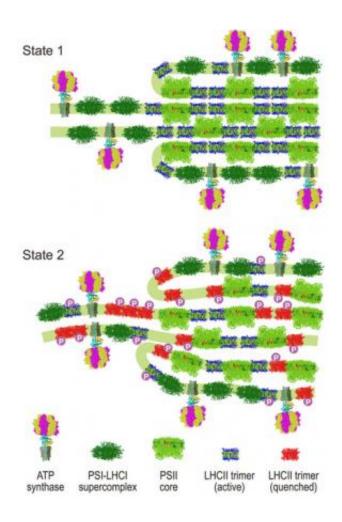


New insight into photosynthesis

May 27 2014, by Martyn Bull



Side views of the light-sensitive thylakoid membranes inside the chloroplast of green algae C. reinhardtii. For State 1, the membrane structure is well-ordered. For State 2, the well-ordered structure is disrupted and the activity of the majority of the light harvesting proteins LHCII is inhibited.

The way that algae and plants respond to light has been reinterpreted



based on results from experiments studying real-time structural changes in green algae. Under particular lighting conditions during photosynthesis, the well-ordered stacking and alignment of light-sensitive membranes in the algae are disrupted. There is no significant movement of the membrane embedded light harvesting proteins, which rather become largely inactive. These new findings challenge widely accepted views of how algae respond to light where the light harvesting proteins were thought to move around the membranes.

The new results from researchers at the Paul Scherrer Institut (PSI) and colleagues in Japan, France and Hungary are published in the *Proceedings of the National Academy of Sciences* and a number of related articles.

Photosynthesis

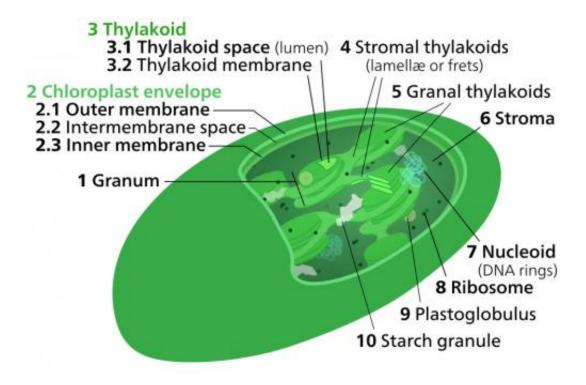
Photosynthesis is the process used by plants, algae and cyanobacteria to convert sunlight into chemical energy that can be stored and later released to fuel cellular activity. This chemical energy is stored in carbohydrate molecules, such as sugars, that are made from carbon dioxide and water, with oxygen released as a waste product. Photosynthesis maintains atmospheric oxygen levels and supplies virtually all of the organic compounds and energy necessary for life on Earth.

"Plants have to operate in very different environments and <u>lighting</u> <u>conditions</u>," says Gergely Nagy at the Laboratory for Neutron Scattering, Paul Scherrer Institute, first author of the PNAS study. "They have to react to the changing colours of <u>light</u> and manage the different illumination of their leaves from the top to the bottom of a plant. The same is true for algae in water at different depths in the sea or lakes."

Algae and plants respond to changes in light quality by regulating the



absorption capacity of their two photosystems. The two photosystems are embedded in membranes ('thylakoid membranes') stacked up inside small compartments known as chloroplasts. "Photosynthesis is a very sensitive process," says Nagy. "The two photosystems are connected together, and an imbalance in absorbed energy between them may damage them."



Photosynthesis takes place on the thylakoid membranes found in chloroplasts within plant cells. Credit: Wikipedia

Understanding state transitions

The conversion of light energy to chemical energy in the two photosystems is balanced by a sophisticated regulatory mechanism called state transitions. Although the core idea of state transitions has been established since 1969, in-depth understanding of this complex process



and the accompanying structural changes has only recently advanced.

"Our data have allowed us to propose a general model for state transitions in algae that explains the molecular mechanism and physiological consequences, and clarify the nature of accompanying structural changes," says Nagy. "We found that state transitions change the stacking and periodicity of the thylakoid membranes and altered the protein–protein interactions within these membranes."



Green algae C Reinhardtii. Credit: Protist Information Server, Courtesy of Dr. Sc. Yuuji Tsukii

These structural changes accompany the conversion of one of the light-



harvesting proteins (LHCII) into an inactive, energy-dissipating mode with only minor movement of the LHCII light harvesting proteins within the membrane. "This new finding differs from previous ideas where it was thought that different amounts of the LHCII proteins - up to 80% in some microalgae - would move between the two photosystems in the membrane during state transitions," says Nagy.

Insights from neutron scattering

The research team has developed a suite of non-invasive techniques to study living cells of the <u>green algae</u> Chlamydomonas reinhardtii in a natural environment: <u>neutron scattering</u> (small-angle neutron scattering – SANS) with supporting experiments using visible light (circular dichroism and time-resolved absorption and fluorescence spectroscopy).

Neutron scattering is a technique well-suited for studying biological samples. It is non-destructive, and there is no need to stain a sample, or fix with chemicals or low-temperatures. Living cells under natural conditions are illuminated with a beam of neutrons and data from the scattered beam can be used to calculate structural information in the sample in the nanometre to micrometre range.





Renata Ünnep und Gergely Nagy at the SANS instrument at PSI's neutron source SINQ. Credit: Paul Scherrer Institut/ Mahir Dzambegovic

"Neutron experiments are extremely well-suited to provide accurate structural information on the membrane ultrastructure," says Nagy. "The state transitions deeply affect the organisation of light-sensitive thylakoid membranes in the chloroplast of the algae. Neutron experiments give a unique perspective where we can watch the state transitions as they are happening. This is the first time that ultrastructure changes have been clearly identified with the aid of neutrons."

The neutron scattering experiments were carried out at the Institut Laue-Langevin neutron source in France and the Swiss SINQ neutron source at the Paul Scherrer Institut.

More information: Nagy, G., Ünnep, R., Zsiros, O., Tokutsu, R., Takizawa, K., Porcar, L., Moyet, L., Petroutsos, D., Garab, G., Finazzi,



G. and Minagawa, J. "Chloroplast remodeling during state transitions in Chlamydomonas reinhardtii as revealed by noninvasive techniques in vivo." *PNAS* April 1, 2014 vol. 111 no. 13 5042-5047. DOI: 10.1073/pnas.1322494111

Related:

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The ultrastructure and flexibility of thylakoid membranes in leaves and isolated chloroplasts as revealed by small-angle neutron scattering. *Biochim Biophys Acta Bioenerg* DOI: 10.1016/j.bbabio.2014.01.017 (Available online 4 February 2014)

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