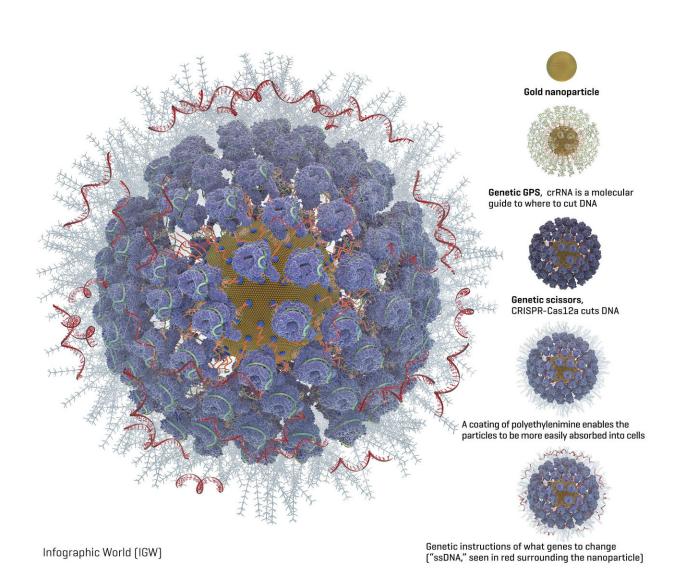


Could gold be the key to making gene therapy for HIV, blood disorders more accessible?

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Graphic of a fully loaded gold nanoparticle with CRISPR and other gene editing tools. Credit: the Adair lab at Fred Hutch.



Scientists at Fred Hutchinson Cancer Research Center took a step toward making gene therapy more practical by simplifying the way geneediting instructions are delivered to cells. Using a gold nanoparticle instead of an inactivated virus, they safely delivered gene-editing tools in lab models of HIV and inherited blood disorders, as reported May 27 in *Nature Materials*.

It's the first time that a <u>gold</u> nanoparticle loaded with CRISPR has been used to edit <u>genes</u> in a rare but powerful subset of blood <u>stem cells</u>, the source of all blood <u>cells</u>. The CRISPR-carrying gold nanoparticle led to successful gene editing in blood stem cells with no toxic effects.

"As gene therapies make their way through clinical trials and become available to patients, we need a more practical approach," said senior author Dr. Jennifer Adair, an assistant member of the Clinical Research Division at Fred Hutch, adding that current methods of performing gene therapy are inaccessible to millions of people around the world. "I wanted to find something simpler, something that would passively deliver gene editing to blood stem cells."

While CRISPR has made it faster and easier to precisely deliver <u>genetic</u> <u>modifications</u> to the genome, it still has challenges. Getting cells to accept CRISPR gene-editing tools involves a small electric shock that can damage and even kill the cells. And if precise gene edits are required, then additional molecules must be engineered to deliver them —adding cost and time.

Gold nanoparticles are a promising alternative because the surface of these tiny spheres (around 1 billionth the size of a grain of table salt) allows other molecules to easily stick to them and stay adhered.





Drs. Jen Adair (left) and Reza Shahbazi examine a test batch of gold nanoparticles at their lab at Fred Hutch. The solution is red because when gold ions clump together to form nanoparticles, electrons on the surface start oscillating and reflecting the light in different wavelengths. Then when light strikes the gold particles, electrons reflect the light differently. Credit: Robert Hood, Fred Hutch

"We engineered the <u>gold nanoparticles</u> to quickly cross the cell membrane, dodge cell organelles that seek to destroy them and go right to the cell nucleus to edit genes," said Dr. Reza Shahbazi, a Fred Hutch postdoctoral researcher who has worked with gold nanoparticles for drug and gene delivery for seven years.



Shahbazi made the gold particles from laboratory-grade gold that is purified and comes as a liquid in a small lab bottle. He mixed the purified gold into a solution that causes the individual gold ions to form tiny particles, which the researchers then measured for size. They found that a particular size—19 nanometers wide—was the best for being big and sticky enough to add gene-editing materials to the surface of the particles, while still being small enough for cells to absorb them.

Packed onto the gold particles, the Fred Hutch team added these geneediting components (diagram available):

- A type of molecular guide called crRNA acts as a genetic GPS to show the CRISPR complex where in the genome to make the cut.
- CRISPR nuclease protein, often called "genetic scissors," makes the cut in the DNA. The CRISPR nuclease protein most often used is Cas9. But the Fred Hutch researchers also studied Cas12a (formerly called Cpf1) because Cas12a makes a staggered cut in DNA. The researchers hoped this would allow the cells to more efficiently repair the cut and while so doing embed the new genetic instructions into the cell. Another advantage of Cas12a over Cas9 is that it only requires one molecular guide, which is important because of space constraints on the nanoparticles. Cas9 requires two molecular guides.
- Instructions for what genetic changes to make ("ssDNA"). The Fred Hutch team chose two inherited genetic changes that bestow protection from disease: CCR5, which protects against HIV, and gamma hemoglobin, which protects against <u>blood disorders</u> such as sickle cell disease and thalassemia.
- A coating of a polyethylenimine swarms the surface of the particles to give them a more positive charge, which enables them to more readily be absorbed into cells. This is an improvement over another method of getting cells to take up gene editing tools, called electroporation, which involves lightly



shocking the cells to get them to open and allow the genetic instructions to enter.

Then the researchers isolated blood stem cells with a protein marker on their surface called CD34. These CD34-positive cells contain the bloodmaking progenitor cells that give rise to the entire blood and immune system.



As gold ions clump together to form nanoparticles, electrons on their surface reflect light at a different wavelength -- hence the red hue. The lab tube in the foreground contains gold ions before they form nanoparticles and loaded with gene editing tools. Credit: Robert Hood, Fred Hutch



"These cells replenish blood in the body every day, making them a good candidate for one-time <u>gene therapy</u> because it will last a lifetime as the cells replace themselves," Adair said.

Observing human <u>blood stem cells</u> in a lab dish, the researchers found that their fully loaded gold nanoparticles were taken up naturally by cells within six hours of being added and within 24 to 48 hours they could see gene editing happening. They observed that the Cas12a CRISPR protein partner was better at delivering very precise genetic edits to the cells than the more commonly used cas9 protein partner.

The gene-editing effect reached a peak eight weeks after the researchers injected the cells into mouse models; 22 weeks after injection the edited cells were still there. The Fred Hutch researchers also found edited cells in the bone marrow, spleen and thymus of the mouse models, a sign that the dividing <u>blood</u> cells in those organs could carry on the treatment without the mice having to be treated again.

"We believe we have a good candidate for two diseases—HIV and hemoglobinopathies—though we are also evaluating other disease targets where small genetic changes can have a big impact, as well as ways to make bigger genetic changes," Adair said. "The next step is to increase how much gene editing happens in each cell, which is definitely doable. That will make it closer to being an effective therapy."

In the study, the researchers report 10 to 20 percent of cells took on the gene edits, which is a promising start, but the researchers would like to aim for 50% or more of the cells being edited, which they believe will have a good chance of combatting these diseases.

More information: Targeted homology-directed repair in blood stem and progenitor cells with CRISPR nanoformulations, *Nature Materials* (2019). DOI: 10.1038/s41563-019-0385-5,



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