

Predictive software can precisely identify most effective ways to target genes with gene editing mechanism CRISPR-Cas9

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This is a computer graphic of an RNA molecule. Credit: Richard Feldmann/Wikipedia



The remarkable ease and accuracy with which scientists can alter genomes using the CRISPR-Cas9 system has led to promising advances toward improving human health and the environment through genetic engineering. Cas9, a protein found naturally in certain bacteria, functions like a pair of molecular scissors to precisely cut sections of DNA and is extremely effective as a gene-editing tool. It can be directed to a specific gene through the use of a matching guide RNA sequence to perform gene mutations, putting programmable control of gene editing in the hands of scientists.

Even though Cas9 can precisely carry out these functions, more than one guide RNA can be a possible match for any given <u>gene target</u>, which leaves scientists with preliminary troubleshooting work to select the best guide RNA for any given <u>gene-editing</u> task.

To eliminate the trial-and-error process of selecting guide RNAs for each job, a team led by George Church, HMS Robert Winthrop Professor of Genetics, Core Faculty member at the Wyss Institute for Biologically Inspired Engineering, and affiliated faculty member of the Harvard-MIT Program in Health Sciences and Technology, has developed a new, straightforward software program for predicting the best guide RNAs for directing Cas9 to gene targets.

"We started off by asking ourselves, is there something in the guide RNA sequence that could suggest one would work better than others?" said the study's lead author Raj Chari, research fellow in genetics at Harvard Medical School.

The software hierarchically ranks how effective any given guide RNA will target a desired gene target based on experimental data gathered using human genomes, which is unlike other gene targeting algorithms currently available to scientists that operate on first principles only.



"This will allow for widespread improvement in the speed and accuracy with which scientists can select the appropriate guide RNA for achieving their desired gene-editing results," said Church.

Made publically available by Church's team, it not only contains information about effective guide RNAs for Cas9 extracted from the bacteria species S. pyogenes, which is the most commonly-used bacterial source of Cas9, but also includes the world's most comprehensive library of guide RNA from a different bacteria species that has been rising in its popularity of use, *S. thermophilus*.

To develop the algorithm that is the basis for the new software, Church's team carried out a high-throughput analysis of the activity between many gene targets and complimentary guide RNAs, searching for patterns in the guide RNA sequences that could indicate how effectively they would bind to any given gene target.

In doing so, they chose a unique sequence from each gene target and synthesized an identical strand of DNA to represent that target, and then they created a very large DNA library and stored it in cell plasmid. The library was then delivered, using a virus as a delivery vehicle, into the genomes of cultured cells. To test which guide RNAs would be the most effective at reaching each of these unique targets, complimentary guide RNAs were inserted with Cas9 into the cells, giving Cas9 the opportunity to make genome mutations at the sites where successful matching occurred. A basic genome extraction and sequencing process then easily revealed which guide RNA was the best match for each target.

From this data, the team developed their novel algorithm to rank and score the most effective guide RNAs for targeting virtually any human gene, even ones lacking experimental data in their software database.



"Analyzing our results, we identified certain features in guide RNA sequences that are indicative of how well they work at directing Cas9 to a desired gene target," said Chari. "By designing an algorithm based on these features, we can now look at any guide RNA sequence and assign it a score for how effective it is predicted to work."

By speeding up this part of the gene-editing process, scientists can focus efforts on applying Cas9 gene editing to developing gene therapies against diseases such as sickle cell anemia, cancer or HIV, and to creating genetically engineered organisms that could help clean up the environment or perform other functions, such as sustainably producing chemical commodities.

"For any scientist using RNA-guided Cas9 in their day-to-day work, it will be greatly beneficial to have a tool like this that quickly narrows down which guide RNA will be the most effective to direct Cas9 to a gene of interest," said Chari.

More information: Unraveling CRISPR-Cas9 genome engineering parameters via a library-on-library approach, *Nature Methods*, <u>DOI:</u> <u>10.1038/nmeth.3473</u>

Provided by Harvard Medical School

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