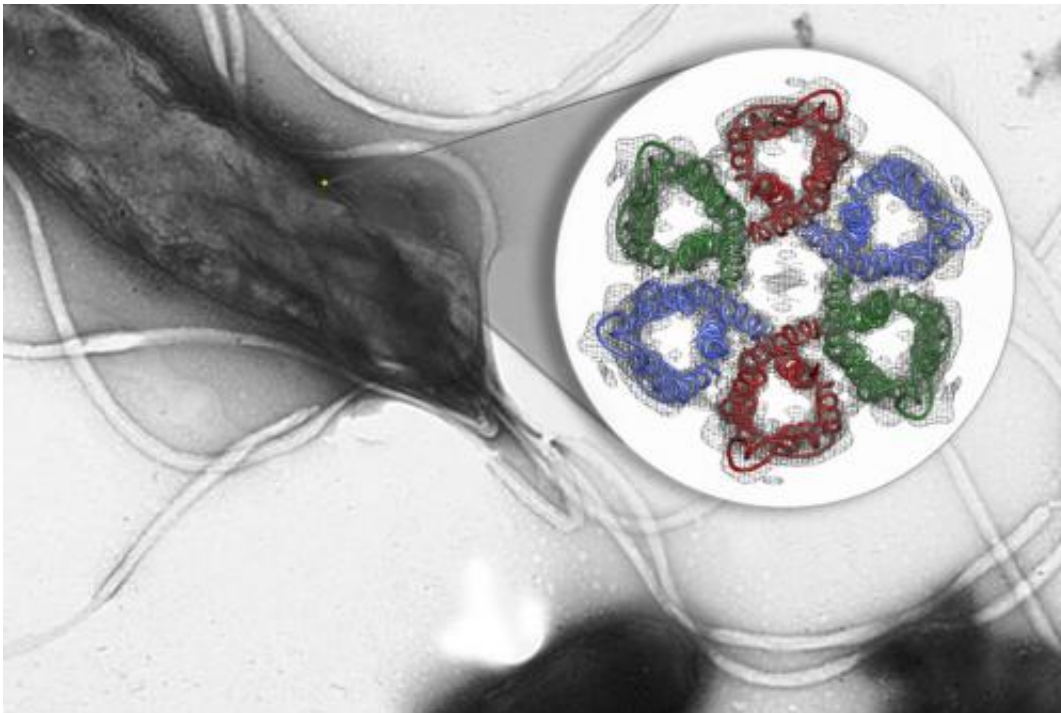


X-rays pinpoint drug target for *Helicobacter pylori*

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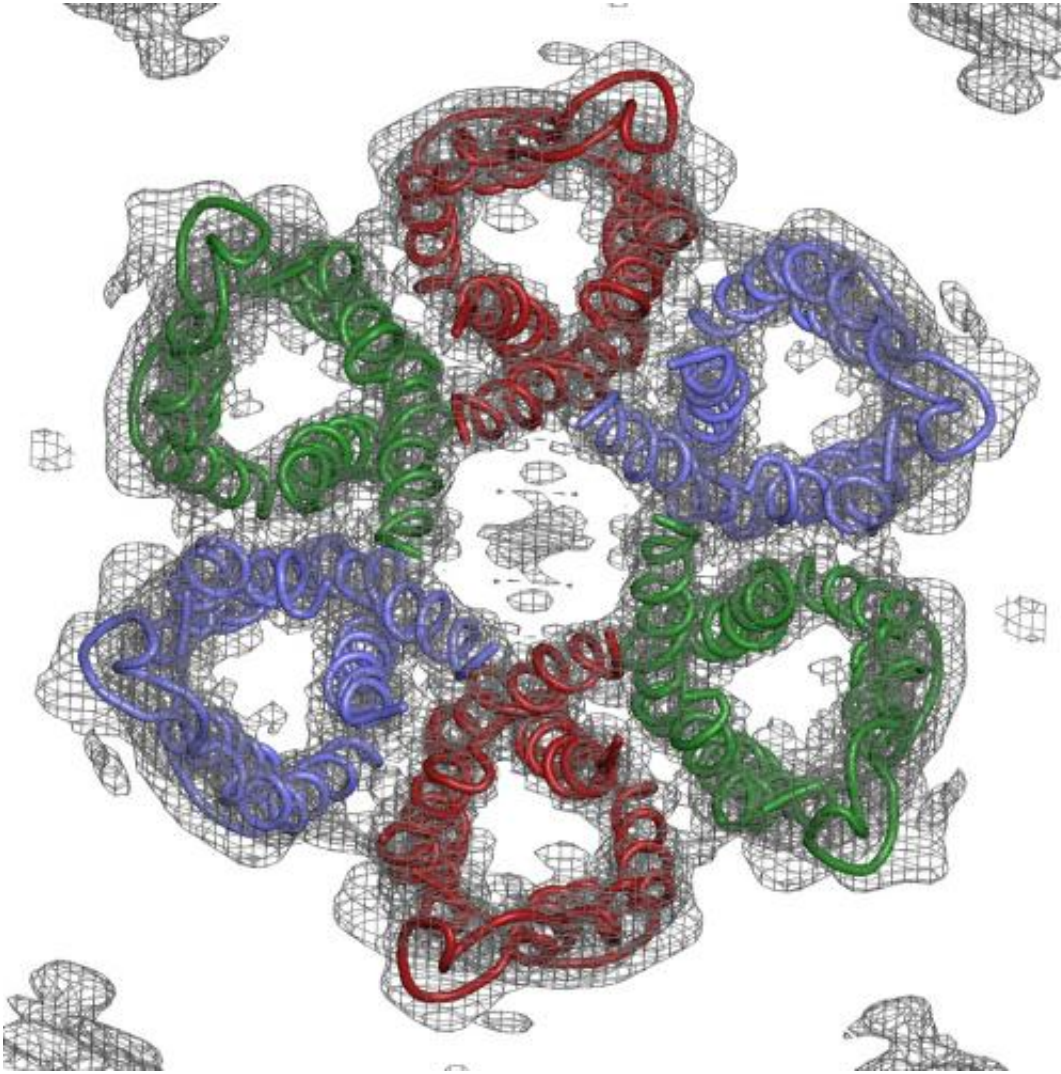
This diagram shows the first-ever glimpse of the six-molecule ring of urea channels embedded in the membrane of a type of bacteria called *Helicobacter pylori*. Understanding the structure of the urea channels will allow researchers to design future drugs to target this bacteria, which harm hundreds of millions of people worldwide. Credit: Hartmut Luecke / UC Irvine

(Phys.org)—Experiments at the U.S. Department of Energy's (DOE) SLAC National Accelerator Laboratory have revealed a potential new way to attack common stomach bacteria that cause ulcers and

significantly increase the odds of [developing](#) stomach cancer.

The breakthrough, made using powerful X-rays from SLAC's Stanford Synchrotron Radiation Lightsource ([SSRL](#)), was the culmination of five years of research into the bacterium [Helicobacter pylori](#), which is so tough it can live in strong stomach acid. At least half the world's population carries *H. pylori* and hundreds of millions suffer health problems as a result; current treatments require a complicated regimen of stomach-acid inhibitors and antibiotics.

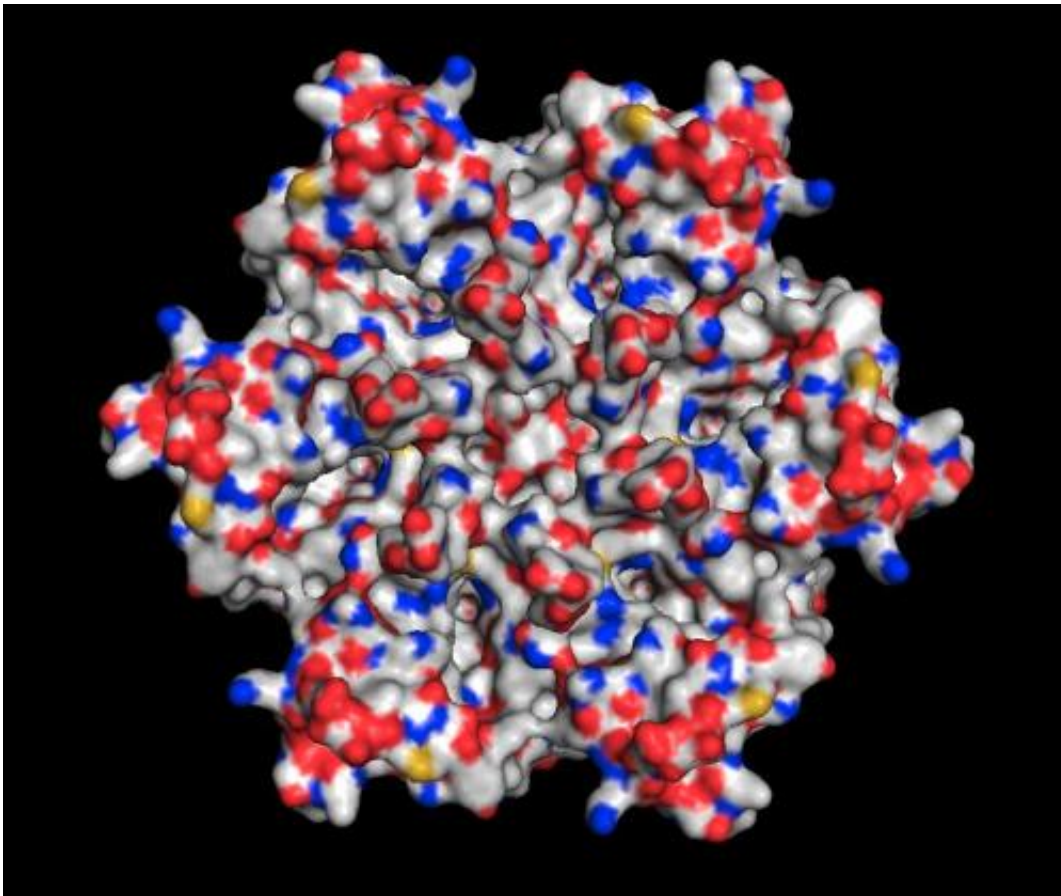
"We were looking for a means to disrupt *H. pylori*'s own mechanism for protecting itself against stomach acid," said Hartmut "Hudel" Luecke, a researcher at the University of California, Irvine, and principal investigator on the paper, published online Dec. 9 in *Nature*. With this study, he said, "We have deciphered the three-dimensional molecular structure of a very promising drug target."



An isolated look at the structure of the six-molecule ring of urea channels embedded in the membrane of *Helicobacter pylori*. Urea passes through the center of each of the six channel molecules. The center of the ring is filled with a lipid bilayer plug. Credit: Hartmut Luecke / UC Irvine

Luecke and his team zeroed in on tiny channels that *H. pylori* uses to allow in urea from gastric juice in the stomach; it then breaks this compound into ammonia, which neutralizes [stomach acid](#). Blocking the channels would disable this protective system, leading to a new treatment for people with the infection.

Solving the structure of the protein to find the specific area to target wasn't easy. The channels are formed by the protein embedded in the bacterium's cell membrane, and membrane proteins are notoriously difficult to crystallize, which is a prerequisite for using [protein crystallography](#), the main technique for determining protein structures. This technique bounces X-rays off of the electrons in the crystallized protein to generate the experimental data used to build a 3-D map showing how the protein's atoms are arranged.



A 3-D view of a newly understood molecular structure in the stomach bacterium *Helicobacter pylori*, built with data derived from protein crystallography experiments at the Stanford Synchrotron Radiation Lightsource (SSRL). Credit: Hartmut Luecke / UC Irvine

The challenge with [membrane proteins](#) is that they are especially hard to grow good quality crystals of, and for this experiment, said Luecke, "We needed to grow and screen thousands of crystals."

"We collected over 100 separate data sets and tried numerous structural determination techniques," said Mike Soltis, head of SSRL's Structural Molecular Biology division, who worked with Luecke and his team to create the 3-D map of the atomic structure. The final data set was measured at SSRL's highest brightness beam line (12-2), which produced the critical data that met the challenge.

"This is the hardest structure I've ever deciphered, and I've been doing this since 1984," Luecke said. "You have to try all kinds of tricks, and these crystals fought us every step of the way. But now that we have the structure, we've reached the exciting part—the prospect of creating specific, safe and effective ways to target this pathogen and wipe it out."

More information: The research paper is available online from *Nature*: www.nature.com/nature/journal/...ull/nature11684.html

Provided by SLAC National Accelerator Laboratory

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