

Chemists develop faster, more efficient protein labeling

February 5 2012

North Carolina State University researchers have created specially engineered mammalian cells to provide a new "chemical handle" which will enable researchers to label proteins of interest more efficiently, without disrupting the normal function of the proteins themselves or the cells in which they are found.

Protein labeling is used by researchers in a variety of fields to help them understand how these important [molecules](#) affect the normal functioning of cells. Currently, proteins are labeled for study simply by fusing them to other [fluorescent proteins](#), which allows researchers to use [microscopy](#) to track their movements through a cell. This approach has several drawbacks, however, not least being that the fluorescent proteins are often large enough to affect the function of the [protein](#) of interest.

Dr. Alex Deiters, associate professor of [chemistry](#), along with colleague Dr. Jason Chin of the Laboratory of [Molecular Biology](#) at the Medical Research Council in Cambridge, U.K., have developed a way to attach a fluorophore – a fluorescent molecule about 20 times smaller than the fluorescent proteins currently in use – to a protein that is expressed in a mammalian cell.

Deiters and Chin developed a special 21st amino acid that they added to cells that were specially engineered to incorporate this amino acid into the protein they wanted to study (there are normally only 20 amino acids). This 21st amino acid has a "chemical handle" that only reacts with a specifically designed fluorophore, but not any cellular

components. According to Deiters, "The reaction between the modified protein and the fluorophore is extremely fast, high yielding, and generates a stable link between both reaction partners. This novel methodology enables future cell biological studies that were previously not possible."

The research appears in the Feb. 5 issue of *Nature Chemistry*.

"We found that our approach gave us a higher yield of labeled proteins and that the binding reaction was 50 times faster than with current methods," Deiters says. "Additionally, it took less reagent to complete the reaction, so overall we have a faster, more efficient method for protein labeling, and less chance of interfering with the normal function of the proteins and cells being studied."

The research was funded by the National Institutes of Health and the National Science Foundation. The Department of Chemistry is part of NC State's College of Physical and Mathematical Sciences.

More information: "Genetically encoded norbornene directs site-specific cellular protein labelling via a rapid bioorthogonal reaction", Feb. 5, 2012 in *Nature Chemistry*.

Provided by North Carolina State University

Citation: Chemists develop faster, more efficient protein labeling (2012, February 5) retrieved 1 May 2024 from <https://phys.org/news/2012-02-chemists-faster-efficient-protein.html>

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