

We need to talk: How cells communicate to activate notch

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During formation of multi-cellular organisms, cells need to talk to each other to make critical decisions as to what kind of cell to become, as well as when and where to become that cell type. The Notch signaling system allows cells to directly talk to each other to program almost every cell type in the body.

Now, researchers from UCLA's Jonsson Comprehensive Cancer Center have shown for the first time that the mechanical force produced by cellcell interactions is critical for programming by the Notch signaling system.

Prior to these findings, it was speculated that interacting cells undergoing Notch signaling pull on each other to unlock and activate Notch. Now, the UCLA scientists detail how the process unfolds, a discovery that could result in potential new targets for therapeutics, said Gerry Weinmaster, a professor of <u>biological chemistry</u> and a researcher with UCLA's Jonsson Cancer Center.

The findings are outlined in two companion papers that appear May 31, 2012 in the early online edition of the peer-reviewed journal, *Developmental Cell*.

"Our findings used optical tweezers as a novel tool to detect and measure mechanical force produced by cells when bound to Notch," said Weinmaster, co-senior author of the studies who has been studying Notch signaling for more than 20 years. "Together with biochemical and



cell biological analyses, our findings provide compelling evidence that pulling on Notch opens a network to deliver information that instructs specific <u>cellular responses</u>."

In its normal state, Notch is folded in an inactive form that protects its proteolytic cleavage, or cutting site, that once exposed, activates signaling in cells. Weinmaster likened the unfolding and activation of Notch by mechanical force to the force required to pull a pin from a grenade to produce an explosion. One of the interacting cells, known as the ligand cell, uses mechanical force to pull the "pin" from the Notch grenade present on the other interacting cell, and this results in an explosion of sorts that sets off cellular programming by Notch.

Weinmaster and her team wanted to obtain evidence that ligand cells actually produce pulling force following interactions with Notch cells and contacted Elliot Botvinick, an assistant professor in biomedical engineering at the University of California, Irvine, who is an expert in <u>optical tweezers</u>, a scientific instrument that uses a highly focused laser beam to detect and measure mechanical forces.

"To detect the pulling force, I recommended that we replace the Notch cell with a Notch bead that could be laser trapped and held just-incontact with the ligand cell," said Botvinick, co-senior author of the studies. "If the cell produced mechanical force, it would displace the bead from the trap, allowing the exact magnitude of the pulling force to be measured."

Botvinick determined that ligand cells could physically pull a Notchbead from the center of the trap.

The studies also shed light on a new role for endocytosis to activate signaling in <u>cells</u> that is unique to Notch, said Laurence Meloty-Kapella, a post-doctoral researcher and first author of one of the studies.



"Our studies provide compelling evidence that ligand endocytosis produces force to pull on Notch and identifies a new role for endocytosis in activation of a <u>signaling system</u>," Meloty-Kapella said.

Going forward, Weinmaster and her team will use cutting-edge biophysical approaches to clearly understand how pulling force is used to activate Notch both in human development and disease. This research is essential for the design of potential therapeutics for numerous Notchrelated diseases, in particular cancer.

"We conclude that the primary role of <u>ligand</u> endocytosis is to generate mechanical force to activate <u>Notch signaling</u>. Future studies to quantify the force required to dissociate Notch, as well as directly demonstrate <u>mechanical force</u> applied to Notch activates signaling, will extend our findings and further test the pulling-force model," the study states.

Provided by University of California, Los Angeles

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