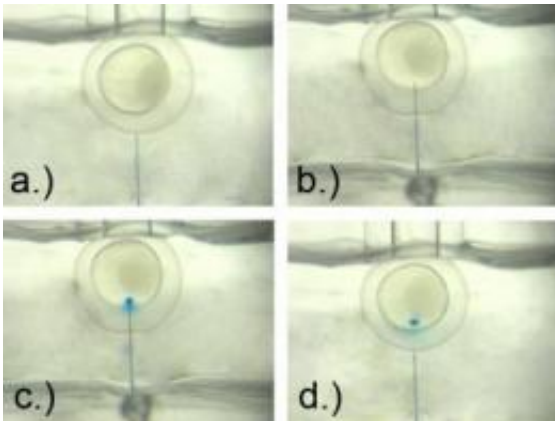


Tiny injector to speed development of new, safer, cheaper drugs

November 4 2009



(a) Zebrafish embryo immobilized by suction capillary. (b) Needle inserted into yolk sack. (c) Electroosmotic pumping of methylene blue solution into the embryo by the application of 25 V for 10 s. (d) Needle retracted from the embryo. Credit: McMaster Engineering

It's no bigger than a stamp packet but it has the potential to allow rapid development of a new generation of drugs and genetic engineering organisms, and to better control in-vitro fertilization.

Engineering researchers at McMaster University have fabricated a palm-sized, automated, micro-injector that can insert proteins, DNA and other biomolecules into individual cells at volumes exponentially higher than current procedures, and at a fraction of the cost. This will allow scientists to vastly increase preclinical trials for drug development and

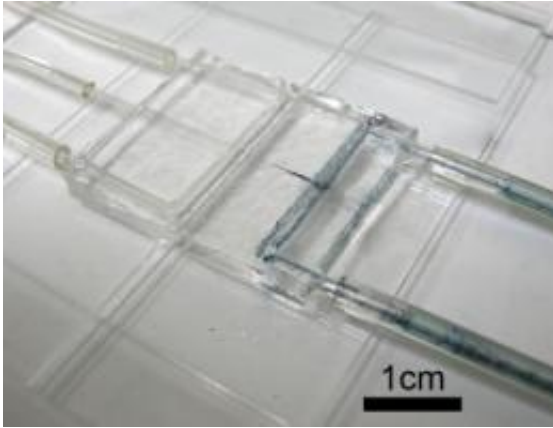
[genetic engineering](#), and provide greater control of the process.

In a paper published in the current issue of *Lab on a Chip* journal, researchers describe the construction and operation of a microfluidic micro-injector, which achieved an almost 80 per cent success rate in injecting [Zebrafish](#) embryos.

"This device is to [drug discovery](#) what the assembly line was to the automobile or the silicon chip to information technology," explains Ravi Selvaganapathy, assistant professor of mechanical engineering at McMaster and lead author of the research. "It turns what was a complex, resource-intensive process available to a few into an automated, predictable, reliable, and low-cost system accessible to almost anyone."

The micro-injector has a cell-wide channel cast on a silicon chip that guides cells and [embryos](#) to the injection site. A similar channel guides the injection reagent to a needle as thin as 10 micrometers (one-tenth the diameter of a human hair). The researchers have developed a buckling method to drive the needle through a cell's pliable [outer membrane](#) accurately and to the proper depth. The injection dosage is controlled electrically, as is monitoring of the needle's position. The researchers have also developed methods to sharpen the needle, ensuring minimal injection damage or interference to the cell.

Notably absent is the need for a [microscope](#) or optical magnification to conduct the process, which is required for manual injection and to monitor transfection methods. The microfluidic device also allows easy integration of post-processing operations including cell sorting and the testing of cell viability on the same chip.



This photo shows a microinjection device loaded into a fixture which immobilizes the left substrate and ensures planar and linear motion of the movable substrate. Credit: McMaster Engineering

"Almost every researcher would be able to have this device at their disposal in their own labs," said Selvaganapathy. "The micro-injectors can easily be run in parallel and allow for scientists to test far greater combinations of materials in a much shorter time than current processes. It also makes it more feasible to pursue drug discovery for many so-called neglected diseases."

The micro-injector also holds great promise for [in-vitro fertilization](#) as it provides far greater accuracy and control than current manual injections procedures, which have high rates of failure, require trained expertise and can be time intensive.

The micro-injector has achieved numerous firsts for cell transfection procedures:

- Buckling based actuation of injection needle providing low cost but precise actuation with uniform injection depth and consistent

alignment;

- Injection format that could allow needles as small as 100 nanometres, half the size of current injectors, virtually eliminating cell damage and interference with cell functions;
- Electro-osmotic injection which provides electrical control of reagent injected into cell for accurate and uniform dosage;
- Elimination of expensive optical magnification needed for manual injection or to monitor quality control.

Source: McMaster University ([news](#) : [web](#))

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