

Corn researchers discover novel gene shut-off mechanisms

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Pamela Green, Crawford H. Greenewalt Chair and professor of plant and soil sciences and marine bioscience, at the University of Delaware. Credit: University of Delaware/Kathy F. Atkinson

University of Delaware scientists, in collaboration with researchers from the University of Arizona and South Dakota State University, have identified unusual differences in the natural mechanisms that turn off, or "silence," genes in corn.

The discovery, which was made by comparing the impact of inactivating a gene that occurs in both corn and in the much-studied laboratory plant *Arabidopsis*, provides new insight into how one of the world's most important crops protects itself from mutation-causing mobile DNA elements and viruses.

The research was led by Blake Meyers, associate professor of plant and soil sciences, and Pamela Green, Crawford H. Greenewalt Chair and professor of plant and soil sciences and marine bioscience, and their laboratory groups at the Delaware Biotechnology Institute, a major center for biotechnology and life sciences research at the University of Delaware.

Collaborating with the University of Delaware team were Vicki Chandler, the Carl E. and Patricia Weiler Endowed Chair for Excellence in Agriculture and Life Sciences Regents' Professor at the University of Arizona, and Yang Yen, a professor at South Dakota State University.

The results were published in the *Proceedings of the National Academy of Sciences* of the United States of America.

Studies of *Arabidopsis thaliana*, a small flowering plant of the mustard family that is easy to grow in the lab, have provided a lot of what scientists know about gene silencing in plants.

An important key to the process is short sequences of ribonucleic acids known as "small RNAs" which act like biochemical switches that shut off genes, thus playing a fundamental role in plant development. Understanding how small RNAs work is a continuing quest for geneticists seeking to breed plants with improved crop yields, disease resistance and other characteristics.

Previously, the Meyers and Green labs had studied *Arabidopsis* plants with nonfunctional versions of a gene known as RNA-dependent RNA polymerase 2 (RDR2). Without an active copy of this gene, the plants were unable to produce a major class of small RNAs, which act to stabilize and protect genes on the chromosomes.

In that prior work, Meyers and Green took advantage of the

nonfunctional gene to study microRNAs, an interesting type of small RNA that is usually "masked" by the major class of small RNAs produced by RDR2.

Independently of the UD groups, Chandler and her team at the University of Arizona had identified from corn an orthologous gene--a gene that has the same function in different organisms. In corn, this gene, which the Chandler lab found, is called the mediator of paramutation (MOP1). Its equivalent in Arabidopsis is the RDR2 gene.

Because the RDR2 and MOP1 genes should both produce the "protective" set of small RNAs, the research groups decided to collaborate to see if the small RNAs in corn behave the same way they do in Arabidopsis. The hypothesis was that the result would be the same in the two plant species, and the lab groups could use the MOP1 corn plants to focus their studies on the harder-to-examine microRNAs, as they had done previously in Arabidopsis.

"Yet we found something that had not been observed before in this plant--an odd class of small RNAs," Meyers said. "I think it's pretty neat to work in a more complex system like corn and see things that Arabidopsis hadn't shown us," he noted.

Using a technique known as sequencing by synthesis (SBS), provided by Illumina in Hayward, Calif., coupled with state-of-the-art bioinformatics in Meyers' lab, the research team found that the MOP1 and RDR2 genes are not fully equivalent based on an assessment of small RNA complexity.

The researchers found that there are lots more RNAs of an unusual class known as "small interfering RNAs" in corn than there are in Arabidopsis.

"This class of RNAs mainly functions to repress repetitive sequences, including mobile DNA elements called transposons," Meyers said.

"Thus, small interfering RNAs act to protect the genome," he noted.

"Corn contains an extra layer of protective small RNAs that had not been observed in Arabidopsis, so there must be additional genes other than MOP1 that produce this," Meyers said.

The scientific community is sequencing the corn genome now, Meyers said. Once the genome is available, the work of matching up small RNAs to specific traits in corn will be much easier, he noted.

"This research is helping us to better understand the biology of corn--one of the most important plants in the world--and gives us new avenues for exploring a novel class of small RNAs," Meyers said.

Source: University of Delaware

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