

Your body's cells use and resist force, and they move. It's mechanobiology

October 20 2017, by Joshua Chou, Kate Poole And Maté Biro



Credit: AI-generated image (disclaimer)

Mechanical forces rule biological processes, from the contractions of the pump-like heart, to muscles that resemble strings and pullies, and cells that carry out microscopic tugs-of-war.

Previously, these mechanical aspects of biology have been largely



ignored, not least due to a lack of technology allowing for intricate mechanical measurements.

But better tools are being developed, and these enable tracking of mechanical activity in cells and tissues.

And because of this visibility, <u>new drugs</u> and treatments are starting to emerge.

Mechanobiology is the science of how cells and tissues sense and respond to <u>mechanical forces</u>.

Just like we humans have muscles and bones that give us the ability to exert forces, each of our cells also has a skeleton: the <u>cytoskeleton</u>. This network of fibres allows cells to exert and resist forces, and enables them to move.

Watching T cells

<u>T cells</u> are part of our immune system: they can act like cellular assassins, killing other cells such as those infected with viruses, or cancer cells.

At the microscale (about one hundredth of a human hair), we can visualise and follow T cells "hunting" for cancer cells as they move and push their way through tissues. This applies the approach known as <u>3-D</u> traction force microscopy (TFM).

Upon finding a cancer cell, a T cell firmly grasps its target, and delivers a "kiss of death".

Techniques known as <u>dual pipette aspiration</u> (DPA) and optical tweezers (see video below) allow us to grab <u>individual cells</u>, and pair them



together in a controlled manner. This allows us to understand and illustrate the mechanics behind this deadly "kiss".

Using mechanobiological techniques to see how T cells find and kill cancer cells, may allow better targeting of anti-cancer immunotherapies.

The first immunotherapy targeting cancer using a patient's own T cells was approved just recently by the <u>United States Food and Drug</u> <u>Administration</u> (FDA).

Sensing the force

Cells use <u>force</u> sensors to detect and distinguish between many of the physical signals that they experience.

A major class of force sensors are "mechanosensitive ion channels". These are holes, or pores, in the surface of the cell that can open and shut.

When the cell senses a physical force or a mechanical stimulus (in essence, like a microscopic prod), these pores can open. Chemicals move in and out, and a tiny electrical current is conducted across the cell wall. This can be measured by attaching small electrodes to the surface of a cell.

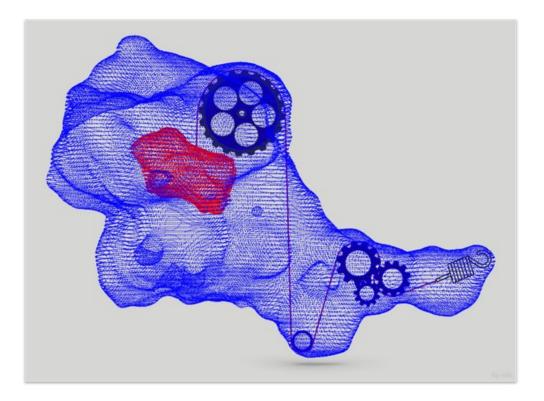
Many types of cells and tissues have such sensors, and respond to changes in mechanical loads. These include the neurons that underpin our sense of touch, metastasising <u>cancer cells</u> and the cells that maintain our cartilage in bones.

The drug <u>EVENITY</u> – which aims to prevent bone loss in osteoporosis – acts through this pathway. The drug blocks sclerostin, a key factor that naturally inhibits bone formation based on the mechanosensing functions



of bone-forming cells.

Trialled in mice travelling to the International Space Station, the treatment has now passed <u>human clinical trials</u> and is awaiting FDA approval for use in the treatment of osteoporotic patients in the United States.



Each cell's cytoskeleton acts like a system of internal pulleys and strings. Credit: Dr Maté Biro, CC BY-SA

Organ on a chip

Organ-on-chip technology is designed to aid drug development, disease



modelling and personalised medicine. Each individual unit is made of a transparent material known as a polymer: it's about the size of a USB stick, and consists of hollow channels lined by living <u>human cells</u>.

These chips differ from other laboratory tests such as cell culture, in that they can mimic the physiology and mechanics of how cells interact with living tissues (rather than just looking at responses in individual cells).

For example, organs-on-chips can recreate the architecture of human organs at the microscopic level, including the <u>intestines, kidney, skin</u>, <u>bone marrow and areas of the brain</u>.

An example using lung <u>tissue</u> is described in the video below. This technology provides a way to see diseases in tissues, and is an alternative to animal testing for drug development.

Many researchers and biotechnology companies hope that technology such as organs-on-chips will accelerate the development of new drugs, and advance personalised medicine.

Using existing techniques, clinical studies can take years to complete and testing a single compound can cost many millions of dollars. Also, preclinical animal studies often fail to predict human responses because animal models do not always accurately mimic human biological responses.

In April 2017, the FDA announced <u>a multi-year research and</u> <u>development agreement</u> to evaluate organs-on-chips technology, starting with a <u>liver-chip</u>.

The agreement may expand in the future to cover additional organ-chips, including kidney, lung and intestine models.



Mechanobiology is integrating physical sciences into biology and driving the development of new technologies. Watching <u>cells</u> in motion, understanding and measuring forces at the cellular scale, and creating mini models of human tissues in the lab are just the beginning.

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