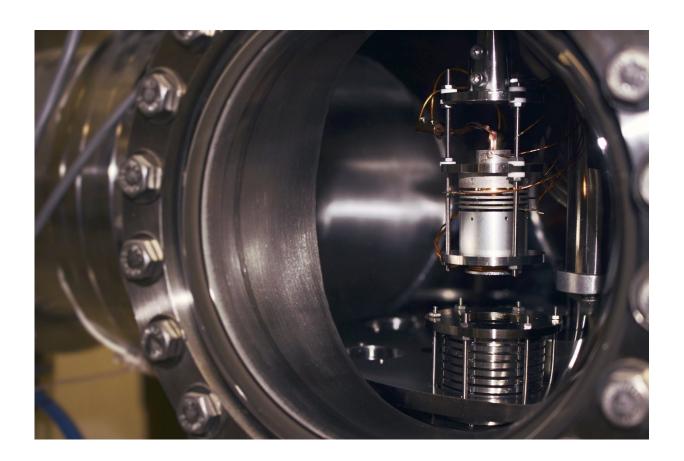


New measurements to guide radiation therapy

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One of many experimental systems for studying LEE-induced damages to condensed biomolecules in the research group of Leon Sanche at the Université de Sherbrooke. In this illustration, we can see a low energy electron gun (top) and a cylindrical multidetector (bottom). This system is used to irradiate (with low energy electrons) short DNA strands condensed onto a cylinder inner surface. The irradiated DNA is afterwards analyzed by HPLC/MS-MS to detect any chemical changes. Credit: Vincent Lemelin, Université de Sherbrooke, Québec, Canada



When ionizing radiation passes through living tissue, it interacts with molecules present in the cells, stripping away electrons and producing charged species known as ions. Ionizing radiation used for cancer treatment includes gamma rays, X-rays and energetic particles such as alpha and beta rays.

The electrons produced by this process, known as secondary electrons, can themselves go on to wreak further havoc, causing even more dramatic changes. This week in the *Journal of Chemical Physics*, a group of investigators reports studies of the impact of secondary electrons on a model of DNA.

The measurements were made in a condensed-phase environment. Compared to isolated electron-molecule experiments, the condensed-phase measurements are made under conditions closer to those found in living tissue. The results will be used to accurately calculate the damage and <u>radiation</u> dose delivered to patients in radiotherapy, when <u>cancer cells</u> are bombarded with ionizing radiation.

Secondary electrons are the most important species created by ionizing radiation in living tissue. These "low energy electrons," or LEEs, interact with biological molecules, sometimes breaking them into fragments. One of the affected molecules is deoxyribonucleic acid, or DNA, the molecule that carries genetic code. The long, chainlike DNA molecule consists of a ladder of base pairs connected to each other through a deoxyribose phosphate group.

The precise way LEEs interact with portions of the DNA molecule, the bases themselves or the phosphate backbone, is still not precisely understood, although LEEs do have enough energy to initiate DNA strand breaks. This can affect cell function, leading to mutations and



even cell death. In this week's report, the investigators employed a model molecule known as dimethyl phosphate, or DMP, to study the interaction of LEEs with the phosphate backbone of DNA.

New radiation treatment methods, currently under development, can precisely target the radiation to specific cancer <u>cells</u> or even specific locations within those cells. This method, known as targeted radionuclide therapy, or TRT, involves the use of molecules labeled with radioactive atoms that are injected into patients and localized in cancer cells. Once in place, the radioactive molecules produce ionizing radiation inside or close to cancer cells. This radiation then goes on to generate localized LEEs.

An important part of the TRT method involves computer simulations used to predict the interactions of LEEs with biological matter and the amount of radiation absorbed by the targeted biomolecules or cells. One of the key parameters in these simulation models are absolute cross sections, which give the probability of interaction between a single LEE and a target molecule. The work reported here represents the first direct measurement of absolute cross sections for the phosphate unit in DNA, values required to calculate strand breaks induced by LEEs.

The DNA present in a living system is surrounded by water and other types of <u>molecules</u>, so studying these processes in a more realistic environment is particularly desirable. In future work, the DNA will be embedded in water and molecular oxygen, known to sensitize cells to radiotherapy.

More information: V. Lemelin et al, Absolute vibrational excitation cross sections for 1-18 eV electron scattering from condensed dimethyl phosphate (DMP), *The Journal of Chemical Physics* (2017). DOI: 10.1063/1.5008486



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