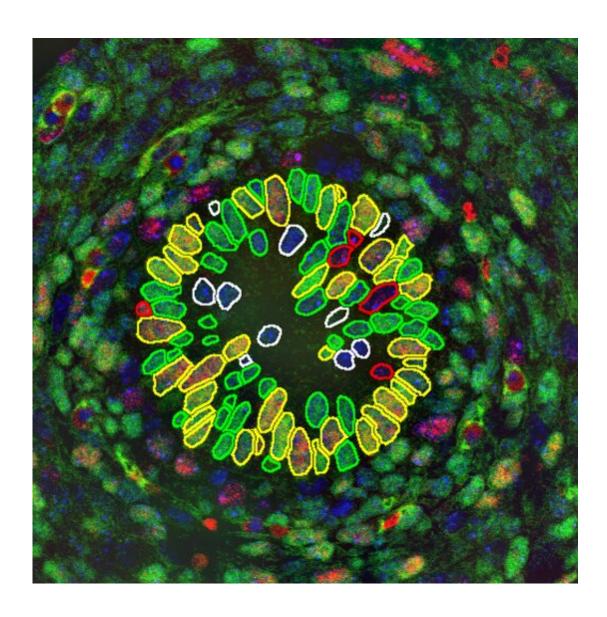


The deep learning dive: how cells regulate division

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Photomicrograph of the embryonic mouse intestine showing expression of E2F3A (green) and E2F8 (red). A nuclear segmentation mask is overlaid on the image. Credit: The images were acquired by Maria C. Cuitiño and the nuclear



segmentation mask generated by Thierry Pécot.

Combining tissue imaging and artificial intelligence, Hollings Cancer Center researchers at the Medical University of South Carolina probed deeper into how cell division cycles are regulated, in this study released online in the May 2019 issue of *Cell Reports*.

Addressing a long unanswered question of how <u>cell division</u> is controlled during development and normal maintenance in multicellular organisms, Maria Cuitino, Ph.D., and colleagues used animal models and applied deep learning tools to measure <u>protein levels</u> and expose cellular mechanisms that previously could only be estimated by <u>cell culture</u> <u>systems</u>. The findings begin to identify the possible early events associated with uncontrolled cell division, a key step in the early progression to cancer.

Past research revealed which molecules direct <u>cells</u> to divide or not divide, but this left many scientific gaps. Since bodies are made up of many different types of cells that come together to form complex organs, studying the whole body at once can be very complicated, but also can be very exciting and revealing, says Gustavo Leone, Ph.D., Hollings Cancer Center director and corresponding author on the study.

Previous studies that looked at <u>individual cells</u> in in vitro cell culture systems provided basic answers to the biology going on inside the cell but missed the interactions that occur when all of the cells are working together to form organs. Leone and his team's findings confirmed at least 80% of the previous knowledge derived from cell culture systems and addressed new important questions that needed to be answered.

"Not knowing when and where "on and off" switches for cell division



are expressed is like having paint with no canvas. Now we have the canvas, and thus the cellular context, for how these proteins behave within cells in the body," Leone says.

The five-year project probed "when" and "where" a critical family of transcription factor proteins (E2F family) is expressed in mammalian cells. Mammals have at least nine different E2F transcription factors that have either activation (on) or repressive (off) functions. All units within cells must work properly to make a functioning organ. "Our DNA provides the code to make the multiple proteins, which are the functional units of our cells. Transcription is the first biological process that makes proteins from DNA, and transcription factors are the on and off switches for this process," Leone explains.





Dr. Maria C. Cuitiño is a researcher at MUSC Hollings Cancer Center. Credit: MUSC Hollings Cancer Center

Cancer is one of the most common diseases that occurs when cells multiply in an uncontrolled manner. Understanding the on and off switches, the transcriptions factors, is essential to understand disease processes like cancer.

Instead of studying cell division regulation in cultured cells (or in vitro), the researchers used a whole-organism approach. Two major discoveries were made in this study. The most surprising discovery during this work was that the same E2F family of proteins are organized into two modules that work similarly in all cell types and organs in our bodies. "So it appears that a universal mechanism has evolved to control cell divisions, regardless of the diversity of cell types existing in our bodies," he says.

The second discovery was the development of tools that allow this level of precision in the analysis of proteins in complex tissues. Critical to the artificial tools developed in this study was Hollings Cancer Center researcher Thierry Pecot, Ph.D. "To be able to develop the tools that can detect the infrequent presence of transcription factors in every cell and quantify them is both clinically and biologically relevant," Pecot says.

The Leone lab harnessed the power of <u>artificial intelligence</u> to quantify transcription factors across numerous cells in mouse tissues. While deep learning-based tools have been used for medical imaging before, it was not advanced enough to recognize individual cells in microscopic images within tissues/organs. The technology used by the lab is similar to how self-driving cars recognize objects on the street and allowed the identification of individual cells.



Often the exact clinical relevance of a particular biological discovery is not clear for decades. Currently other papers and emerging data from this group show hints of the role of E2F in cancer. The Leone laboratory uncovered that three transcription factors (E2F3A, E2F8, E2F4) work together to control cell division, while two others combine to stop cell division. These findings provide the strong foundation for further studies to understand these complex mechanisms.

This National Institutes of Health-funded research was driven by an international team of researchers with broad training and expertise, with the lead authors originating from Argentina, France, and China, which allowed for a great breadth of thought processes and determination, Leone says. "Solid elegant research leads to more questions, and that is certainly true in this comprehensive in-depth study of mammal E2F transcription factors."

The study prompts important questions for future research, says Leone. Using advanced technology, "we uncovered when and where the on and off switch modules for cell division are expressed in intact organisms". However, we do not know why there are multiple on and off switches, and whether these switches have redundant roles."

More information: Maria C. Cuitiño et al, Two Distinct E2F Transcriptional Modules Drive Cell Cycles and Differentiation, *Cell Reports* (2019). DOI: 10.1016/j.celrep.2019.05.004

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