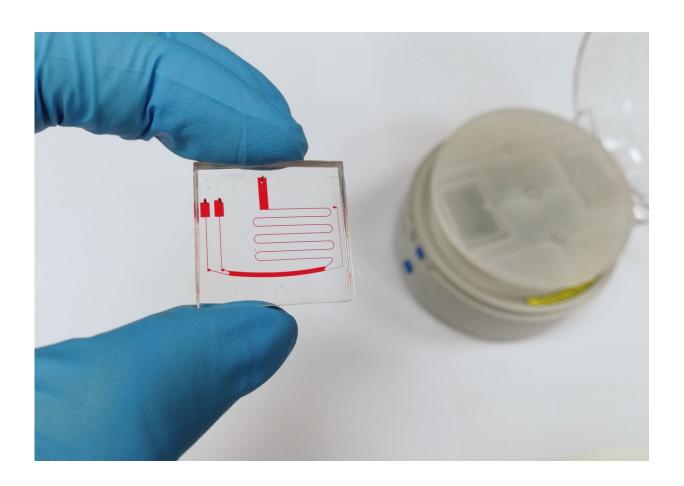


## **Engineering a new spin for disease diagnostics**

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μCENSE chip with the benchtop centrifugal platform in the background. Credit: Joo Chuan Yeo

Researchers at the National University of Singapore have created a new platform with the potential to extract tiny circulating biomarkers of



disease from patient blood. This simple, fast and convenient technique could help realize liquid biopsy diagnostics—a less invasive procedure than the current gold standard: tumor biopsies. Details of the new technique, which utilizes standard laboratory equipment, are reported this week in *Biomicrofluidics*.

Extracellular vesicles are cell messengers that can be found in blood. In cancer, cardiovascular and blood diseases, vesicles transport specific disease-related molecules (biomarkers) that can be used to diagnose these diseases. However, it's tricky to isolate vesicles from blood, because they are miniscule particles, only 30-1,000 nanometers in size.

Current methods of extraction are clinically tedious, time-consuming and expensive, with low throughput and poor extract purity. In this research, scientists utilized a microfluidic centrifugal technique, where a spinning rotor generates pressure, forcing the patient's blood sample to flow through microscopic channels of a specially designed microfluidic chip. The centrifugal force that drives this extraction is similar to the G-force experienced when riding a rollercoaster, only a lot stronger.

A <u>blood sample</u> is first added to the chip's inlet, and then the chip is placed into the centrifugal nanoparticle separation and extraction ( $\mu$ CENSE) platform.  $\mu$ CENSE is then loaded into a standard laboratory benchtop centrifuge and spun. It takes less than eight minutes for the blood and vesicles to separate, and extract can be removed from the chip outlet. This is a hundredfold faster than the high speed ultracentrifugal method that has been used in the past. The  $\mu$ CENSE platform was designed to increase the external force field within a smaller radius, minimizing the <u>centrifugal force</u> and time requirements.

"As we spin the microfluidic chip, the sample in the inlet starts to migrate or move into this curved channel," said Chwee Teck Lim, the team's study lead. "Once there, the centrifugal forces start to separate the



smaller vesicles from the larger particles, because the forces acting on the different sized vesicle are different. So, as they move from inlet to outlet, they start to separate into different zones. The smaller particles remain near the inner wall of the channel and the larger particles move towards the outer wall of the channel, and this separates them into two outlets." This process is similar to how a separator spins to remove cream from the top of milk.

Once isolated the vesicles molecular content can be tested for certain biomarkers of disease. This process includes examining the nucleic acid and protein content. For this study, the group successfully demonstrated that the  $\mu$ CENSE was capable of separating and enriching vesicles from liquid medium exposed to cells grown in a laboratory, by showing that the protein biomarker for vesicles, CD63, was present.

The  $\mu$ CENSE platform is highly versatile for multiple microscale manipulations, as the <u>microfluidic chip</u> can be redesigned for the nanoparticle that needs to be extracted.

Currently, Lim's group is developing the prototype chip design to increase its throughput, and streamline for clinical testing. "We are already looking at trying to conduct a trial on patient clinical samples," Lim said. Ultimately, he hopes to use this technique to identify which biomarkers will be useful in detecting cancer.

**More information:** "Label-free extraction of extracellular vesicles using centrifugal microfluidics," *Biomicrofluidics* March 6, 2018. <u>DOI:</u> 10.1063/1.5019983

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