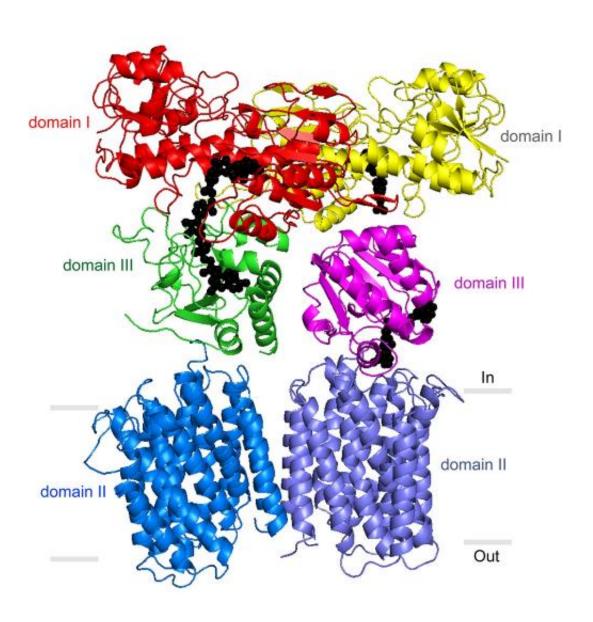


## X-ray study reveals division of labor in cell health protein

January 29 2015



A recent study performed in part at SLAC's SSRL X-ray facility has provided new insights into how the critical mitochondrial enzyme transhydrogenase, or



TH, works in a key process that maintains healthy cells. The crystal structure of TH shows two copies of the molecule (left and right), each of which contains three domains (I, II, III). Structural asymmetry is observed for domain III: One of the structures is facing up (green) to catalyze the production of NADPH from its precursor (black spheres); the other is facing down (magenta) towards the transmembrane domain II to facilitate the transit of a proton. Labels "in" and "out" denote the mitochondrial matrix and the space outside the inner mitochondrial membrane, respectively. Credit: C. David Stout/The Scripps Research Institute

Researchers working in part at the Department of Energy's SLAC National Accelerator Laboratory have discovered that a key protein for cell health, which has recently been linked to diabetes, cancer and other diseases, can multitask by having two identical protein parts divide labor.

The TH enzyme, short for transhydrogenase, is a crucial <u>protein</u> for most forms of life. In humans and other higher organisms, it works within mitochondria – tiny, double-hulled oxygen reactors inside cells that help power most cellular processes.

"Despite its importance, TH has been one of the least studied mitochondrial enzymes," said C. David Stout, a scientist at The Scripps Research Institute whose group led the research. The new study, published Jan. 9 in *Science*, is an important step toward understanding how this protein manages to perform two crucial cellular tasks at the same time.

## **Two Crucial Processes**

As a mitochondrion burns oxygen, it pumps protons out of its innermost compartment, or matrix. Part of the TH protein extends through the membrane that surrounds the matrix; it allows a one-by-one flow of



protons back through the membrane. This proton influx, in turn, is linked to the production of NADPH, a compound crucial for defusing oxygen radicals that are harmful to cells.

But how do TH enzymes couple proton transport and NADPH production? Although Stout's laboratory and others have previously described portions of the TH enzyme that protrude from the membrane into the mitochondrial matrix, a precise understanding of TH's mechanism has been elusive. The enzyme has an exceptionally loose structure that makes it hard to evaluate by X-ray crystallography, the standard tool for determining structures of large proteins.

"Key details we've been lacking include the structure of TH's transmembrane portion, and the way in which the parts assemble into the whole enzyme," said Josephine H. Leung, a graduate student in the Stout laboratory who was the lead author of the new study.

For the first time, the team was now able to determine precise details of the transmembrane portion using X-rays from SLAC's Stanford Synchrotron Radiation Lightsource (SSRL) and Argonne National Laboratory's Advanced Photon Source (APS), both DOE Office of Science User Facilities. Together with X-ray and electron microscopy data of the whole protein, the study provided major clues as to how TH works.

## **Flipping Functions**

The analysis revealed that two identical copies of TH are bound together in what is called a dimer, and that one copy appears to be involved in proton transport while the other takes part in NADPH production. "Our new study helps clear up some mysteries – suggesting how the enzyme structure might harness protons and indicating that its two sides are able to alternate functions, always staying in balance," Stout said.



Attached to TH's transmembrane structure, just inside the mitochondrial matrix, is the "domain III" structure, which binds NADPH's precursor molecule during NADPH synthesis. Previously, scientists did not understand how two such structures could work side by side in the TH dimer and not interfere with each other's activity.

The new data suggest that these side-by-side structures are highly flexible and always have different orientations.

"Our most striking finding was that the two domain III structures are not symmetric – one of them faces up while the other faces down," said Leung.

In particular, one of the structures is apparently oriented to catalyze the production of NADPH, while the other is turned towards the membrane, perhaps to facilitate transit of a proton. The new structural model indicates that with each proton transit, the two domain III structures flip and switch their functions. "We suspect that the passage of the proton is what somehow causes this flipping of the domain III structures," said Leung.

But much work remains to be done to determine TH's precise structure and mechanism. For example, the new structural data provide evidence of a likely proton channel in the TH transmembrane region, but show only a closed conformation of that structure. "We suspect that this channel can have another, open conformation that lets the <u>proton</u> pass through, so that's one of the details we want to study further," said Leung.

**More information:** "Division of labor in transhydrogenase by alternating proton translocation and hydride transfer." *Science* 9 January 2015: Vol. 347 no. 6218 pp. 178-181 DOI: 10.1126/science.1260451



## Provided by Stanford University

Citation: X-ray study reveals division of labor in cell health protein (2015, January 29) retrieved 20 April 2024 from <a href="https://phys.org/news/2015-01-x-ray-reveals-division-labor-cell.html">https://phys.org/news/2015-01-x-ray-reveals-division-labor-cell.html</a>

This document is subject to copyright. Apart from any fair dealing for the purpose of private study or research, no part may be reproduced without the written permission. The content is provided for information purposes only.