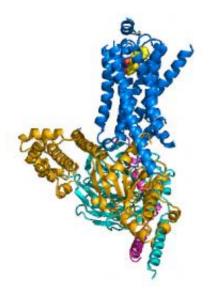


## How receptors talk to G proteins

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The structure of the  $\beta$ 2AR-Gs complex: The  $\beta$ 2AR (blue) occupied by a high affinity agonist (yellow spheres) forms a complex with Gs, the stimulatory G protein for adenylyl cyclase. Gs is composed of three proteins: G $\alpha$  (gold), G $\beta$  (cyan) and G $\gamma$  (purple).

(PhysOrg.com) -- The mechanism by which cells respond to stimuli and trigger hormonal responses, as well as the senses of sight, smell, and taste, has for the first time been brought into focus with the help of highbrightness x-rays provided by the U.S. Department of Energy Office of Science's Advanced Photon Source (APS) at Argonne National Laboratory. This breakthrough will pave the way to new research avenues in drug discovery, cell signaling, and cellular regulation.



Receiving a signal, interpreting it, and responding correctly: these are three activities that <u>cells</u> must be good at in order to respond to stimuli. For responding to hormones, and for the senses of sight, smell, and taste, the activated receptors are coupled with G proteins. These G proteincoupled receptors (GPCRs) are involved in a complex set of steps in which an extracellular hormone or neurotransmitter binds to and activates a GPCR in the cell membrane. The activated receptor passes on the signal to a G-protein inside the cell, triggering reactions that ultimately create a cellular response to the stimulus.

The intricacy of this set of reactions has been appreciated—and studied—for some time. What was lacking, however, was a finely-detailed <u>crystal structure</u> of the GPCR as it actually signals across the membrane. The importance of acquiring this knowledge and knowing a lot more about how GPCRs function is apparent when we consider that the human genome contains over 800 GPCR genes.

The structure of protein receptors that are involved in cellular responses to <u>stimuli</u> is now available thanks to experiments carried out at the National Institute of General Medical Sciences and National Cancer Institute Collaborative Access Team (GM/CA-CAT) beamline 23-ID-B at the APS, and the process of cell signaling has come into crystal clear focus. This crystal structure is the first high-resolution look at transmembrane signaling by a GPCR and adds critical insight about signal transduction across the plasma membrane.

The general model for GPCR signaling is that the G-protein is activated by a receptor that has received a stimulus from outside the cell. The research team, comprising members from Stanford University, the University of Copenhagen, the University of Michigan, the University of Wisconsin, Vrije Universiteit Brussel, and Trinity College studied a specific model system for GPCR signaling that has long been used by biochemists and about which much was already known. In this system, a



 $\beta$ 2 adrenergic receptor ( $\beta$ 2AR) is bound by the outside stimulus, known as an agonist, and then activates Gs, the stimulatory G protein for adenylyl cyclase.

The researchers were able to capture the  $\beta$ 2AR bound to the Gs protein before the latter went on to the next step in the dance, which would be binding a nucleotide. One technical challenge, and perhaps the reason that the crystal structure had been so elusive to date, was to create a stable  $\beta$ 2AR-Gs complex in detergent solution. Once this problem was solved, the team could proceed to unveiling the crystal structure of this active-state  $\beta$ 2AR-Gs complex (see the accompanying figure).

The new data allowed the research team to construct the early stages of GPCR signaling. Using their own observations, and other data collected on this model system, a mechanism for the initial stages of complex formation came into being. The data provided a pinpoint determination of exactly which changes in the molecules permitted that binding stage to carry on to completion.

Shape changes in the Gs, particularly in the nucleotide-binding pocket, prepared the protein for the next step. In the  $\beta$ 2AR, the researchers were able to identify major changes in two areas of the molecule that fit well with understanding of how the  $\beta$ 2AR interacts with molecules outside the cell and how the two molecules— $\beta$ 2AR and Gs—interact each other.

Article co-author Brian Kobilka, of Stanford University, recalled how the project reached this important milestone: "For the past five years, Roger Sunahara and I have been working together to understand how receptors and G proteins interact with each other. This was the next logical step after getting the first inactive-state structures of the  $\beta$ 2AR alone in 2007.

"The receptor and <u>G protein</u> purification procedures were well



established at the time we started the project, but most of the other methods were developed during the project. The minibeam and the rastering‡ capabilities of the GM/CA-CAT beamline at the APS were essential for collecting the diffraction data on these crystals."

Future research, Kobilka said, involves "how the complex forms and dissociates after activation."

**More information:** Søren G. F. et al. "Crystal structure of the b2 adrenergic receptor–Gs protein complex," *Nature*, advance online publication, 19 July 2011. <u>DOI: 10.1038/nature10361</u>

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