

# Novel small antibody fragment—a valuable tool in crystallography

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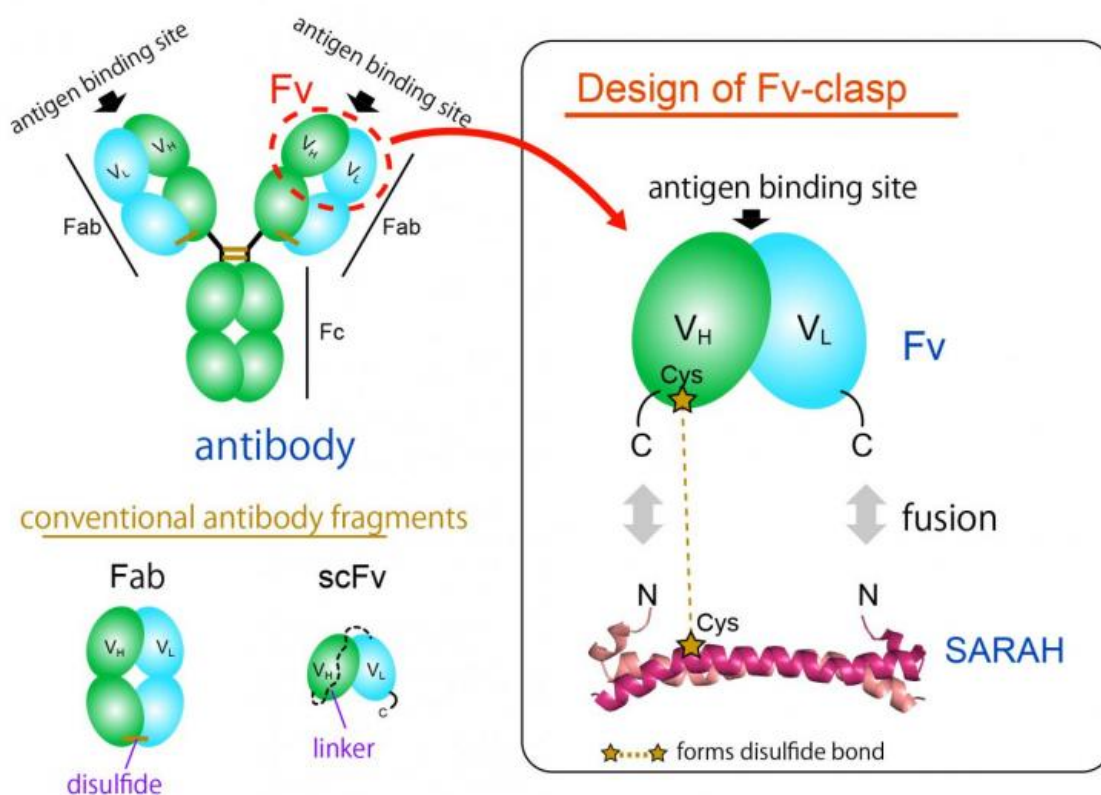


Figure.1. Design concept of a novel antibody fragment format "Fv-clasp". Fv-clasp is a simple fusion of the SARAH domain derived from human Mst1 protein to the Fv (green: heavy chain, cyan: light chain) fragment of an antibody. V<sub>H</sub>/V<sub>L</sub> dimer is stabilized by the dimer conformation of the SARAH domain (dark and light magenta). Credit: Osaka University

Antibodies are a powerful weapon system in defending our body against invaders such as bacteria and viruses. Each antibody consists of four amino acid polypeptide chains: two heavy chains and two light chains joined to form a Y-shaped molecule. Antibodies recognize a specific antigen unique to its target as they possess the antigen-binding sites located at the upper tips of the Y. While antibody-based therapeutics have been established as front-line drugs, little headway has been made in the use of antibodies as research tools in small molecule drug discovery, particularly in the field of x-ray crystallography.

X-ray crystallography is a technique that uses x-ray diffraction patterns to determine high-resolution, three-dimensional structures of [molecules](#) such as proteins, small organic molecules, and materials. The major challenge in X-ray crystallography approaches remains the production of high-quality diffracting crystals.

In recent years, there is increasing use of antibody fragments as crystallization chaperones to aid the structural determination of otherwise "uncrystallizable" or "undruggable" target proteins. The basis for the strategy is to increase the probability of obtaining well-ordered crystals by minimizing the conformational heterogeneity in the target protein.

The most commonly used antibody fragment format is the Fab format, but it is difficult to produce in bacterial expression systems because of its large and complex structure. The Fv fragment of the antibody, which contains only one heavy and one light chain, would be ideal for this application thanks to its simple and small architecture. However, Fv is rarely used because the two chains dissociate easily, leading to a loss of function.

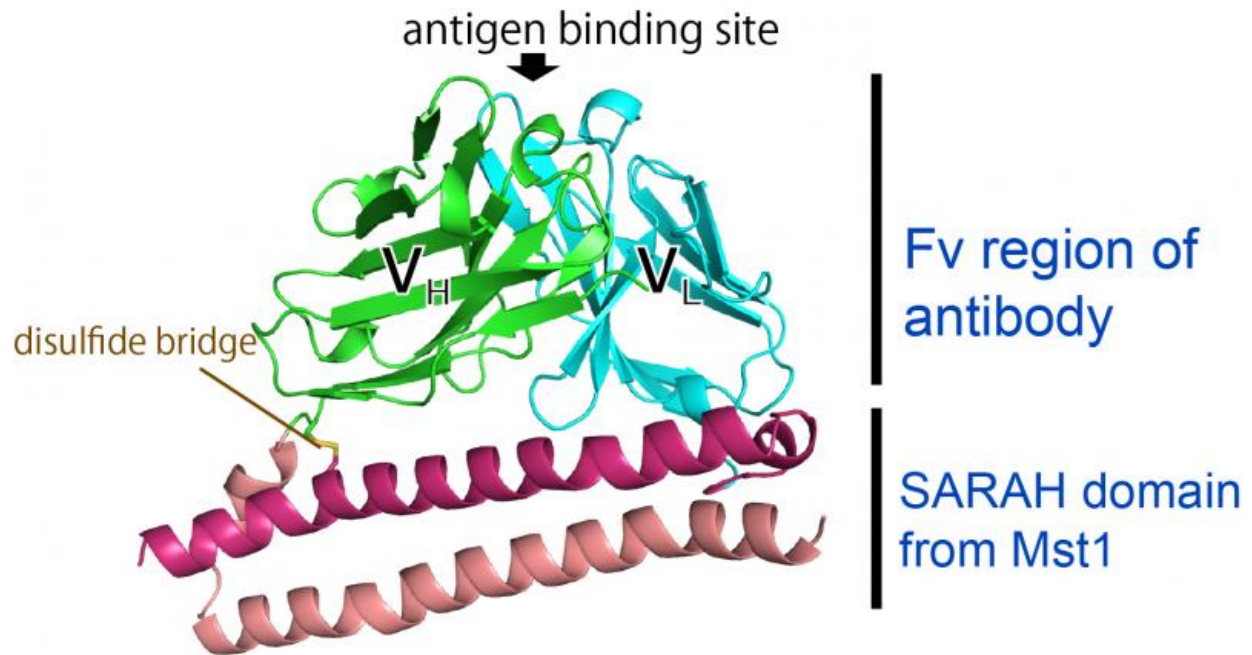


Figure.2. Crystal structure of Fv-clasp. Actual three dimensional structure of Fv-clasp determined by X-ray crystallographic analysis. Credit: Osaka University

Now, a team of Osaka University researchers has designed a novel single-chain Fv fragment with improved production compatibility, stability and crystallizability, while maintaining the binding ability of the original molecule. They recently published their findings in *Structure*.

"We successfully produced a new fragment by fusing an anti-parallel coiled-coil structure derived from a particular domain of a human enzyme, Mst1 kinase, to the antigen-binding sites of an antibody," study lead author Takao Arimori explains. "The resulting chimeric molecule, Fv-clasp, was functionally and structurally equivalent to the Fv of the original antibody."

Notably, switching from Fab to Fv-clasp format markedly enhanced the

antibody-assisted crystallization of two biologically important proteins, the extracellular domains integrin  $\alpha 6 \beta 1$  and sorLA. Integrin  $\alpha 6 \beta 1$  plays a significant role in the attachment of iPS cells and ES cells as well as many liver cancer cells to the basement membrane, an extracellular protein network foundation present in tissues in most cells and organs, while sorLA is [protein](#) receptor implicated in Alzheimer's disease.

"The universal applicability of the Fv-clasp design to large-scale and inexpensive production makes it desirable for industrial applications. Furthermore, its high heat stability is a great advantage for immunotherapies," corresponding author Junichi Takagi says. "Aside from the field of structural biology, we anticipate that the Fv-clasp design will contribute to the expansion of already eminent antibody use in both basic and applied sciences."

**More information:** Takao Arimori et al. Fv-clasp: An Artificially Designed Small Antibody Fragment with Improved Production Compatibility, Stability, and Crystallizability, *Structure* (2017). [DOI: 10.1016/j.str.2017.08.011](https://doi.org/10.1016/j.str.2017.08.011)

Provided by Osaka University

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