

## New research aims to shut down viral assembly line

January 11 2011, by Richard Harth



This is a depiction of the SARS coronavirus by Luke Jerram. Credit: Courtesy of Heller Gallery, New York

Under the electron microscope, a coronavirus may resemble a spiny sea urchin or appear crownlike, (the shape from which this family of pathogens takes its name). Previously recognized as the second leading cause of the common cold in humans and for economically important diseases in many domesticated animals, a new disease form abruptly emerged as a major public health concern in 2002, when the SARS coronavirus (CoV) surfaced in Asia.

The rapid spread of the virus caused significant social and economic



disruption worldwide, infecting over 8000 people with Sudden Acute Respiratory Syndrome or SARS and killing about 10 percent of them. While SARS-CoV was brought under control through decisive action by health officials, the sudden scourge underlined the threat posed by coronaviruses and spurred new research into the inner workings of these infectious agents.

Brenda Hogue and her colleagues at the Biodesign Institute at Arizona State University are studying the intricate formation of these viruses—a process known as viral assembly. The research may offer fresh insight, leading to a new generation of antiviral agents that can disrupt the ability of coronaviruses like SARS to assemble viable infectious particles. Such strategies may prove applicable against other classes of virus as well.

The group's work recently appeared in the Journal of Virology.

Viruses, Hogue stresses, differ fundamentally from other common microscopic <u>pathogens</u> like bacteria, in that viruses are structurally primitive, lacking the means to independently replicate. Viruses are composed of genetic information (DNA or RNA), encased by proteins. They exist in a shadowy region between living and non-living entities.

In order for a virus to replicate, it must commandeer machinery of a host cell it has infected. Nevertheless, viruses have evolved to be highly adept at this sort of replication-by-proxy, and can infect virtually all types of organisms, from animals and plants to bacteria and even Archaea. Viruses—of which millions of forms are known to exist—are far and away the most numerous (and successful) parasitic invaders on earth.

"Coronaviruses are a very large family of RNA viruses," Hogue says. "They infect humans and a broad range of animals." While the symptoms produced by coronavirus infection in humans tend to be respiratory, in animals, such viruses can cause a range of severe



problems, from neurological ailments to immunosuppressive effects. Various coronaviruses are responsible for common colds in humans, though the combined upper and lower respiratory symptoms and gastrointestinal complications seen in SARS patients are unusual.

In the study reported in *Journal of Virology*, Hogue and her team closely examined one of the major proteins found in the coronavirus that is crucial to the pathogen's process of assembly. Known as the M, it is one of four proteins, in addition to S, N and E, required to produce a fully assembled viral particle, capable of infecting a host.

The membrane (M) protein makes up the bulk of the outer shell or envelope of the virus, forming a lattice that surrounds and shields the viral genome. The spike protein (S)—named for its spike-like or crownlike appearance under electron microscopy, is critical for allowing the coronavirus to attach to the host cell's receptors, prior to viral entry into the cell. The nucleocapsid (N) protein encapsidates the genomic RNA. The envelope or E protein is the least plentiful protein known to play a central role in virus assembly, though its presence is very important. In addition to assisting viral assembly, the E protein also appears to be involved in shuttling the newly assembled virions out of the cell, enabling these particles to escape and infect other host cells in the exponential process of viral infection.

The group wanted to determine the requirements for the M protein to function during assembly of the viral envelope. To establish this, coronaviruses were genetically manipulated to form mutant versions, exhibiting varying degrees of viability. Much of this manipulation focused on domains within the viral genome coding for a distinct structural and/or functional domain of the M protein. Conserved domains, as they are known, contain genetic sequence patterns or motifs that tend to recur across a number of different viruses or within a particular virus type, like catch phrases recurring in different books.



These conserved domains are generally involved in functions essential to viral formation, survival or replication, making them an attractive target for therapeutic efforts designed to short-circuit viral assembly.

Viral pathogens like the SARS <u>coronavirus</u>, (along with hepatitis C and influenza), use RNA rather than DNA as their genetic material. In general, such RNA viruses mutate more rapidly than DNA viruses, posing particular challenges to virologists hoping to combat them. They can also acquire alterations that allow them to hop from one species to another. Something like this now appears to be at the root of the SARS outbreak.

"We think that the reservoir for this virus is bats, because a large number of SARS-like viruses have been isolated from bat populations around the world," Hogue says. SARS-CoV was subsequently able to infect a secondary animal host, now believed to be the civet cat—a mongooselike creature found in Asia and sometimes used as a food source, particularly in China, where the SARS outbreak originated. Contact with infected civets in the open markets may have caused the initial human cases of SARS, which then rapidly spread— human to human—from the Guangdong provinces in China to 37 countries.

Viruses that act as respiratory pathogens, including SARS, are highly transmissible from person to person through contact with respiratory droplets that become aerosolized from coughs or sneezes. Virions may also persist on surfaces that come in contact with an infected individual. Following transmission, virions initiate the infectious process of host cells, which transpires in several important phases.

First, viral proteins located on the virus' outer capsid bind to particular receptors on the host cell's surface. Next, virions enter the cell, either by fusing their membranes with those of the host cell or through the process of endocytosis, in which the host cell takes in the virion in a membrane-



bound vesicle. The virion is now in a position to begin the replication cycle, releasing its genetic material into the cell. The viral genome encodes genes that when expressed, yield the protein components necessary to assemble new virus particles.

Hogue stresses the importance of in silico analysis, in which large libraries of proteins can be screened through analysis, in order to identify conserved and non-conserved protein regions, thus greatly accelerating the pace of discovery. "The more we learn about these particular regions of proteins that are critically important for the assembly process," she says, "the likelier it is we can design molecules that will be able to interfere with this process."

While some conserved domain alterations in the M protein proved lethal to coronaviruses, others undermined viral assembly without shutting it down completely, often causing compensatory efforts on the part of the virus, (known as second site changes) which may offer insights into the virus' adaptive capabilities. In the coming year, Hogue plans to examine the non-lethal changes introduced, studying these mutant viruses under high-resolution cryo-EM, to determine how alterations of specific domains affect overall coronaviral structure.

Additionally, Hogue's group is closely examining the under-represented envelope or E protein. "One reason we are excited about this is that a number of enveloped viruses, including hepatitis C, influenza and others, that are of real medical significance, have small ion channel proteins," Hogue says. "If we can develop ways to target and obliterate the ion channel activities of these proteins, we may be able to disable these viruses and prevent or reduce infections."

Provided by Arizona State University



Citation: New research aims to shut down viral assembly line (2011, January 11) retrieved 2 May 2024 from <u>https://phys.org/news/2011-01-aims-viral-line.html</u>

This document is subject to copyright. Apart from any fair dealing for the purpose of private study or research, no part may be reproduced without the written permission. The content is provided for information purposes only.