

At the molecular level, the predator is the prey

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An evolutionary arms race between predatory garter snakes and their newt quarry is turning out to be something of an illusion. At the molecular level, another battle rages. And in this second, miniature realm, it's the newt who's the aggressor.

Biologists at Indiana University Bloomington, Utah State University and the University of Utah present evidence in this week's *Nature* that a toxin produced by the rough skinned newt, *Taricha granulosa*, has forced several evolutionary changes in the garter snake *Thamnophis sirtalis* or, more specifically, in the snake nerve cell protein that endures the toxin's attacks.

Image: Some garter snakes (Thamnophis sirtalis) have evolved the ability

*to eat super-toxic newts (*Taricha granulosa*) in the Pacific Northwest*
Photo by: Edmund Brodie III

Embedded in the surface of garter snake nerve cells is $\text{tsNa(V)}1.4$, a tube-shaped protein that allows sodium ions to flow into the cell. When nerve cells' ability to move sodium in and out is hampered, paralysis and death can result. Tetrodotoxin (TTX), a powerful paralytic poison concentrated in the newts' skin, can bind to garter snake nerve cell channels and prevent sodium ions from flowing freely.

"These channels are absolutely fundamental to every aspect of nerve and muscle function and are highly specific gateways for sodium ions," said IUB biologist Edmund Brodie III, one of the paper's coauthors. "If the channels change too much or in the wrong way, they can't perform their basic, everyday functions. Sodium channel genes in different vertebrates are virtually identical to each other, but not in these snakes. We're finding a molecular arms race is driving rapid and repeated changes in the gene within this group of beasts."

For TTX to bind successfully to the sodium channel, the toxin needs something to bind to. At this moment in the garter snake's evolutionary history, TTX infiltrates a hole on $\text{tsNa(V)}1.4$'s surface, binding to an aromatic amino acid and causing enough of a change in the sodium channel's shape to impair its function. Three of the four Pacific Northwest snake populations the scientists examined have evolved some degree of resistance to TTX by making this aromatic amino acid harder for TTX to grasp -- or by removing it altogether.

One-thirtieth of the TTX normally found in a *T. granulosa* newt is enough to kill the average human being. The only organisms on Earth that can eat *T. granulosa* newts and survive are some *T. sirtalis* garter snakes. TTX is a defensive compound found in some puffer fish, octopuses and primitive chordates. It is used in low concentrations to

treat morphine and heroine addicts. It's also the "zombie" drug used by Haitian voodoo ritualists.

Despite its action at the molecular level, the evolution of TTX in some organisms is viewed by ecologists as a defense mechanism. In the case of *T. granulosa* newts and *T. sirtalis* garter snakes, the interaction has gone far beyond that simple fangs-off arrangement, evolving into a lethal contest of toxification/detoxification one-upsmanship.

"One might think that this sort of change in the sodium channel would be too costly to the snakes," said Utah State University biologist Shana Geffeney, who conducted the gene expression experiments. "What will be interesting in the future is to understand if there is a balance between the costs of the changes in the channel pore structure on channel function and the benefits of changes in TTX binding."

The evolution of new traits often happens one of two ways, either by altering existing genes or by changing patterns and amounts of expression. The current Nature report shows that snakes' ability to detoxify TTX involves changes in the sodium channel gene.

"That is generally the story as it is developing," Brodie said. "Ecological arms races that go on between predator and prey are really driven at the molecular level. We have no evidence, nor reason to believe, that TTX is changing too, but rather that the toxin responds in quantity. Pour on more toxin, change the snake's sodium channel. Add more toxin, force further changes in the channel."

Source: Indiana University

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