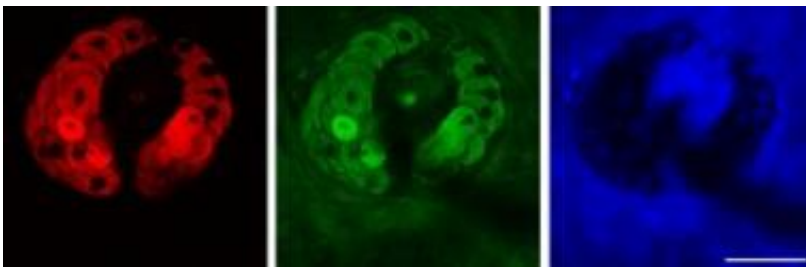


New microscopy tracks molecules in live tissue at video rate

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Based on characteristic vibrational spectra of endogenous molecules, many important structural components of tissue such as lipids (CH₂-vibration, red), proteins (CH₃-vibration, green), and water (OH-vibration, blue) can be imaged. The images show a sebaceous gland wrapping around a hair in the viable epidermis of mouse skin. (left) CH₂-image in red shows the lipid-rich gland cells with sub-cellular resolution. Nuclei appear as dark circles due to the lack of lipids. (center) CH₃-image in green shows residual lipid signal but also new protein-rich structures such as a hair in the center of the image, collagen fibers surrounding the gland and red blood cells in the top left. It can be seen that hair (green solid center) is surrounded by lipid-rich sebum (red circle). (right) OH-image due to water in blue shows inverse image contrast from the sebaceous gland because the lipid rich cells exclude water. Recent developments have allowed the acquisition of SRS images in reflection mode and at video-rate imaging speed in living mice and humans. Scale: 20 μ m. Credit: Image courtesy of Dr. Brian Saar and Christian Freudiger.

A novel type of biomedical imaging, made possible by new advances in microscopy from scientists at Harvard University, is so fast and sensitive

it can capture "video" of blood cells squeezing through capillaries.

Researchers led by Harvard's Brian G. Saar, Christian W. Freudiger, and X. Sunney Xie describe the work this week in the journal *Science*.

The new technique, based on stimulated Raman scattering (SRS), makes a complementary partner to MRI, widely used to capture static images of organs, tumors, and other large structures. For the first time, SRS microscopy makes possible label-free chemical movies, with streaming footage at the subcellular level, catching video of proteins, lipids, and water within cells.

"When we started this project 11 years ago, we never imagined we'd have an amazing result like this," says Xie, professor of chemistry and [chemical biology](#) at Harvard. "It took MRI more than 30 years to reach patients, but we're already looking forward with great anticipation to applications of SRS microscopy in hospitals. It's now clear that stimulated Raman scattering will play an important role in the future of biological imaging and medical diagnostics."

Xie says SRS microscopy could aid, and speed, surgery to remove tumors and other [lesions](#). Surgeons must now send excised samples for histological analysis -- a process that takes about 20 minutes -- while a patient waits on the operating table. SRS microscopy provides equivalent insights through real-time scanning.

Xie's team has already used SRS microscopy to track migration of medications in skin, shedding new light on the absorption of topical drugs. In conjunction with endoscopy, the technique can also view three-dimensional sections of tissue, layer by layer.



(left) SRS images of the viable epidermis at the CH₃-vibration mainly highlight proteins. A capillary with individual red blood cells (arrow) is visible. The cells are imaged without motion blur due to video rate acquisition speed. (right) An x-t plot acquired by line-scanning across a capillary at the position of the arrow. Individual red blood cells are captured on the fly. Scale: 25 μ m. Credit: Image courtesy of Dr. Brian Saar and Christian Freudiger.

"Previous SRS microscopy captured only about one image per minute, far too slow for use in live animals or humans," Xie says. "We were able to speed the collection of data by more than three orders of magnitude, attaining video-rate imaging."

Because SRS microscopy works by detecting the intrinsic vibrations in chemical bonds between atoms, it doesn't require intrusive fluorescent labeling. An optical technique, it compliments MRI, whose depth of penetration is better suited to imaging organs and other large objects deep within the body.

The current work greatly improves detection of signals -- backscattered by tissues in the body -- by rearranging photodetectors to surround a small aperture through which a beam of light is directed at the tissue being examined. Using this approach, the scientists were able to collect and analyze almost 30 percent of the laser light directed at a biological sample, a more than 30-fold increase over previous SRS microscopy.

Scientists currently use a variety of techniques to view biomolecules, but most have significant limitations that are sidestepped by SRS

microscopy. Labeling with green fluorescent protein (GFP) provides sharp images, but the bulky protein can perturb delicate biological pathways, overwhelming smaller biomolecules. Conventional infrared (IR) microscopy has low spatial resolution and requires desiccated samples, while spontaneous Raman microscopy requires high laser power and long integration times, limiting use in live specimens. Coherent anti-Stokes Raman scattering (CARS) [microscopy](#), pioneered by Xie's own group, lacks the contrast to image most molecules beyond lipids.

Provided by Harvard University

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