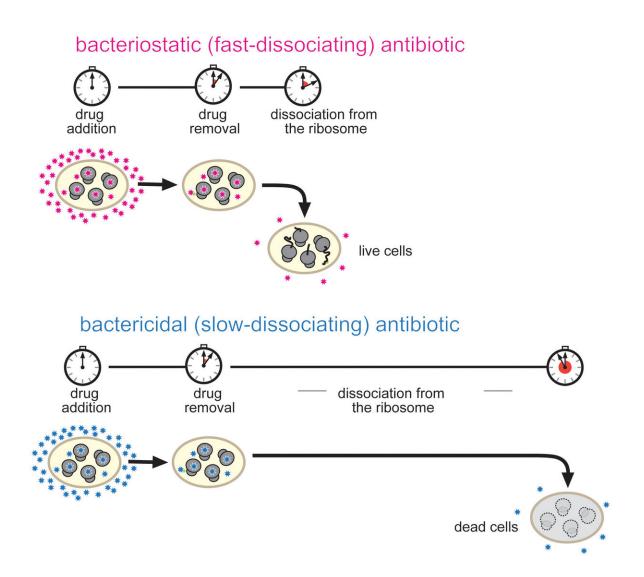


Teaching antibiotics to be more effective killers

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Research shows prolonged inhibition of protein synthesis leads to cell death. Credit: Proceedings of the National Academy of Sciences



Research from the University of Illinois at Chicago suggests bond duration, not bond tightness, may be the most important differentiator between antibiotics that kill bacteria and antibiotics that only stop bacterial growth.

While both types of antibiotics are used to treat a variety of bacterial infections, <u>bactericidal antibiotics</u> – those that kill bacteria – can be taken for shorter periods, are associated with a lower risk of infection recurrence and often cure the infection much better than bacteriostatic antibiotics.

"Bacteriostatic antibiotics work by slowing the growth of bacterial cells while the individual's immune system fights the infection," said Alexander Mankin, lead author of the study and director of the Center for Biomolecular Sciences in the UIC College of Pharmacy. "Often, this is enough to treat an infection; but if the immune system is not strong enough, the <u>infection</u> may persist."

Unfortunately, not much is known about why the sometimes-similar antibiotic molecules interact differently with bacteria in the body.

Mankin and his colleagues looked at one class of antibiotics called macrolides, which work by binding to the <u>ribosome</u> of the bacteria to stop protein synthesis. However, while <u>macrolide antibiotics</u> are all structurally similar and act on the same molecular target (the ribosome), some are bactericidal and others are bacteriostatic.

Using a new technique developed by Maxim Svetlov, a postdoctoral researcher in the lab of Mankin and Nora Vázquez-Laslop, an associate professor in the center, the researchers studied the differences between the bacteriostatic and bactericidal macrolides. The technique makes it



possible to analyze how tightly the drugs interact with the ribosome and to measure the speed with which the antibiotics can disconnect from the target.

"Researchers usually think that it is the tightness of the drug's binding to the ribosome that makes the difference between an antibiotic that kills bacteria and one that only slows <u>bacterial growth</u>," Mankin said. "Therefore, we were surprised to find that tightness of the drug binding does not define the ability of the drug to kill <u>bacteria</u>.

"Instead we understood that bactericidal drugs dissociate from the ribosome at a significantly slower rate," Mankin said.

The presence of an extended side chain in the structure of the antibiotic functions "like an extra hand to hold on to the ribosome; it allows the drug to bind for a longer period," says Mankin.

The findings, published in the *Proceedings of the National Academy of Sciences*, offer a new and previously unexplored possibility.

"The results of this study suggest that when we talk about the mechanism of antibiotic action, we need to talk about more than 'how tight' a drug binds," Mankin said. "We also need to talk about kinetics and the rate of a drug's disassociation from the ribosome."

In addition to improving outcomes for individuals who need <u>antibiotic</u> <u>treatment</u>, particularly for patients who are immunocompromised, understanding the difference between bacteriostatic and bactericidal <u>antibiotics</u> may also help to address the problem of rising <u>antibiotic</u> <u>resistance</u> in society.

"If we can understand these mechanisms, we may be able to reduce the rate at which antibiotic resistance develops and repeat infections occur,"



Mankin said.

More information: Maxim S. Svetlov et al. Kinetics of drug–ribosome interactions defines the cidality of macrolide antibiotics, *Proceedings of the National Academy of Sciences* (2017). DOI: 10.1073/pnas.1717168115

Provided by University of Illinois at Chicago

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