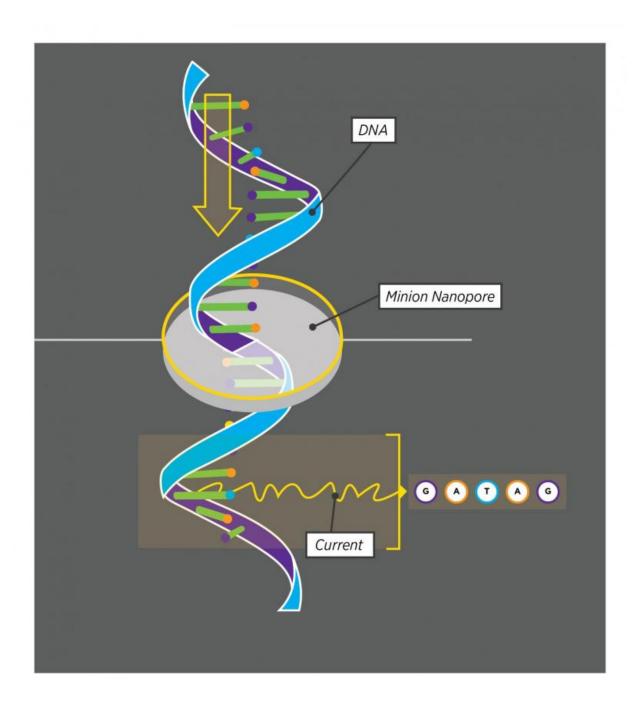


A better way to read the genome

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As a strand of DNA moves through the MinION nanopore, the MinION 'sees' five nucleotides at a time. Every set of five nucleotides produces a characteristic current, allowing the MinION to read out the DNA code in a single, continuous stream. Credit: Yesenia Carrero/UConn

UConn researchers have sequenced the RNA of the most complicated gene known in nature, using a hand-held sequencer no bigger than a cell phone.

If DNA is the blueprint of life, RNA is the construction contractor who interprets it, so sequencing RNA tells you what's really happening inside a cell.

Genomicists Brenton Graveley from the UConn Institute of Systems Genomics, postdoctoral fellow Mohan Bolisetty, and graduate student Gopinath Rajadinakaran teamed up with UK-based Oxford Nanopore Technologies to show that the company's MinION nanopore sequencer can sequence <u>genes</u> faster, better, and at a much lower cost than the standard technology. They published their findings on Sept. 30 in *Genome Biology*.

If your genome was a library and each gene was a book, some genes would be straightforward reads - but some would be more like a "Choose Your Own Adventure" novel. Researchers often want to know which version of the gene is actually expressed in the body, but for complicated choose-your-own-adventure genes, that has been impossible.

Graveley, Bolisetty, and Rajadinakaran solved the puzzle in two parts. The first was to find a better gene-sequencing technology. In order to sequence a gene using the old, existing technology, researchers first

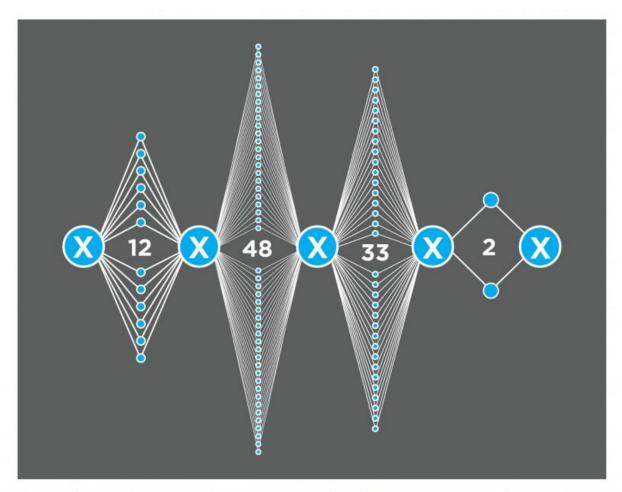


make lots of copies of it, using the same chemistry our bodies use. They then chop up the gene copies into tiny pieces, read each tiny piece, and then, by comparing all the different pieces, try to figure out how they were originally put together. The technique hinges on the likelihood that not all the copies got chopped up into exactly the same pieces. Imagine watching different scenes from a movie, out of order. If you then watched the same movie, but cut into scenes at slightly different places, you could compare the two versions and start to figure out which scenes connect to which.

That technique won't work for choose-your-own-adventure genes, because if you copy them the way the body does, using RNA, each copy can be slightly - or very - different from the next. Such different versions of the same gene are called isoforms. When the different isoforms get chopped up and sequenced, it becomes impossible to accurately compare the pieces and figure out which versions of the gene you started with.

If the gene were a movie, "you wouldn't be able to tell that scenes 1 and 2 were present together," Bolisetty says.





There are four variable sections in the gene Dscam1. Every blue dot represents one potential version of its section. Each time Dscam1 is expressed, only one version is actually expressed in each of the four variable sections. If you do the math, you find Dscam1 has 38,016 potential versions (isoforms) in total.

Then last year, the nearly impossible suddenly became possible. Oxford Nanopore, a company based in the UK, released its new nanopore sequencer, and offered one to Graveley's lab. The nanopore sequencer, called a MinION, works by feeding a single strand of DNA through a tiny pore. The pore can only hold five DNA bases - the 'letters' that spell out our genes - at a time. There are four DNA bases, G, A, T, and C, and



1,024 possible five-base combinations. Each combination creates a different electrical current in the nanopore. GGGGA makes a different current than AGGGG, which is different again than CGGGG. By feeding the DNA through the pore and recording the resulting signal, researchers can read the sequence of the DNA.

For the second part of the solution, Graveley, Bolisetty, and Rajadinakaran decided to go big. Instead of sequencing any old chooseyour-own-adventure gene, they chose the most complex one known, Down Syndrome cell adhesion molecule 1 (Dscam1), which controls the wiring of the brain in fruit flies. Dscam1 has the potential of making 38,016 possible isoforms, and every fruit fly has the potential to make every one of them, yet how many of these versions are actually made remains unknown.

Dscam1 looks like this: X-12-X-48-X-33-X-2-X, where X's denote sections that are always the same, and the numbers indicate sections that can vary (the number itself shows how many different options there are for that section).

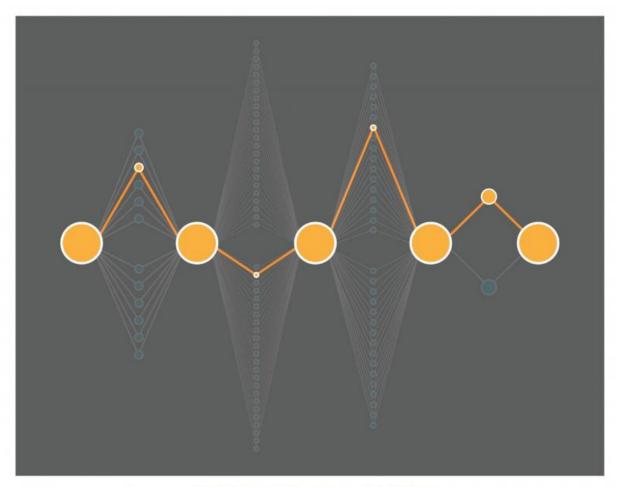
To study how many different isoforms of Dscam1 actually exist in a fly's brain, the researchers first had to convert Dscam1 RNA into DNA. If DNA is the book or set of instructions, RNA is the transcriber that copies the book so that it can be translated into a protein. The DNA includes the instructions for all 38,016 isoforms of the Dscam1 gene, while each individual Dscam1 RNA contains the instructions for just one. No one had yet used a MinION to sequence copies of RNA, and though it was likely it could be done, demonstrating it and showing how well it worked would be a substantial advance in the field.

Rajadinakaran took a fruit fly brain, extracted the RNA, converted it into DNA, isolated the DNA copies of the Dscam1 RNAs, and then ran them through the MinION's nanopores. In this one experiment, they not



only found 7,899 of the 38,016 possible isoforms of Dscam1 were expressed but also that many more, if not all versions are likely to be expressed.

"A lot of people said 'The MinION will never work,'" Graveley says, "but we showed it works using the most complicated gene known."



[Paths in orange] show one potential isoform



The study demonstrates that <u>gene sequencing technology</u> can now be accessed by a much broader range of researchers than was previously possible, since the MinION is both relatively inexpensive and highly portable so that it requires almost no lab space.

"This type of cutting-edge work puts UConn at the forefront of technology development and strengthens our portfolio of genomics research," says Marc Lalande, director of UConn's Institute for Systems Genomics. "Also, thanks to the investments in genomics through the University's Academic Plan, Brent Graveley can leverage his expertise so that faculty and students across our campuses will successfully compete for grant dollars and launch bioscience ventures."

Graveley will speak about the research at the Oxford Nanopore MinION Community Meeting at the New York Genome Center on Dec. 3.





The MinION gene sequencer in Brenton Graveley's lab is state-of-the art technology that costs about \$1,000 and is roughly the same size as an iPhone. Credit: Kim Krieger/UConn Photo

As for next steps, the researchers plan on going even bigger: sequencing every bit of RNA from beginning to end inside a single cell, something that cannot be done with traditional gene sequencers.

"This technology has amazing potential to transform how we study RNA biology and the type of information we can obtain," says Graveley. "Plus the fact that the MinION is a hand-held sequencer that you plug into a laptop is simply unbelievably cool!"

More information: Mohan T. Bolisetty et al. Determining exon connectivity in complex mRNAs by nanopore sequencing, *Genome Biology* (2015). DOI: 10.1186/s13059-015-0777-z

Provided by University of Connecticut

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